

Genome Sequencing and Phylogenetic Analyses as a Basis for Molecular Subtyping of Male-Specific (FRNA) Coliphages

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Abstract

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(Under the direction of Mark D. Sobsey)

Monitoring programs for recreational waters utilize indicator bacteria concentrations as predictors of sewage-exposure related illness risks. However, most illnesses contracted through exposure to recreational waters may be of viral etiology. Identifying the fecal sources (non-human vs human) is also valuable information for risk management and source mitigation. Male-specific (FRNA) coliphages are proposed as sensitive enteric viral indicators for source-tracking fecal pollution in environmental waters. Classified as family *Leviviridae* of two genera, *Levivirus* and *Allolevivirus*, and four genogroups (I, II, III, IV) the genogroups provide information regarding animal or human fecal sources. In order to design an assay for molecular identification of specific genogroups, a genomic sequence database of sufficient size must be generated from several FRNA coliphages collected from diverse sources or locations. The complete genome of 21 FRNA strains was sequenced and compared with 11 strains available in GenBank. Sequences of 30 out of 32 FRNA coliphages demonstrated very similar conserved regions, Open Reading Frame positions, amino acid compositions and gene maps when compared to the FRNA reference strains. The sequence of two strains could be placed in a new subcluster of genogroup I and further analysis suggests that these viruses are natural recombinants. Among viruses within each

genogroup, nucleotide sequence similarities ranged from 75-99%, 83-93%, 69-95% and 74-95% for genogroups I, II, III and IV, respectively. Genogroup II lysis protein tree formed a unique branch that was not observed in the full-length nucleotide tree. Thus, both full-length nucleotide and individual protein sequences need to be evaluated when genotyping or phylogenetically clustering these FRNA coliphages. From conserved regions within each genogroup, four genogroup-specific primer sets were designed for a reverse-transcription polymerase chain reaction (RT-PCR) assay. The assay was then evaluated successfully on a panel of environmental FRNA strains demonstrating their usefulness to assess the sanitary quality of recreational waters and provide data identifying and subsequently eliminating the contamination source.

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I. Introduction

A link between waterborne transmission of disease and sewage was first observed in 1854 by the historical achievements of Dr. John Snow. The observations by Dr. Snow pioneered the sciences of epidemiology, preventative medicine and public health intervention. His observations of the cholera outbreak led to the removal of the Broad Street pump handle in London followed by a rapid, subsequent decline in cholera-related deaths ([Snow, 1855](#)).

Sewage disposal into marine and freshwater systems has occurred since the establishment of community populations. In the USA and most developed countries treated wastewaters are commonly discharged into aquatic environments, including both drinking-water resources and recreational waters. Fecal contamination introduced into waterways may potentially contain both human and/or zoonotic pathogens, including various enteric and respiratory viruses. A virus- infected individual may fecally shed viral particles for weeks at levels up to 10^5 to 10^{12} per gram of stool ([Bosch, 1998](#); [Griffin et al, 2003](#); [Gerba, 2000](#)), depending upon the viral species or type. The majority of the shed viruses are thought to enter the aquatic environment primarily from sewage discharge, but additional routes include solid waste applications ([Bosch, 1998](#)), and shedding during recreational bathing ([Gerba, 2000](#)). Although treating wastewater by conventional primary and secondary treatment and terminal disinfection reduces the majority of fecal pathogens, resistant bacteria, parasite cysts, oocysts and spores and some viruses may not be removed adequately. Besides

municipal wastewater discharges, inputs such as stormwater runoff, septic-tank seepage from on-site systems, agricultural runoff, urban runoff and other fecal waste sources can enter aquatic environments. Other naturally occurring, non-pathogenic and potentially pathogenic viruses are already present in aquatic environments, thus adding to the complexity of the viral ecology of aquatic systems.

To minimize adverse impacts and to protect the public health and aquatic environments, management systems based on fecal indicators of microbial origin were implemented as a “warning flag” and a means by which to estimate fecal contamination based on direct microbial measurements. The World Health Organization ([WHO](#); [Ashbolt et al, 2001](#)) defines microbial indicators of public health concern by one of three groups:

- “(1) process indicator - group of organisms that demonstrates the efficacy of a process, such as total heterotrophic bacteria or total coliforms for chlorine disinfection
- (2) fecal indicator - a group of organisms that indicates the presence of a fecal contamination, such as thermotolerant coliforms or *E. coli*. Hence, they only infer that pathogens may be present
- (3) index and model organisms - a group or species indicative of pathogen presence and behavior, respectively. For example, *E. coli* as an index for Salmonella; F-RNA coliphages as models of human-enteric viruses.”

Criteria for an ideal fecal indicator selection are as follows: (1) consistently present in feces at higher concentration than those of pathogens; (2) cannot/should not replicate outside the intestinal tract; (3) easily detected and quantified; (4) non-pathogenic; (5) at least as resistant as pathogens to disinfection treatments and environmental conditions/inactivation rates; (6) indicator concentration in water is quantitatively associated with potential risks to human populations, typically from enteric illness; and (7) applicable to all water types (marine, freshwater, estuarine).

Existing bacteriological culture methods for determining fecal contamination in water

provide quantitative estimates of *E. coli* and enterococci. These methods are neither real-time nor do they provide information regarding source. The best that current methods can do is indicate that possible fecal contamination occurred within the last 24 hours. To minimize risks to human health, resource managers and human health advisors need an early-warning indicator that will enable them to assess the sanitary condition of waters in real-time or at least shortly after the sample is collected for analysis. An additional limitation of the current indicators, enterococci and *E. coli*, is that they do not correlate with the presence and concentrations of all potential water-borne pathogens (Griffin et al., 2003). Most importantly, current EPA recreational water-quality criteria using two bacterial indicators have little or no correlation to the presence and concentration of human pathogenic viruses. To date, no viral indicator has been mandated for regulatory purposes in recreational waters in the USA.

Male-specific coliphages have been suggested as a viral indicator for: (1) fecal contamination (Osawa, 1981; Furuse, 1983), (2) enteric bacterial contamination (Gerba, 1987), (3) enteric viral contamination (Grabow, 2001; Leclerc et al, 2000) and (4) risks of gastro-intestinal illness from recreational water exposures (Colford et al., 2007). Male-specific coliphages, specifically the ssRNA *Leviviridae* family, are superficially indistinguishable from most human enteric viruses (Grabow, 2001), occur in higher numbers in sewage and wastewater effluents than viral enteric pathogens (Grabow, 2001), their presence implies the presence of pathogenic viruses (Grabow, 2001), and, in a majority of cases, they provide human/animal fecal-source specificity (Vinjé et al, 2004; Cole et al, 2003; Furuse, 1987; Schaper et al, 2002; Scott et al, 2002; Stewart, 2002; Long et al, 2005).

The focus of this research was to (1) generate a nucleotide sequence database by

sequencing at least three to five strains from each FRNA coliphage genogroup (I, II, III and IV), (2) develop and analyze for identification of preferred targeted genomic regions the sequence database representing environmental and prototype strains and (3) based on the primers identified in step 2, design and validate a molecular assay to detect and ultimately subtype the different genogroups. To develop a genetic database, 19 FRNA strains were sequenced. In addition, two new undescribed *Levivirus* strains were sequenced. In addition, a one-step reverse transcription polymerase chain reaction (RT-PCR) detection method for the four FRNA coliphage groups (I, II, III and IV) was designed to distinguish human vs. animal fecal source. Highly precise forward and reverse genogroup-specific primers were designed based on a total of 30 FRNA sequences containing several strains from all four genogroups.

II. Background

To develop valid fecal-indicator criteria based on credible epidemiology design, US EPA undertook a series of marine and freshwater public-beaches studies ([EPA-600/1-84-004](#); [EPA-600/1-80-031](#)). Objectives were to assess the mathematical relationship between microbiological indicator concentrations in bathing water and illness rates resulting from recreational water exposure (swimming), in order to correct for perceived deficiencies in US Public Health Service studies conducted before 1972. Additional goals were to provide a statistically sound study outcome correlating the concentration of the best bacterial indicator with magnitude of health effects in bathers, resulting in specific bacterial concentration-health risk outcome relationships associated with swimming in sewage-contaminated waters. Two key US EPA documents were published from these epidemiological studies “Health Effects Criteria for Fresh Recreational Waters” ([EPA-600/1-84-004](#)) and “Health Effects Criteria for Marine Recreational Waters” ([EPA-600/1-80-031](#)). Epidemiological data supported the use of *E. coli* and enterococci as primary fecal indicators because they were associated with statistically significant increased gastrointestinal illness rates to swimmers/bathers at increasing concentration in water. Subsequently, EPA guidelines for recreational waters were established in the 1986 “Ambient Water Quality Criteria for Bacteria in Recreational Waters.”

To overcome deficiencies in the 1986 US EPA guidelines, the agency was compelled to implement the Beaches Environmental Assessment and Coastal Health Program (BEACH)

Act of 2000. The BEACH Act was an amended Section 303 of the Clean Water Act. The focus was to improve public health and recreational water quality programs by 1) strengthening beach testing and standards, 2) providing faster testing methods, 3) predicting fecal pollution, 4) better defining the criteria as to which fecal indicators and water quality standards are based, 5) investing in health and methods research and 6) informing the public ([EPA, 2003](#)).

There are limitations associated with the two current bacterial fecal indicators for recreational water quality, enterococci and *E. coli*. They do not provide a complete assessment of the sanitary quality of water. Their presence and densities do not correlate with all potential water-borne pathogens causing adverse environmental effects ([Griffin et al., 2003](#)). For example, most illnesses contracted by swimmers appear to be of viral etiology. Environmental and health risks associated with microbes can be as much a consequence of viral contamination as it can of contamination by any other microorganisms. Both microbial and epidemiological evidence suggests that the current EPA recreational water-quality criteria using two bacterial indicators have little or no correlation to the presence of pathogenic viruses or to health-risks from non-point source fecal contamination. In microbial water quality studies conducted in Florida, indicator bacteria counts satisfied EPA criteria limits in the presence of detectable human pathogenic viruses ([Griffin et al., 2003](#); [Griffin et al., 1999](#)). In other studies, waters satisfying *E. coli* standards for shellfish harvesting often harbored shellfish that contained human enteric viruses ([Dore et al., 2000](#)). In laboratory studies, bacterial indicators have been erroneous predictors of virus survival because viruses survived longer and were inactivated at slower rates than were bacteria ([Burkhardt III et al.,](#)

2000; Sinton et al., 1999; Sinton et al., 2002). Even when current bacteriological standards are met in recreational waters, risks to human health may be posed by viruses.

In other studies, *E. coli* and enterococci were detected in environmental niches of tropical or temperate climates (Hawaii, Guam and Florida) where there was no evidence of human fecal contamination. Such environmental reservoirs of these bacteria led to high indicator counts that were not associated with fecal contamination (Hardina and Fujioka, 1991; Roll and Fujioka, 1997; Byappanahalli and Fujioka, 1998; Fujioka and Byappanahalli, 2000; Solo-Gabriele HM et al., 2000; Genthner et al., 2005). In some cases, elevated bacterial indicator counts exceeding EPA water-quality criteria were influenced by soil run-off and not a result of sewage input (Byappanahalli and Fujioka, 2004).

The apparent weaknesses in current bacterial indicators and the need to develop better, faster methods to accurately measure fecal contamination and determine source (animal vs human) supports the proposed use of a suite of indicator species, including at least one viral indicator for more specific and timely water-quality public health assessments. An attractive viral candidate is male-specific (F+), ssRNA bacteriophages (FRNA coliphages). They are viruses that are morphologically similar to most enteric viruses (Grabow, 2001), their presence in sewage and wastewater-impacted waters implies the presence of pathogenic viruses (Grabow, 2001), they are associated with “fresh” fecal contamination, they are easily detected in environmental samples, they survive disinfection treatments similar to other pathogenic viruses (Duran et al, 2003, Nappier et al, 2006) and they occur in higher numbers in sewage and wastewater effluents than viral enteric pathogens (Grabow, 2001). FRNA phages have been isolated from treated wastewater samples in the absence of fecal coliforms (Stewart, 2002), they provide insight as to contamination source (Vinjé et al., 2004; Cole et

al., 2003; Furuse, 1987; Schaper et al., 2002; Scott et al., 2002; Stewart, 2002; Long et al., 2005) and the potential exists to develop and use rapid nucleic-acid based molecular detection for real-time public-health risk management.

To design and develop a robust molecular genogroup-specific assay, the analytical system must be based on a genetically-representative sequence database. To identify appropriate genomic regions to target for primer design and amplification the use of only the few FRNA phages that have been fully sequenced over the past few decades is inadequate. As of 2007, eleven full-length genome sequences were available in GenBank (GenBank/EMBL/DDBJ). The use of more comprehensive and representative genetic sequence data for these coliphages would provide greater assurance of the development of a reliable molecular detection system based on RT-PCR amplification.

III. Research Objectives

Specific Goal:

1. To develop and validate a rapid genetic typing assay for the detection and characterization of FRNA coliphages from various geographical locations and sources representing each genogroup (I, II, III, IV).

Study Design:

1. Increase the full-genomic FRNA sequence database by selecting and sequencing representative strains from each genogroup.
2. Conduct phylogenetic analyses, identify nucleotide similarities, locate Open Reading Frame (ORF) positions and gene locations from these sequences.
3. Perform bioinformatics analysis to reveal protein domains, family conservations and discrete amino acid sequence features, or motifs, by Pfam and PROSITE patterns.
4. Identify optimal target regions for the development of a one-step reverse transcription-PCR assay specific for each genogroup.
5. Design and validate the one-step RT-PCR assay specific for each genogroup.
6. Identify optimal target regions to design primers and probes for the development of a real-time PCR assay specific for each genogroup.
7. Design the real-time assays to detect and discriminate the four *Leviviridae* genogroups, I, II, III and IV.

IV. Literature Review

Coliphage History

Between 1915-1917, bacterial viruses or bacteriophages, were first noted independently by Frederick William Twort and Felix Hubert d'Hérelle. Approximately 20 years earlier, plant viruses (1892 by Ivanowski) and animal viruses (1902 by Loeffler and Froesch) had been described ([Duckworth, 1987](#)). During the late 1930's, Emory L. Ellis isolated phages from Pasadena, CA, sewage by using *E. coli* as a host bacterium. In 1942, bacteriophages were first observed using electron microscopy ([Ackermann, 2006](#)) by Tom Anderson (www.asm.org). Bacteriophage lambda was discovered in 1951 by Esther Lederberg while observing a lysogenic phage isolate from *E. coli* ([Ackermann, 2006](#)). Throughout the 20th century, the culmination of phage research laid the foundation of modern molecular biology.

An RNA phage, strain f2, was isolated and characterized in 1961 by Loeb and Zinder ([Loeb and Zinder, 1961](#); [Zinder, 1965](#); [Furuse, 1987](#); [van Duin, 1988](#)) and in 1959, Dr. A.J. Clark isolated the most well-studied RNA phage, strain MS2 (personal communication). These ssRNA coliphages infect gram-negative bacteria expressing a sex-pili (F⁺), are sensitive to RNase and are collectively known as male-specific phages (FRNA). Using electron microscopy, Bradley ([1964](#)) first described RNA-containing phage morphology as an icosahedral form. In 1976, strain MS2 was the first virus to be completely sequenced

(Fiers et al., 1976). Genomic sequences could now be compared to serological and physicochemical typing.

Taxonomy

Early taxonomic classification of phages, between 1920s and 1930s, was based on bacterial host specificity. In the 1940s and 1950s, the advent of electron microscopy propelled phage taxonomy to morphological descriptions (Nelson, 2004). Techniques to isolate nucleic acids and determine the genome composition, i.e., dsDNA, dsRNA, ssRNA, ssDNA, provided a system of viral taxonomy based on nucleic acid type and morphology published by Lwoff, Horne and Tournier in 1962 (Ackermann, 2006). Bradley proposed a classification scheme based on basic phage features, i.e., nucleic acid type (DNA or RNA, ss or ds, circular or linear), capsid morphology, tailed or filamentous phages, enveloped or non-enveloped, etc. By the late 1960s to early 1970s, the International Committee on Taxonomy of Viruses (ICTV) formalized phage classification into six genera based on Bradley's proposed scheme. As new phage types are discovered, they are added to the ICTV classification of one order and 13 families. The largest viral group is the bacteriophages, with the predominant phage type having dsDNA and a smaller number of phage types having ssRNA (Ackermann, 2006).

Genomic or partial genomic sequencing has become the norm in the modern virology laboratory displacing the historical electron microscopy visualization of phage structures. Rohwer and Edwards proposed a sequence-based taxonomic system based on “signature genes” elucidated in phage genomes comparable to the 16S rDNA classification in bacteria (Rohwer and Edwards, 2002). Since no single protein marker or motif was conserved throughout the range of phage genomes, phage relatedness was based on common

characteristics. The more individual characteristics that were shared such as protein sequences, nucleic acid form, etc, the stronger the relationship. Protein-distance programs calculated the number of amino acid changes from protein-to-protein and were a resourceful tool in determining the fine parameters of the phage proteomic tree. The resulting proteomic tree was compatible with the ICTV arrangement ([Rohwer and Edwards, 2002](#)).

Male-specific phages belong to either the *Leviviridae*, single-strand RNA, or *Inoviridae*, a single-stranded DNA family ([Sobsey et al., 2005](#)) ([Fig 4.1](#), [Fig 4.2](#)). Male-specific RNA phages (FRNA) are non-enveloped, positive sense, single-strand RNA (ssRNA) contained within a 26 nm diameter icosahedral-shaped capsid ([Buchen-Osmond, 2003](#)). The *Leviviridae* family ([Table 4.1](#)) comprises two genera, *Levivirus* and *Allolevivirus*. These genera are further divided into four genogroups (I, II, III and IV). *Levivirus* are genetically divided into genogroups I and II, and *Allolevivirus* are subdivided into genogroups III and IV.

Leviviridae FRNA phages were initially grouped primarily according to their serological properties in that anti-phage sera prepared against specific phage strains could neutralize some of the other closely-related phages ([Watanabe et al., 1967](#); [Miyake et al., 1971](#)). Other methods of grouping were through membrane filtration, elution patterns and buoyant densities ([Miyake et al., 1969](#)). Until 1969, three phage groups existed, groups I, II and III and subgroups a,b and c in group III. With the isolation and characterization of strains SP and FI, serological assays indicated that these two strains did not cross-react to anti-sera from groups I, II or III. A new group IV and group V were proposed for strains SP and FI, respectively ([Miyake et al., 1969](#); [Sakurai et al., 1988](#)). Eventually, four major groups, I, II, III and IV and subgroups a,b, c and d in group III and subgroups a and b in

group IV were assigned to FRNA phages based on template specificity of RNA replicase (Miyake et al., 1971; Miyake et al., 1973).

Based on a limited number of complete sequences four FRNA genes could be identified (reviewed by Bollback & Huelsenbeck, 2001). These genes code for an assembly or maturation protein, capsid protein, lysis protein and replicase protein in the *Leviviruses* whereas in *Alloleviviruses*, the lysis protein is replaced by a read-through protein. Each levivirus virion contains one molecule of positive sense ssRNA, 180 copies of the capsid or coat protein, one copy of the maturation protein and, in alloleviviruses, approximately 15 copies of the read-through protein (Weber & Konigsberg, 1975; van Duin, 2000; van Duin and Tsareva, 2006). Read-through protein synthesis occurs at a rate of about 6% and, although the exact function is unknown, the combination of the read-through protein with the maturation protein is required for an infectious viral stage (van Duin and Tsareva, 2006). The *Leviviridae* maturation protein is needed for viral infection and virus particle maturation (Olsthorn et al., 1995) whereas the numerous coat proteins in the virion are used for assembly of phage progeny (Klovins et al. 1997; Weber and Konigsberg, 1975). A single, small (< 100 amino acids), hydrophobic peptide, the lysis peptide, is responsible for cell lysis at the end of the infection cycle in the *Levivirus* genogroups (van Duin, 1988). No distinct lysis protein is present in *Alloleviviruses*; lysis is mediated by the maturation protein (Karnik and Billeter, 1983). The replicase in both genera, also known as viral RNA-dependent RNA polymerase, is required in small amounts early in the infection process. Twenty minutes after infection, synthesis of this protein ceases (van Duin, 1988).

Table 4.1 Bacteriophage classification.

Family	Number of Genera	Morphology	Nucleic Acid	Enveloped
Cystoviridae	1	isometric	ssRNA, L, S	Yes
Leviviridae	2	icosahedral	ssRNA, L	
Corticoviridae	1	isometric	dsDNA, C, T	
Microviridae	4	icosahedral	ssDNA, C	
Tectiviridae	1	icosahedral	dsDNA, L	
Inoviridae	2	rod	ssDNA, C	
Myoviridae	6	contractile tail	dsDNA, L	
Siphoviridae	6	noncontractile tail	dsDNA, L	
Podoviridae	3	short tail	dsDNA, L	
Lipothrixviridae	1	rod	dsDNA, L	Yes
Rudiviridae	1	rod	dsDNA, L	
Plasmaviridae	1	pleomorphic	dsDNA, C, T	Yes
Fuselloviridae	1	lemon-shaped	dsDNA, C, T	Yes

ss - single stranded; L - linear; S - segmented; C - circular; T - superhelical; ds - double stranded

Ecology and Source Specificity

Male-specific ssRNA phages, or FRNA, strains MS2 (isolated by A. Clark), R17 (isolated by Paranchych and Graham) and f2 collected in the late 1950's to early 1960's from the United States were typed into group I. Throughout the 1970's, K. Furuse, I. Watanabe and colleagues conducted a systematic survey by isolating several thousand ssRNA coliphage strains from sewage in domestic drains, feces from humans and various animals, municipal raw sewage, river water, seawater, irrigation, pond or lake waters from across the globe. They classified these phages into four groups, I, II, III and IV, based on antisera testing or sero-typing and by physicochemical characteristics such as a cesium chloride density profile. Some strains were further subdivided into three to seven subgroups. Group III phages were

predominant in the southwest islands of Japan, the Philippines, Indonesia, Taiwan and Singapore and group II phages were most abundant in mainland Japan (Furuse et al., 1978; Miyake et al., 1971). Domestic drains located in Asian countries had very few FRNA from groups I and IV. Phages isolated from domestic drainage in Korea were from groups II and III whereas southeast Asia phages were predominantly group III (Furuse, 1987). FRNA phages isolated from sewage water in Australia were from group II and, in the USA, groups II and III FRNA phages were collected (Furuse et al., 1975).

FRNA phage isolation frequencies from various countries varied greatly. Only 2.5% of phages collected from sewage in a study in Peru were FRNA, 5% were FRNA in Brazil and by comparison, FRNA were 38% in Taiwan and 30% in Japan (Furuse et al., 1975). The FRNA phages from sewage samples in Brazil and West Germany belonged to group I exclusively. It was unknown whether or not these sewage treatment plants received slaughterhouse waste (Furuse, 1987). Furuse states “it can be reasonably assumed that group I phage observed in raw sewage from treatment plants are most likely introduced from animal sources, and group II and group III phage from human sources.” A separate study in Brazil found that thirteen out of 353 sewage and/or fecal samples contained FRNA phages, or approximately 4% and of these thirteen, five were typed as group I, two as group II and six as group III (Miyake et al., 1973).

Out of 93 phages isolated from the Bangkok, Thailand collection, only eight were FRNA phages. Three independent samples were collected from river water, two samples were collected from sewage and no FRNA phages were isolated from stool samples. Of the eight phages, one strain was group I whereas three and four strains were typed as groups II

and III, respectively ([Aoi et al., 1972](#)).

Domestic sewage samples in Korea had 56% RNA phages belonging primarily to groups II and III along with 4 group I phages ([Osawa et al., 1981](#)). Taiwan sewage and fecal samples included 20% FRNA with 2 group IV, 22 group III, 8 group II and 6 group I isolates ([Miyake et al., 1971](#)).

Furuse and colleagues continued to further explore the distribution patterns of the four genogroups of FRNA phages in samples from the following sources: 1) gastrointestinal contents of cows and pigs, 2) feces of domesticated animals (pigs, horses, cattle and fowl), 3) human feces 4) animal feces and 5) sewage obtained from slaughterhouse treatment plants. FRNA groups II and III were isolated in almost equal proportions from human feces whereas the gastrointestinal tract of swine harbored groups I and II. Group I was isolated from feces and gastrointestinal contents from all other animals. Slaughterhouse samples were predominately group I along with a few isolates from group II. In these studies, group III was only isolated from human subjects and from no other host organism. Group IV displayed the greatest habitat diversity as they were isolated from feces of animals, humans, domestic drainage and raw sewage ([Furuse, 1987](#); [Osawa et al., 1981](#)). The Furuse study concluded group I phages were not found in humans or domestic drainage sewage but only in animals or

slaughterhouse sewage. However, their results conflict with the fact that group I strains MS2 and R17 were isolated from municipal sewage systems located in the USA.

FRNA phages isolated in The Netherlands from fecal samples obtained from humans and domesticated animals were serologically typed. As in the Furuse studies, groups I and IV phages were isolated exclusively from animals whereas human sources harbored groups II and III ([Furuse, 1987](#))

Groups II and III were isolated from human feces in FRNA distribution studies from South Africa and Spain ([Schaper et al., 2002](#)). In contrast to earlier reports of groups II and III occurring only in human feces and I and IV in animals, the Schaper study isolated group II from cattle and swine and group II and III from poultry. This study casts doubt on the absolute association of phage genotype and source-specificity, but emphasized the observation that group I has not been isolated from human feces. Nonetheless, group I has been isolated in domestic sewage and whether or not animal waste was present in these municipal treatment facilities is a question that cannot be easily answered.

The lack of consistent numbers of FRNA in human feces and the discrepancies of phage types harbored in various animals does not minimize the fact that FRNA are isolated from sewage in numbers ranging from 10^2 - 10^4 PFU/ml ([Leclerc et al., 2000](#)). From multiple samples over time, FRNA were isolated from raw sewage at an average of 4.2×10^4 PFU/L ([Brion et al., 2002](#)).

FRNA Coliphages as Indicators of Fecal Pollution

FRNA coliphages have been recommended as a possible indicator of enteric viruses as their presence indicates fecal pollution from either humans or animals. A highly significant statistical correlation was observed between the presence of FRNA and

enterovirus virus concentrations in river water, coagulated effluent, chlorinated and UV-irradiation effluents, coagulated river water and lake water but not in raw sewage or biologically-treated sewage (Havelaar et al., 1993). FRNA was proposed as a viral indicator in recreational waters due to the strong correlation between FRNA and enteroviruses. Limitations to this study were that only freshwater, not marine or other environmental water bodies were studied, and only a small geographical area was evaluated (Havelaar et al., 1993).

Marine, freshwater and estuarine waters were selected to study the relationship between coliphages, their *E. coli* host and a few pathogenic bacteria. Multiple sampling stations were located near domestic and industrial sewage discharges. A correlation between male-specific coliphage concentration and *E. coli* concentration were found to be dependent upon the direction and distance from the effluent plume. A greater statistical relationship was noted for male-specific coliphages and the pathogens *Salmonella*, *P. areuginosa* and *C. albicans* when compared to fecal and total coliforms (Borrego et al., 1987). In this study, FRNA and FDNA phages were not resolved separately.

Drinking water sources were analyzed for the presence of male-specific phages, somatic phages and *Bacteroides fragilis* phages for 30 months. Bacterial indicator assays, total coliforms and fecal coliforms, were also evaluated to determine if a relationship existed between the bacterial and viral indicators. *B. fragilis* HSP40 host was used to select the human-specific phage. Throughout the survey, total or fecal coliform positive sites were also positive for at least one or all three phage types. In some instances, coliform negative sites were bacteriophage positive. When the data was tabled as frequency of indicator organisms by year, F+ specific phages had the highest percentage of positive samples two out of three years. This study did not differentiate between FRNA and FDNA male-specific phages.

Phage *B. fragilis* was isolated less frequently two out of three years. Male-specific phages were suggested as an indicator similar to total coliforms and phage *B. fragilis* was similar to fecal coliforms as a measure of fecal pollution ([Armon and Kott, 1995](#)).

For a period of two years, raw sewage and surface water samples were surveyed once or twice per week from sites reflective of different land use and agricultural areas. Following double-agar overlay and RNase sensitivity, plaques were genotyped by hybridization using Hsu's method ([1995](#)). FRNA phages were the most abundant F+ phage collected in surface waters (67% out of 105 samples) and sewage (87.5% out of 288) samples. Surface waters were positive for type I (81%) and only one sample had type III, suggesting these surface waters were not influenced by human-impacted effluent. However, type III was the predominant genotype as 57% were isolated from raw sewage ([Brion et al., 2002](#)). During this study, type III FRNA were recovered more frequently than the other genotypes when a group of male campers were staying near the sampling site. Following their departure, the presence of type III declined and was no longer detectable after a week. Source-tracking with type III may indicate sewage contamination occurred within the past 7 days ([Brion et al., 2002](#)).

A ratio of FRNA to FDNA densities were compared from samples collected in animal feces, municipal wastewater facilities, in potentially impaired surface waters and in agricultural livestock wastewaters. To establish a correlation between FRNA genogroups and sample source, FRNA isolates were serotyped. Background samples were defined as an upstream or background site that was sampled concurrently with the surface water sample. FRNA phages were isolated from bovine waste (18%), swine wastewater (50%), gull (96%), goose (100%) and municipal/human wastewater samples (23%). In comparison, FDNA

isolation frequencies were bovine waste (82%), swine wastewaters (50%), gull (4%), goose (0) and human wastewater (77%) FDNA. The remaining animals, horse, buffalo, cat, cormorant, rooster, dog, llama, donkey and pig did not yield F+ coliphages. In animals, goose had the highest percentage of group I (approximately 98%) followed by swine (51%) and cattle (30%). Groups II and III were only isolated from human (50% group II, 15% III), swine (5% II, 22% III) and bovine (15% II, 0 III). Both FDNA and FRNA phages were isolated from surface waters impaired by human, swine, bovine and background sources. Group I FRNA were predominant in background surface waters (97%) whereas bovine-impacted surface waters had 82% group I followed by 75% group I from human-impacted waters. Human land use sites had the greatest percentage of group II isolates (12%) with a low group II recovery from background (2%) and bovine-waste sites. A correlation of FRNA source-associated genogroups was confounded by differential survival rates for each genogroup and/or strain. The authors concluded that there was a statistically significant link of group II FRNA associated with human-land use sites ([Cole et al., 2003](#)).

The US EPA sponsored a field validation of Methods 1601 and 1602 to detect coliphages, both male-specific and somatic, in groundwater systems. In addition, fecal indicators *E coli*, enterococci, total coliforms, *Clostridium perfringens* spores and the presence of enteric viruses (enterovirus, hepatitis A, norovirus, rotavirus and adenovirus) were monitored monthly for one year. Male-specific coliphages, but not specifically FRNA, were recommended by the proposed Groundwater Rule as an alternate fecal indicator as they were detected more often in groundwater sources than somatic coliphages by Method 1602. The study concluded that the use of both a bacterium indicator and a coliphage increased the predictability of fecal contamination in groundwater wells ([American Water Works](#)

[Association., 2004](#)).

Distinct geographical locations (New Mexico, Massachusetts, Connecticut, Michigan, the Carolinas and Southern California) across the USA were surveyed in an ecological study of FDNA, FRNA and somatic phages. Phage concentrations were compared to fecal coliform counts. Viruses were isolated and enumerated on double-agar overlay and the appropriate phage-specific *E. coli* host. Further separation of phage types were determined by RNase sensitivity followed by serotyping. RT-PCR methods followed by Reverse Line Blot (RLB) methods were used to genotype the phages. Direct fecal samples did not have detectable levels (<3 PFU/gram feces) of male-specific phages in feces other than chicken litter, gull and goose. Only FRNA groups I and IV were detected in these avian species. Cow lagoon isolates typed to group I, whereas hog lagoon predominantly typed group I (32%), 3% group IV along with 18% group III. Hog lagoon had the highest percent of group III FRNA. Wastewater influent and effluent were predominately group II with 12 and 15%, respectively. In addition, wastewater influent had 6% group III, 2% I and 1% IV of FRNA phages. Interestingly, septic water samples did not contain FRNA, but had 100% FDNA strain M13. Grazing animal feces contained large numbers of somatic phages. It was concluded that FDNA M13-like was most prevalent in wastewaters, FDNA were detectable in fecal samples and the link between FRNA group III as a human-associated effluent was not absolute ([Long et al., 2005](#)).

To add to the body of information on FRNA occurrences from different sources, phages were collected from wastestreams at two hospitals, a cattle feedlot, pig farm and a poultry farm. Environmental river water samples were taken adjacent to farming fields. Male-specific phages were selected on *S. typhimurium* WG 49 host by double-agar overlay,

transferred and fixed onto a membrane and hybridized with the Beekwilder et al., (1996) designed probes. Hybridization probes detected groups II and III in hospital wastewaters, groups III and IV from swine waters, groups I, III and IV from poultry and groups I and IV from cattle wastes. As a whole, river water samples contained all four genogroups. However, seven of the individual river samples were positive for only genogroup II. Analysis of FRNA from an assortment of sources led the authors to conclude that group III was not necessarily specific for human excreta, but the trend on the specificity of groups I and IV for animal sources and II and III for human sources supported previous findings (Sundram et al., 2006).

The ambiguous association of swine isolates of FRNA coliphages periodically grouping as type III led to the hypothesis that perhaps a refined genogrouping system could clarify if sub- or unique group III clusters existed. Forward and reverse primers for reverse-transcription polymerase chain reaction (RT-PCR) were designed to the 5' untranslated region spanning into the maturation gene region in group III phages. Primers were based on known group III sequences to strains MX1, M11 and Q β . Isolates were first classified to group III by hybridization (Hsu et al., 1995; Beekwilder et al., 1996). Thirty-two type III coliphage strains were isolated from swine lagoons, surface and wastewater sources from North Carolina and South Carolina. RT-PCR amplification was performed on those isolates testing positive for group III. RT-PCR products of about 567 nucleotides were sequenced and phylogenetic trees were generated from these sequences. Phages isolated from North Carolina lagoons, surface and wastewaters grouped as Q β -like whereas those from South Carolina were closely related to prototype strain M11. In this case, group III isolates could not be genetically separated as human vs swine clusters. The authors noted genetic similarity

evidence from human and swine hepatitis E virus populations being consistent with their finding for FRNA phages ([Stewart et al., 2006](#)).

Ninety-six surface water stations on the State of South Carolina's impaired list were selected as study sites to quantify FRNA and somatic coliphages. Typing FRNA occurred by serological and/or nucleic acid hybridization methods. Fourteen of these sites identified FRNA genogroups II (5%) and III (1%) while the majority of isolates typed as group I (94%). Direct wastewater samples typed 73% group III, 14% II and 11% group I. Isolates collected and typed from a swine lagoon had 70% group I, 19% group III and 6% group IV. Chicken litters contained approximately equal amounts of group I and IV FRNA. The presence of groups II and III from the 14 surface water sites were mainly located downstream of wastewater effluent discharges and considered to be contaminated by human fecal pollution ([Stewart-Pullaro et al., 2006](#)).

Sewage-polluted tropical river waters and animal fecal samples were examined for male-specific RNA, male-specific DNA and somatic coliphages. Male-specific DNA and FRNA occurred at similar quantities per gram feces or per ml of water at 7% and 6.5%, respectively. F-specific phages were isolated by plaque assay from 50% of the river samples and 4.4% from the animal fecal material. The study concluded the presence of FRNA phages in the tropical waters of Klang Valley, Malaysia, and proposed FRNA as a tool for monitoring fecal-polluted waters in their country ([Yee et al., 2006](#)).

A small-scale study using male-specific, somatic phages and F+ *E. coli* as tracers of sewage, of monthly fluctuations and of various populations were investigated in a small Israeli community, Kibbutz Yagur, near Haifa, Israel. The well-defined sewage lines collect from daycare centers, residential, factory, dairy, greenhouse, dining hall, laundry room and

clinic with manholes specific to each location. Phages were not isolated from the manholes that served the laundry area and were found to occur in very low numbers (<10 PFU/ml) from the elderly home, the day-care center that used pampers, the greenhouse and the factory. The dairy farm was prevalent in somatic phages (10^2 - 10^3 PFU/ml) but had low counts of male-specific phages (1- 10 PFU/ml). Direct fecal material from newborn infants contained male-specific phages at concentrations of 10 - 10^5 PFU/g feces with one child excreting phages for almost 8 months. Absence or presence of F+ *E. coli* in the same water sample(s) correlated with low or high male-specific phage counts. Throughout the year-long study, there were higher numbers and more positive samples of male-specific phages than somatic phages with approximately 96-98.5% of typed phages being FRNA (Gino et al., 2007). Phages were not genogrouped or typed in this study.

FRNA Host Specificity

E. coli K-12 strain was studied in terms its ability or inability to transfer a sex factor, termed F. If cells transferred F to other cells by means of chromosomal markers, this was termed Hfr strains. If the F factor was transferred independent of the chromosome but through an extra-chromosomal state, or plasmid, the strains were known as F+ (Clark, 1963). Electron microscopy shed light onto the aggregate nature of FRNA phages adsorbed to the host cell's fimbriae, the F pili (Bradley, 1964) (Fig 4.1). The conjugative pili serve to transfer genetic information by horizontal gene transfer in gram-negative bacteria. Both Hfr and F+ strains are derived from *E. coli* K-12 (Havelaar and Hogeboom, 1984).

The male-specific DNA and RNA coliphages adsorb to these conjugative pili (Daehnel et al., 2005), a fertility (F+) sex-pili, coded in *E. coli* by the F-plasmid (Paranchych, 1975). DNA and RNA coliphages bind to the F-pili in different manners.

DNA phages bind to the tip of the F-pili and RNA phages attach to the sides (Daehnel et al., 2005).

The Furuse studies enumerated FRNA phages by incorporating male strains of *E. coli* (F⁺, F' or Hfr) into the media plates followed by RNase treatment (Furuse, 1987).

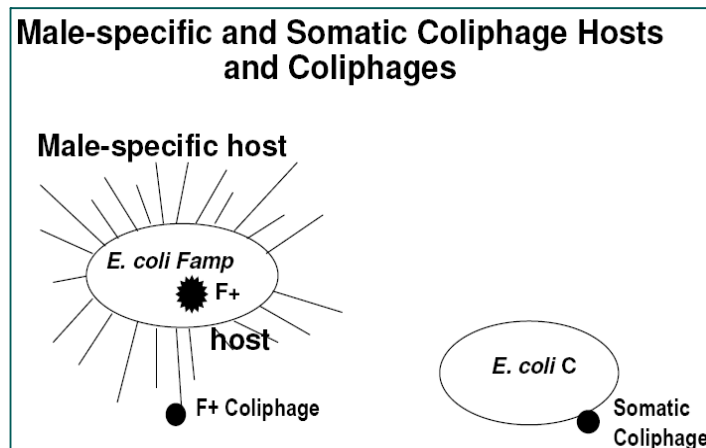
Salmonella typhimurium strain WG 49 that express F⁺ by the presence of an F-plasmid have also been used as a selective host to isolate male-specific phages (Havelaar et al., 1984). *S. typhimurium* detected somatic and FDNA phages, however, the host selected 90-95% FRNA phages (Sundram et al., 2006). *E. coli* strain F_{amp} (ATCC 700891) is commonly used in host-selection procedures for F⁺ coliphages.

Phage replication is restricted to environments such as the intestinal tract of warm-blooded animals as the sex-pili are only expressed at temperatures greater than 30 °C (Grabow, 2001). Interesting to note, Zinder (1963) and Bradley (1964) theorized that in order for male-specific phages to be plentiful in nature, then subsequently, the *E. coli* male strains must either be equally prevalent in nature or they are not the natural environmental host. However, bacteria from animal intestines harbor hosts with pili at 10⁴ CFU/ml in wastewater, thereby possibly allowing phage replication (Long et al., 2005) in municipal wastewater systems.

FRNA phages are more abundant in sewage waters (10³ - 10⁴ PFU/ml) than in human and animal feces (up to 10³ per gram feces) (Gerba, 2006). To examine this phenomenon, FRNA strain GA was inoculated into pasteurized sewage, seawater, and/or river water. Infectious strain GA underwent multiplication at 20° C but only when host bacterium was cultivated at 37° C (Havelaar and Pot-Hogeboom, 1988). Due to temperature restrictions of

the host-pili expression, it seems plausible that FRNA phage environmental replication would be restricted to $> 30^{\circ}\text{C}$. Optimal phage growth temperatures were found to be 30°C for genogroup II and 37°C for genogroups I, III and IV when three strains per genogroup were evaluated (Furuse, 1987). Other studies also reported different survival temperatures in water temperatures $\geq 15^{\circ}\text{C}$ (Sobsey, ftp.sccwrp.org). In addition, phage replication requires a host density of approximately 10^4 bacteria/ml to be successful (Goyal et al., 1987). The need for concomitant events of optimal temperatures and the presence of an F⁺ host in log phase would restrict environmental replication of FRNA coliphages.

Figure 4.1 Somatic and male-specific coliphage hosts.



Additional Potential Viral Indicators of Fecal Pollution

Bacteriophages are ubiquitous in nature and as a whole, are not suitable as an environmental water quality fecal indicator. However, unique bacteriophages are associated with sewage. Various phages have been proposed as viral indicators. Somatic coliphages (Fig 4.1), FRNA coliphages, FDNA coliphages and the *Bacteriodes* phage have been recommended. Advantages and limitations of each indicator will be addressed.

Somatic coliphages are present among four bacteriophage taxonomic groups, *Myoviridae*, *Styloviridae*, *Podoviridae* and *Microviridae*, hence the morphologic heterogeneity. Three families of somatic phages contain dsDNA and one family, *Microviridae*, contain ssDNA. The somatic phages are non-enveloped and are present in sewage, often in high abundance in untreated sewage, ranging from 10^4 - 10^5 PFU/ml (Gerba, 2006). These coliphages have been detected in humans, chickens, pigs, other animals (Gerba, 2006) and are easily cultured using *E. coli* strain CN-13 (ATCC 700609). The concentrations of somatic phages are similar to FRNA, 10^2 - 10^4 infectious units per liter (Sobsey, <ftp.sccwrp.org>) The phage adsorb to the *E. coli* host and other enterobacteria species via the cell wall with a basic receptor site. A limitation of somatic phages is the possibility that the bacteria host origin, especially from environmental reservoirs, may not have originated from a fecal source. Some somatic phages may replicate in environmental waters in the absence of fecal pollution (Ashbolt et al., 2001; Gerba, 2006). If the presence of somatic phages were unrelated to fecal contamination, then it would not serve to predict human health risk (Leclerc et al., 2000).

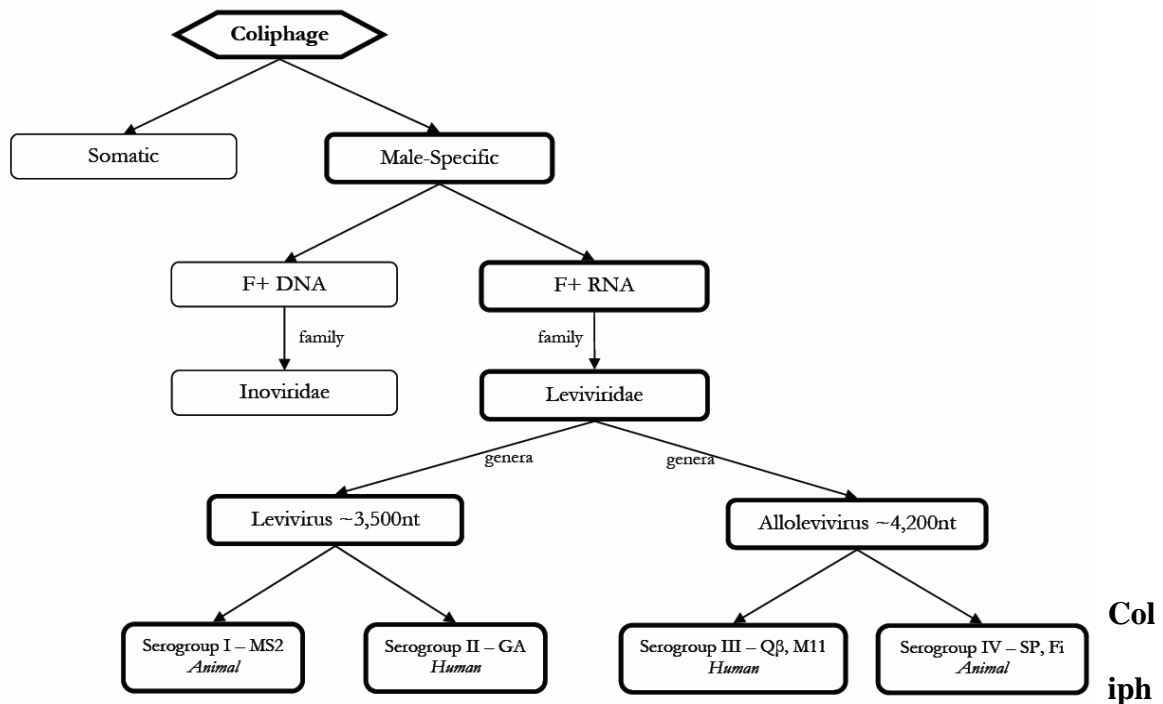
Inoviridae, a male-specific circular, single-stranded family of DNA viruses (Fig 4.2)

are filamentous, non-enveloped with a genome size of 6 kb to 9 kb and a virion length of approximately 700 nm to 2000 nm (www.virustaxonomyonline.com). Male-specific DNA coliphages are not morphologically similar to human enteric viruses and their ecology has not been extensively studied ([Leclerc et al., 2000](#)). Contradictions in the literature exist on whether FDNA or FRNA are more abundant in nature. Leclerc et al., (2000) noted FDNA were less abundant when compared to FRNA phages. Different distributions of FDNA and FRNA have been reported ranging from 52% FRNA and 48% FDNA ([Vinjé et al., 2004](#)) to 77% FDNA and 23% FRNA ([Long et al., 2005](#)). Cole et al., (2003) found higher numbers of FDNA in bovine wastes. Using Reverse-Line Blot hybridization and reverse-transcription-PCR followed by sequence analyses, one study found no link between FDNA type and source ([Vinjé et al., 2004](#)). However, a second study observed FDNA strain M13 as the dominant phage in septic system samples ([Long et al., 2005](#)). Thus, a definitive association between FDNA and human fecal pollution has not yet been established.

Certain phages belonging to the *Siphoviridae* family are approx 60 nm virion size, contain dsDNA with long non-contractile tails and infect via cell wall attachment to the host bacteria, strain *Bacteroides fragilis*. Host strain *B. fragilis* HSP 40 selects for a human-specific phage. The *B. fragilis* human-specific phage has been isolated in sewage, human fecal samples, polluted groundwater, seawater, river water and sediments but not detected in animal feces ([Bitton, 2005](#); [Cornax et al., 1990](#); [Tartera and Jofre, 1987](#)). Unlike other viral or bacterial indicators, the *Bacteroides* phage is considered to be an exclusive viral indicator of human fecal pollution and unlikely to replicate in the environment as the host is a strict anaerobe ([Tartera et al., 1989](#)). Human-specific phages can be rapidly detected with molecular primers without direct historical cultivation since animal vs human specific probes

are available (Bernhard et al., 2003). A limiting factor lies in the observations of a low isolation frequency, 0-15%, of *B. fragilis* phages were isolated from human feces (Gantzer et al., 2002). *B. fragilis* host are anaerobic and die-off rapidly under ambient water environmental conditions. As with other viruses, molecular detection only reveals the nucleic acid presence or persistence of the phage, not necessarily the infective or biologically active form. Another *Bacteroides* host, strain RYC2056, is more abundant but not human specific (Ashbolt et al., 2001). *B. fragilis* strain RYC2056 occurs in domestic sewage and was used to select phage from 30% of swine and 28% of human fecal samples (Puig et al., 1999).

Figure 4.2. Schematic of male-specific coliphage classification.



As sewage and its accompanying microbial population are dispersed from point or non-point sources, microbes entering marine and freshwaters are subjected to the surrounding ecological conditions. Water-quality parameters such as pH, hardness, salinity, degree and intensity of sunlight exposure and temperature (climate) strongly influence microbe survival. Survival differs among microbial types, even from virus to virus. Virus types, enveloped or non-enveloped, vary greatly in environmental survival, as enveloped viruses tend to survive poorly outside of their host compared to the survival of non-enveloped viruses. F+ phages and enteric virus groups such as adenovirus, enterovirus, norovirus (calicivirus) and rotavirus are all non-enveloped viruses.

Survival was evaluated for the indicator organisms *E. coli*, *C. perfringens*, fecal coliforms and F+ phages in estuarine conditions. A series of parameters were selected such as temperature, salinity, dissolved oxygen, solar radiation, season and geographic location to study inactivation rates. The highest decay rates were influenced by sunlight and/or temperature. F+ phages exhibited the least decay (83%) whereas fecal coliforms had the highest decline at 99%. Phages and *C. perfringens* were least affected by temperature. This study provides corroborative evidence that F+ phage inactivation rates differ from bacterial indicators ([Burkhardt III et al., 2000](#)).

Representative prototype and genotyped field isolated strains of FRNA were spiked at concentrations of 10^5 - 10^7 infectious units into FRNA-free, untreated surface waters (freshwater). Samples were incubated at 25 ° C in the dark and titered over time to determine phage decline or inactivation. Aliquots were removed for FRNA quantitation at day 4, 7, 13 and 36. Initial titers were determined by double-agar overlay plaque assay. The last sample taken, day 36, was enriched and assayed with the two-step enrichment procedure.

At day 4, rates of inactivation for prototype strains were as follows: genogroup I MS2 4.7 log₁₀, genogroup II GA 2.6 log₁₀, genogroup III QB 1.7 log₁₀ and genogroup IV FI 2.7 log₁₀ and SP (inactivation log data not provided). Prototype strain SP had the poorest survival and was not detectable after 4 days. Neither strain SP or QB could be detected by the two-step enrichment at day 36. Strains MS2, GA and FI were detected by enrichment at day 36. GA had the longest survival duration and was detectable by DAL throughout the experiment (day 36) ([Brion et al., 2002](#)). Survival of genotyped environmental isolates for FRNA coliphages was compared to their respective prototype strain. Of four, group I field strains, two survived similar to MS2 whereas the remaining two isolates were detectable by DAL to the end of the experiment, day 36. In group II, GA and one field II strain survived the longest (day 36) and the remaining three, group II field isolates declined by day 15. Type III isolates were below detection limits by day 13. Survivability was strain-specific and not necessarily influenced by genotype or prototype vs field isolates in freshwater systems ([Brion et al., 2002](#)).

A non-water matrix was used to compare the inactivation of MS2 to other pathogenic viruses, norovirus and poliovirus type 1, and *E. coli*. Different microcosms were filled with representative soils, i.e., organic muck, clay and sand. For one month, viruses seeded in a groundwater matrix at approx 10⁶ were dosed into the soil microcosm twice/week. The following month, the columns were dosed with a simulation of rainwater. Microcosm effluent was drained and viral strains were detected by RT-PCR or infectivity assays. With the exception of *E. coli*, the viruses were detected and shown to pass through the column. The results suggested that *E. coli* was not a reliable viral indicator as the bacteria did not pass through the columns in the manner as the virus and ultimately, environmental bacterial

transport would not mimic virus transport. None of the tested indicators passed through the clay column ([Meschke and Sobsey, 2003](#)).

Freshwater, specifically lake waters, were used in a microcosm format to evaluate survival rates on the four FRNA genogroups for 110 days. Strains MS2, two environmental group I, a group II, a group III and two group IV phages were inoculated (2×10^4 PFU/ml) into the microcosms. Incubation temperatures were 4 and 20 °C. Survival time was greater at lower temperature (4 °C). Group IV had the fastest decay rate (5 log₁₀ within 10 days, 4 °C) followed by group III (5 log₁₀ at 3 weeks, 4 °C), where these isolates reached the limits of detection at 10 days and 3 weeks, respectively. Rates of inactivation for groups I and II at 110 days, 4 °C were 1 log₁₀ and 3 log₁₀, respectively. The more persistent genogroups, groups I and II, were detected at 110 days ([Long and Sobsey, 2004](#)).

Prototype strains MS2, GA, Qβ and FI were spiked into seawater at concentrations of approximately 10⁶ PFU/ml and incubated, in the dark, at 21-23 °C. One ml aliquots were removed daily and titers observed by double-agar overlay (DAL) and real-time PCR, concurrently. Within 7 days, all four subgroups in seawater were no longer detected by DAL. In contrast, the real-time PCR detected the four groups after 20 days ([Kirs and Smith, 2007](#)). PCR detection of RNA, as demonstrated in these experiments, does not predict or indicate the presence of infectious phage but only the presence of RNA.

Viruses, including phage MS2, were tested as a molecular model for human norovirus. Spiked surface and groundwater samples were assayed for viral nucleic acid presence at 25 and 4 °C using QRT-PCR. In some strains, cell-culture infectivity and the DAL method for MS2 were used as concurrent methods of detection. MS2 nucleic acid detection was not effected by water source. Infectivity reduction rates were observed to

significantly increase at 25 ° C compared to 4 ° C. Infectivity reduction rates were not observed at 4 ° C (Bae and Schwab, 2008).

FRNA inactivation rates are influenced by sunlight intensity, fresh vs saltwater matrices and soil composition. For example, increasing salinity and temperature demonstrated faster rates of inactivation in FRNA strains. In contrast, lower temperatures (4 ° C) and freshwater environments decreased inactivation rates on FRNA coliphages.

FRNA Coliphage Detection Methods - Plaque Assays

To quantify and accurately detect FRNA phages, initial purification methods are imperative. Depending upon the application, detection methods include Single Agar Overlay, Double-Agar Overlay, enrichment, Most Probable Number, RNase sensitivity, serotyping and genotyping.

To prepare a Single Agar Overlay (SAL) for enumeration of FRNA, collect 250 ml environmental water sample and place on ice up to 6 hrs. Beforehand, begin a log-phase of the *E. coli* host F_{AMP} (ATCC # 700891) by adding a 1ml inoculum from an overnight (O/N) culture into 50 ml Trypticase Soy Broth. Gently shake at 37 ° C for 4 hr. Host culture will be in log-phase at 4 hr. Divide the environmental sample by dispensing two aliquots of 100 ml into sterile bottles. To the 2X TSA (trypticase soy agar) add 1 ml of 100 X streptomycin/ampicillin, 0.5 ml sterile 4M MgCl₂. Add 10 ml log-phase F+ host to the 100 ml sample. Quickly combine the sample/*E. coli* host with the 2X TSA/antibiotics flask. Gently swirl and allow host to contact sample for at least 3 minutes before pouring plates. Pour the agar/sample mix into a series of 20mm petri dishes at approx 20 ml/petri dish. Allow to harden and dry. Invert and incubate 36 ° C \pm 1 ° C. Count plaques per plate as

PFU/100 ml. Questionable plaques can be verified by spot plate assay ([EPA Method 1602, 2001](#)).

A presence/absence method involves a two-step enrichment. Using a sterile bottle, divide the 1 L environmental water sample into two, 500 ml aliquots. To each 500 ml, add the following: 50 ml of 10X cold TSB per L (media is cold to diminish phage growth), 12.5 ml 4M MgCl₂, 10 ml of 100 X streptomycin/ampicillin and 5 ml log-phase F+ host. Cap and invert to mix. Incubate overnight (16-24 hr) at 36 ° C \pm 1 ° C. The following day, spot 10 ul from each enrichment onto a grid spot plate (TSA) containing the appropriate host and incubate overnight. Score as positive or negative lysis zones in spots ([EPA Method 1601](#)).

An MPN (Most Probable Number) can be estimated from a 1L environmental water sample. Enrich the 1L sample with 2X TSB, antibiotics, 4M MgCl₂ and *E. coli* host as stated above. Once the reagents are added, quickly aliquot the enrichment into the following amounts in triplicate (A,B,C): (i) 300 ml A, 300 ml B, 300 ml C (ii) 30 ml A, 30 ml B, 30 ml C and (iii) 3 ml A, 3 ml B, 3 ml C. This dilution series is for a 3x3 MPN matrix. Incubate bottles/tubes at 37 ° C O/N. Remove 12 ml from each enrichment of 300 and 30 ml (A,B,C) except the 3 ml. Centrifuge all samples for 10 minutes at approx 5000 rpm. Transfer 10 ml of each supernatant into clean tubes; transfer most of the 3 ml supernatant into a clean tube. Briefly vortex the MPN supernatant and apply 10 ul from the appropriate supernatant onto the respective labeled spot (spot plate). Allow the spots to dry approx 20-30 min. Invert and incubate at 37 ° C O/N. Score as presumptive positive if a clear zone of lysis is visible in the spot and calculate the MPN from the number of positive (lysis zone) and negative (no lysis zone) spots. If necessary, verify the lysis zone by isolation of material from it and a repeated spot plate of this material.

Applications to detect somatic phages are the same as those for male-specific except the selection host is one that is F-minus (F-) such as *E. coli* strain CN-13 (ATCC 700609). For this host a 1% stock of antibiotic nalidixic acid is used in the media instead of strep/amp as used for *E. coli* Famp ([EPA Method 1601, 2001](#)). *E. coli* host C-3000 (ATCC 15597) selects for both F+ and somatic phages but findings from field studies reveal strain C-3000 recovers lower numbers of somatics and male-specific phages when compared to the sum of phages recovered by their respective host-specific strain. In a recent study, *E. coli* strain CB390 was effective at recovering the sum of both phage types ([Guzman et al., 2008](#)).

Coliphage antisera are obtained by inoculating experimental animals to elicit an immune response that results in polyclonal antibodies against the desired phage strain. For serotyping, isolated plaques are spotted (10 ul) onto an agar plate containing a specific antisera, either to MS2, GA, Q β , SP or FI. The agar plate contains the antisera and log phase host. Once the plaque is applied, the plate is incubated O/N at 37 ° C. Phage growth suppression on one of the plates, which contains homologous neutralizing antibodies to the phage group, is scored as positive. Serotyping is not 100% reliable and can produce ambiguous results ([Beekwilder et al., 1996](#)). For example, two isolates in the UNC collection were first serotyped to group II but later sequenced and genotyped to group I.

RNase sensitivity is determined by re-plating phage isolates on plates containing or lacking Ribonuclease A. On the plates with RNase, FRNA phages do not form lysis zones, however, FDNA phages will form zones of lysis.

FRNA Coliphage Detection Methods - Genotyping

Identification of male-specific phage by host selection does not discriminate between

FDNA and FRNA phages. An FRNA assay applicable for microbial source-tracking must first and foremost precisely identify FRNA. Genogroup-targeted methods solve two considerations, (i) FRNA selection and (ii) genotype-associated source tracking.

To distinguish the individual FRNA groups I, II, III, IV and a combination of groups I and II, III and IV, a molecular hybridization assay was developed. Genogroup-specific hybridization probes were designed using multiple alignment software to strains MS2, GA, Q β , and SP. Environmental isolates of FRNA phage as lysis zones on agar plates were adsorbed to a membrane, denatured to unfold RNA secondary structures and linked to the membrane using UV light. Digoxigenin (DIG-dUTP) labeled probes were incubated overnight with the fixed membranes and visualized with alkaline phosphatase-conjugated anti-DIG antibody. Assay development involved optimization of various hybridization solutions, selection of the most efficient membrane and denaturation solutions. To test the hybridization assay, 203 FRNA field isolates from sewage, oysters, surface waters and feces were first grouped by serotyping. Serotyping and hybridization classified 79 and 109 isolates, respectively, from surface waters, adult and piglet swine feces, treated and untreated sewage and oysters as genogroup II. Almost half of the piglet isolates, 12 out of 26, were type IV. Isolates from chickens were grouped almost equally into I and IV. Serotyping cross-reactivity was observed for 37 isolates neutralized by anti-sera GA and partial neutralization to anti-sera MS2. However, these 37 serotyped strains were eventually genotyped by hybridization to be in group II. Similar classification results were obtained by both serotyping and hybridization genotyping ([Hsu et al., 1995](#)).

A hybridization assay ([Beekwilder et al., 1996](#)) was developed similar to Hsu et al., ([1995](#)), but oligonucleotide probes targeted different regions of the genome. Probe to group I

was designed to MS2 nucleotide region 1248 (Hsu) and 1260 (Beekwilder), group II was designed to GA region 431 (Hsu) and 2100 (Beekwilder), group III was designed to Q β region 27 (Hsu) and 660 (Beekwilder) and group IV probe was designed to SP region 35 (Hsu) and 40 (Beekwilder). Probes were designed to each genogroup by Beekwilder and colleagues by alignment of 3-5 strains/group. If the completed sequenced genome was not available, they proceeded to sequence partial regions to provide adequate sequence representation for each genogroup. In addition, a probe A for *Levivirus* (groups I and II) and probe B for *Allolevivirus* (groups III and IV) was developed. To validate the hybridization probes, a combination of “blinded” but previously serotyped samples and field samples were analyzed. Approximately 78% of the samples were correctly identified by both the genogroup and genus-specific probes. Isolates collected from human impacted areas in the form of hospital waste and domestic wastewater identified 1 group I, 1 group II, 4 group III and 1 group IV. The two human feces samples had ambiguous classifications in that both samples were weakly positive for group I and positive for group IV but hybridized to both genus-specific probes, *Levivirus* and *Allolevivirus*. Twenty phages isolated from animal sources hybridized to groups I or IV in all cases except two. Of those two, one isolate was positive to groups I and II, and the second isolate (porcine slaughterhouse) was positive to group III (Beekwilder et al., 1996). These hybridization studies lend supporting evidence to the trends that genogroups II and III are associated with human waste and groups I and IV occur more often in animal sources.

Reverse-line blot is a nucleic hybridization-based assay designed to genotype both FRNA and FDNA coliphages. Using Clustal W 1.4 software, alignment of complete or partial sequences available in GenBank was the basis for primer and probe design. Six

cluster-specific oligonucleotide probes for hybridization were designed to FRNA strains, 1 group I probe for MS2-like, 1 probe for GA-like, 2 group III probes, Q β and M11, and 2 group IV probes, SP and FI-like. Three cluster-specific FDNA probes were also designed, M13, fd and CH and one generic FDNA consensus-specific probe, termed “con.” In order to initially divide FRNA from FDNA isolates, a generic RT-PCR assay was performed. Broadly reactive primer sets yield different PCR amplicon sizes for *Allolevivirus* vs. *Levivirus*. *Allolevivirus* was amplified by primers MJV82 forward, JV41 reverse and *Levivirus* was detected by primers MJV82 forward and JV81 reverse. Primer pair SL2 forward and SL3 reverse amplified FDNA phages. Assay validation began by collecting a total of 557 environmental samples. Phages were isolated by single or double-agar overlay plating, followed by RNase sensitivity testing and serotyped by anti-sera neutralization or genotyped by hybridization ([Hsu et al., 1995](#)). RT-PCR identified 100% of the FRNA and FDNA strains. Identified strains were used to validate the reverse-line blot hybridization (RLB), resulting in 98% agreement of the FRNA strains and 100% confirmation of the FDNA phages ([Vinjé et al., 2004](#)).

When comparing the basic hybridization assays, the RLB involves a two-step process since the RT-PCR step occurs prior to hybridization. Nonetheless, RLB typing had a higher predictability, 100% FDNA, 98% FRNA, when compared to 38% by serotyping ([Hsu et al., 1995](#)), 54% by genotyping ([Hsu et al., 1995](#)) and 78% by genotyping ([Beekwilder et al., 1996](#)).

Prior to 2007, 10 full-length or nearly full-length FRNA phage genomes were available at NCBI GenBank. In 2007, Kirs sequenced strain FI ([Kirs and Smith, 2007](#)), for a total of 11 FRNA genomes. Studies describing molecular detection of FRNA phages base their primer design on these 10 or 11 sequences and partial sequences available in GenBank.

A universal forward and reverse primer set was designed (Kelly Reynolds, Univ of AZ) to a consensus sequence in the replicase gene to detect all FRNA strains. A two-step reverse transcription polymerase chain reaction (RT-PCR) was performed. Prior to RT-PCR, environmental samples were column filtered to remove inhibitors. RT-PCR sensitivity was improved when the samples were column filtered and detection limits were 0.10 PFU of laboratory control MS2. RT-PCR amplified FRNA from 2 samples that were plaque-negative by soft agar overlay and was in agreement with the plaque-positive overlay methods (Rose et al., 1997). According to the authors, the primers amplified all four FRNA coliphage groups, but specific strains were not mentioned.

Phage MS2 is routinely used as a surrogate for pathogenic and environmental studies. Five sets of real-time primers and probes were designed to strain MS2. Two sets targeted the assembly gene, 1 for the coat region, 1 targeted the lysis gene and 1 primer and probe set targeted the replicase gene. Cross-reactivity of the real-time PCR was tested against non-targeted organisms, specifically pathogenic bacteria. MS2 primer sets did not cross-react with bacteria (O'Connell et al., 2006). Detection or cross-reactivity to other FRNA phages was not discussed.

Two independent real-time assays to detect each FRNA genogroup were developed for microbial source-tracking. Primer and probe sets were designed to the limited genomic NCBI GenBank database, by aligning 2 to 3 strains/genogroup and partial sequences. Purified RNA and a two-step QRT-PCR assay were common to both investigations. Kirs and Smith (2007) used a multiplexed assay whereas Ogorzaly and Gantzer (2006) analyzed each genogroup in separate vials. QRT-PCR primer and probe specificity were evaluated in the reaction containing an RNA cocktail of the strains MS2, GA, Q β , SP (Ogorzaly and

Gentzer, 2006) and MS2, GA, Q β , SP and FI (Kirs and Smith, 2007). Although the authors state their assay was template specific and lacked cross-hybridization, assay validation was limited to four or five FRNA strains. Raw sewage (Ogorzaly and Gentzer, 2006) and raw sewage and chicken litter (Kirs and Smith, 2007) samples were collected for QRT-PCR field validation. Ogorzaly and Gentzer detected 100% of groups I, II and 85% group III by QRT-PCR of double-agar overlay plaques. Group IV was not detected. Twenty plaques from sewage and 20 plaques from chicken litter were isolated, RNA purified and subjected to QRT-PCR (Kirs and Smith, 2007). Sewage isolates were group III and chicken stool isolates were group IV. Three genogroup-specific primer pairs were developed for a two-step RT-PCR assay and validated with strains MS2, GA and SP. Primer design for genogroups I, II and IV were based on GenBank genomic sequences for MS2, GA and SP, respectively. Phages isolated and enumerated from individual septic systems, poultry farm, municipal sewage and a background site were enumerated by SAL after filtration of the water sample. An MPN was also conducted. Purified phage RNA, from October 2004 environmental samples, tested positive for groups I and IV with RT-PCR and was in agreement with plaque-positive samples. The septic system tested negative with RT-PCR but positive by SAL and MPN. In January, 2005, 12 positive SAL and MPN samples tested positive for group I by RT-PCR. The May, 2005, sampling season had 3 out of 7 samples positive by SAL and MPN detection. In those 3 samples, RT-PCR identified group II upstream from the wastewater treatment plant and group I from the poultry farm-area of the lake. In contrast, SAL-positive samples downstream from the sewage treatment plant were negative by RT-PCR. Similar results were obtained in the August, 2005, sampling season as 4 samples were positive by SAL and MPN. Of those 4 samples, RT-PCR identified group II in the upstream

wastewater treatment plant sample but the 3 remaining samples taken downstream of the sewage plant, upstream of a different sewage plant and lake waters near the poultry farm were negative by RT-PCR. The authors concluded FRNA indicators were not useful to distinguish between human and non-human sources and suggested the presence of either somatic or FDNA phages in their positive samples ([Dryden et al., 2006](#)). Limitations to their study design are as follows. Primers were designed to three individual strains and not an alignment of multiple strains per genogroup. These primers would not detect all four genogroups or environmental strains as reflected in the results. A group III primer set was not designed and yet, the authors concluded FRNA could not distinguish between human and non-human sources. SAL and MPN host selection was *E. coli* strain C-3000 (ATCC 15597), a host that selects for both somatic and male-specific phages.

An antibody-based agglutination assay, termed “latex agglutination”, was developed to rapidly detect, <24 hr, genogroups of FRNA coliphages. Environmental strains were collected from bird droppings, shellfish and water bodies from diverse geographical locations across the USA. A two-step enrichment protocol was modified to a culture time of 180 min based on preliminary sampling and phage measurement at 0, 30, 60, 90, 120, 180 and 360 min on TSA plates containing *E. coli* host F_{AMP}. Male-specific plaques were confirmed as FDNA or FRNA by RNase sensitivity, FRNA genera were determined by broadly reactive RT-PCR and preliminary genogrouping by RLB ([Vinjé et al., 2004](#)). Serotyping and sometimes even RLB genotyping, at times, yield ambiguous results. To address this ambiguity, an RT-PCR assay to the capsid region with primers DL10 forward and DL11 reverse and sequencing of the PCR amplicon was conducted. To develop the coliphage latex

agglutination and typing (CLAT) assay, polyclonal antibodies were first generated against MS2, GA, Q β , SP and FI. Phage strain-specific antisera was bound to polystyrene particles and used in the agglutination step of the CLAT. Coliphage enriched samples showed agglutination within 30-60 sec when 2.5 ul enriched phage cultures were mixed with 2.5 ul strain-specific antibody particle on an agglutination card. Out of 192 FRNA field isolates, CLAT correctly identified 185 (96%) and RLB identified 92%. When the two methods were compared, some ambiguity was noted in that CLAT identified more group II and RLB identified more groups III and IV. Twenty-four strains yielded inconsistent typing. Sequencing of the capsid region and phylogenetic clustering of these 24 strains yielded 19 group I and 5 group II. CLAT clustered 17 of the 19 group I strains as I and II, but matched 100% of the 5 sequenced group II isolates. A total of 164 FDNA isolates were also identified by CLAT at a rate of 97.7% ([Love and Sobsey, 2007](#)).

FRNA Relationship to Enteric Viruses and as Predictors of Health Risk

An important attribute of an ideal indicator is the relationship between the indicator density in polluted waters to human-health risks. Few epidemiology studies exists correlating health risks with F+ coliphage densities. A European cohort study ([Lee et al., 1997](#)) showed a statistically significant relative risk (RR) with increased F+ coliphage exposures. However, a threshold value was not extrapolated and the slight increased RR (2.6-2.8) is minimal compared to the large coliphage density range of 26 - 32 and 69-308 PFU/10 ml, respectively.

A recent California beach study suggested an association between F+ coliphage densities (both FRNA and FDNA combined) and gastrointestinal illness rates, nausea, cough

and fever. The authors noted that when F+ phages were detected, a low number of beachgoers were exposed (Colford et al., 2007).

Meta-analysis of epidemiological studies revealed no correlation between bacteriophage in marine waters to GI illness, whereas the freshwater studies reported an elevated GI risk with elevated bacteriophage exposure (Wade et al., 2003). The meta-analysis did not clarify which type of bacteriophage showed associations with human health risks.

Six wastewater treatment facilities from FL, AZ and CA producing and distributing reclaimed water were monitored for indicator and pathogen load. Although reclaimed water is routinely assessed for fecal or total coliforms, the degree of microbial indicator and pathogen removal has not been evaluated. Results using male-specific coliphages did not show a correlation with enteric viral load. However, the coliphage predicted an absence of enteric viruses at levels less than 10 coliphage/100 ml (Rose et al., 2004).

Adenovirus presence was statistically correlated to FRNA ($r = 0.99$) in a brackish (salinity from 9-34 ppt) but no correlation to FDNA phages were observed in coastal waters (Jiang et al., 2001).

Future Applications

Molecular detection methods, i.e., PCR, real-time PCR and microarrays, allow a more timely assessment of the microbial quality of recreational waters. A prospective study at two Great Lakes beaches using real-time PCR detection found that enterococci was statistically associated with increased gastro-intestinal (GI) illness at both beach sites. A strong positive trend was noted with the presence of *Bacteroides* at one of the beaches (Wade et al., 2006).

Microarrays have been constructed with genomic DNA purified from raw wastewaters (Lee and el., 2006) and 16S rRNA and cpn60 genes extracted from several specific pathogens (Maynard et al., 2005). Primers and oligonucleotide microarray probes were designed from sequences derived from specific pathogenic bacteria strains. Validation of the microarrays generated a positive hybridization signal in raw sewage samples for the *E. coli* gene *uidA* (Lee et al., 2006).

Commercially-available field PCR instruments could potentially be applicable to molecular detection of FRNA genogroups. For example, a hand-held fluorogenic real-time PCR instrument, the Advanced Nucleic Acid Analyzer (ANAA), was used to detect bacteria spores and MS2 virions. The microbes were analyzed using a micro-chip in the ANAA and a positive signal was viewed within 18-26 min (Belgrader et al., 1998).

Portable real-time thermocyclers (Cepheid, Inc) and nucleic acid sequence based amplification (NASBA) instruments (BioMereux, Inc) can be transported to provide on-the-spot analysis of environmental samples. NASBA, unlike PCR, does not require a thermocycler. NASBA relies on an isothermal process (41 °C), three RNA-associated enzymes and a molecular beacon. Single-stranded RNA is generated in a single-tube, emitting fluorescence by the molecular beacon upon hybridization with ssRNA and a target-specific oligonucleotide. NASBA technology targets RNA viruses and would be applicable as a portable instrument to detect FRNA coliphages.

Summary and Conclusion

Microbial source-tracking is defined as a group of analytical protocols, typically microbial applications, which are used to ascertain or regulate the source of fecal input into a water body. Use of a potential microbial source-tracking “toolbox” is a complex process

whereby decisions must be made to select and validate the selection of such tools. For example, (1) define the problem (nearby source discharge or runoff, history of monitoring data, weather patterns, hydrology, etc), (2) formulate objectives as to the suspected source and category (human vs. non-human), (3) presence/absence vs. quantification of loading values (4) consider if the application linked to regulatory values for microbial indicator concentration or if fecal presence or absence is an acceptable result for decision making, (5) consider if fecal presence is linked to public health risk, (6) consider if there are legal ramifications, (7) select the most appropriate source-tracking protocol (8) define a sampling strategy or study design, (9) define a method of data collection and quality control, (10) define data analyses and (11) have a plan for data interpretation ([Stoeckel, 2005](#)).

Currently, the toolbox consists of DNA fingerprinting, antibiotic resistance, ribotyping, pulse-field electrophoresis and carbon utilization profiles of *E. coli* and/or enterococci (library-dependent), host-specific *Bacteroides* and *Prevotella* bacteria markers (library-independent) and serotyping or genotyping of FRNA or FDNA coliphages (library-independent). The “library” per se, is a collection of bacterial or viral isolates from which the source of collection is known as well as the fingerprint, marker, genotype or serotype. One aspect of method selection depends upon the analytical question, library-dependent (epidemiological matching or clustering) vs. library-independent (source could be traced in any water body type or geographical location).

FRNA source-specificity displayed by the four genogroups render FRNA phages applicable for microbial source-tracking. Groups I and IV are typically associated with animal wastes whereas groups II and III generally occur from human waste ([Havelaar et al., 1986](#); [Schaper et al., 2002](#)). FRNA library collections began with known sources, i.e.,

domestic sewage/drainage or hospital waste sites, cattle, swine and poultry waste lagoons or litters, animal-specific fecal voids, and/or animal intestinal content and, following FRNA purification, these sources were further defined by serotyping or genotyping (Furuse et al., 1978; Miyake et al., 1971; Furuse et al., 1975; Miyake et al., 1973; Osawa et al., 1981; Schaper et al., 2002; Beekwilder et al., 1996; Hsu et al., 1995; Calci et al., 1998; Cole et al., 2003; Long et al., 2005; Sundram et al., 2006; Stewart-Pullaro et al., 2006; Kirs and Smith, 2007). Building upon trends in FRNA source specificity, investigators have isolated environmental phages, purified them and genotyped/serotyped as a strategy to classify fecal contamination (Dryden et al., 2006; Love and Sobsey, 2007; Stewart-Pullaro et al., 2006; Vinjé et al., 2004; Stewart et al., 2006; Yee et al., 2006; Sundram et al., 2006; Brion et al., 2002; Sobsey et al., 2006).

Types and trends of known FRNA host specificity were reported to indicate human source from sewage-collected FRNA as follows: genogroup I 38% , 15%, 12%, 11%, 2%; genogroup II 50%, 38% , 21% , 15%, 14%; genogroup III 100%, 73%, 58%, 57%, 50 % , 46%, 15%, 6%; and genogroup IV 1%, 5% IV and animal sources: genogroup I 98%, 70%, 51%, 50%, 50%, 32% 30%; genogroup II 15%, 5%; genogroup III 22%, 19%, 18%; and genogroup IV 100%, 50%, 50%, 46%, 6%, 3% (Aoi et al., 1972; Miyake et al., 1973; Brion et al., 2002; Miyake et al., 1971; Cole et al., 2003; Hsu et al., 1995; Kirs and Smith, 2007). Although the data from these studies is not comprehensive or exhaustive, a host-specific FRNA genogroup trend exists.

In this dissertation, a library-independent method to differentiate between sources of human and non-human fecal pollution was developed. Initially, nineteen FRNA strains were sequenced and compared to the eleven FRNA full-length sequences available in the National

Center for Biotechnology Information (NCBI) genetic database (GenBank) for a total of 30 FRNA strains. FRNA phages were collected from water, sewage, and various animals representative of diverse geographical locations (Table 7.1). The field-collected FRNA strains and prototype strains were represented by phages isolated from the United States, Japan, Europe, Brazil, Taiwan and/or Mexico (Table 7.1). FRNA sequences generated in this study tripled the genetic information currently available in the national genetic database for the *Leviviridae*. Based on the intensive sequencing effort, a robust, one-step reverse transcription polymerase chain reaction (RT-PCR) was designed to distinguish the four FRNA coliphage groups (I, II, III, IV). In the more immediate future this data can be applied to methods using FRNA coliphages as a fecal and viral indicator and as a source-tracking tool.

To conclusively apply the “FRNA group II, III human fecal pollution” association for recreational or other surface water regulatory purposes, one must caution that one or two or a few isolates does not confirm a human-impacted area. Perhaps a more plausible application would be to propose an absolute percentage or value of group II and III isolates from a pre-determined FRNA sample size detected from an area where wastewater discharges and/or other source delineations have been examined.

References

- Ackermann HW. 2006. Classification of bacteriophages. In: *The Bacteriophages* by S T Abedon. R L Calendar, ed. Oxford University Press. pp:8-16.
- American Water Works Association. 2004. Field testing of USEPA methods 1601 and 1602 for coliphage in groundwater. Prepared by MR Karim, MW LeChevallier, M Abaszadegan, A Alum, J Sobrinho and J Rosen. AWWA Research Foundation, Denver, CO.
- Armon R and Y Kott. 1995. Distribution comparison between coliphages and phages of anaerobic bacteria (*Bacteroides fragilis*) in water sources, and their reliability as fecal pollution indicators in drinking water. *Wat Sci Tech*, 31(5):215-222.
- Ashbolt NJ, WOK Grabow and M Snozzi. 2001. Indicators of microbial water quality. In: *World Health Organization, Water Quality: Guidelines, Standards and Health*. L Fewtrell and J Bartram, eds. IWA Publishing, London, UK. pp:289-316.
- Aoi T, K Furuse, T Shiba and I Watanabe. 1972. Isolation and grouping of RNA phages III. A survey in Arabia, India and Thailand. *J Keio Med Soc*, 49:361-368.
- Bae J and KJ Schwab. 2008. Evaluation of murine norovirus, feline calicivirus, poliovirus, and MS2 as surrogates for human norovirus in a model of viral persistence in surface water and groundwater. *Appl Environ Microbiol*, 74(2):477-484.
- Beekwilder J, R Nieuwenhuizen, AH Havelaar and J van Duin. 1996. An oligonucleotide hybridization assay for the identification and enumeration of F-specific RNA phages in surface water. *J Appl Bacteriol*, 80:179-186.
- Belgrader P, W Bennett, D Hadley, G Long, R Mariella, Jr, F Milanovich, S Nasarabadi, W Nelson, J Richards and P Stratton. 1998. Rapid pathogen detection using a microchip PCR array instrument. *Clin Chem*, 44(10):2191-2194.
- Bernhard AE, T Goyard, MT Simonich and KG Field. 2003. Application of a rapid method for identifying fecal pollution sources in a multi-use estuary. *Wat Res* 37(4):909-913.
- Bitton, G. 2005. Microbial indicators of fecal contamination: application to microbial source tracking. Report submitted to the Florida Stormwater Association, Tallahassee, FL. June, 2005.
- Borrego JJ, MA Morinigo, A de Vicente, R Cornax and P Romero. 1987. Coliphages as an indicator of faecal pollution in water. Its relationship with indicator and pathogenic microorganisms. *Wat Res*, 21(12):1473-1480.
- Bradley DE. 1964. The structure of some bacteriophages associated with male strains of

- Escherichia coli*. J Gen Microbiol, 35:471-482.
- Brion GM, JS Meschke and MD Sobsey. 2002. F-specific RNA coliphages: occurrence, types, and survival in natural waters. Water Res, 36:2419-2425.
- Buchen-Osmond, C. (Ed.) 2004. Levivirus. In ICTVdB - *The Universal Virus Database*, version 3. ICTVdB Management, Columbia University. New York, NY.
- Burkhardt III W, KR Calci, WD Watkins, SR Rippey and SJ Chirtel. 2000. Inactivation of indicator microorganisms in estuarine waters. Wat Res, 34(8):2207-2214.
- Clark AJ. 1963. Genetic analysis of a “double male” strain of *Escherichia coli* K-12. Genetics 48:105-120.
- Calci KR, W Burkhardt III, WD Watkins and SR Rippey. 1998. Occurrence of male-specific bacteriophage in feral and domestic animal wastes, human feces, and human-associated wastewaters. Appl Environ Microbiol, 64(12):5027-5029.
- Cole, D, SC Long and MD Sobsey. 2003. Evaluation of F+ RNA and DNA coliphages as source-specific indicators of fecal contamination in surface waters. Appl Environ Microbiol, 69(11):6507-6514.
- Colford JM, Jr., TJ Wade, KC Schiff, CC Wright, JF Griffith, SK Sandhu, S Burns, M Sobsey, G Lovelace and SB Weisberg. 2007. Water quality indicators and the risk of illness at beaches with nonpoint sources of fecal contamination. Epidemiology 18(1):27-35.
- Cornax R, MA Morinigo, IG Paez, MA Munoz and JJ Borrego. 1990. Application of direct plaque assay for detection and enumeration of bacteriophages of *Bacteroides fragilis* from contaminated water samples. Appl Environ Microbiol, 56(10):3170-3173.
- Daehnel K, R Harris, L Maddera, and P Silverman. 2005. Fluorescence assays for F-pili and their application. Microbiology, 151:3541-3548.
- Dryden SK, B Ramaswami, Z Yuan, DE Giammar, LT Angenent. 2006. A rapid reverse transcription-PCR assay for F+ RNA coliphages to trace fecal pollution in Table Rock Lake on the Arkansas-Missouri border. Water Res, 40:3719-3724.
- Duckworth DH. 1987. History and basic properties of bacterial viruses. In: *Phage Ecology*. SM Goyal, CP Gerba and G Britton, eds. John Wiley & Sons, Inc., NY. pp: 1-43.
- Fiers W, R Contreras, F Duerinck, G Haegeman, D Iserentant, J Merregaert, W Min Jou, F Molemans, A Raeymaekers, A Van den Berghe, G Volckaert and M Ysebaert. 1976.

- Complete nucleotide sequence of bacteriophage MS2 RNA: primary and secondary structure of the replicase gene. *Nature*, 260:500-507.
- Furuse K. 1987. Distribution of coliphages in the environment: general considerations. In: *Phage Ecology*. SM Goyal, CP Gerba and G Britton, eds. John Wiley & Sons, Inc., NY. pp 87-124.
- Furuse K, A Ando and I Watanabe. 1975. Isolation and grouping of RNA phages VII. A survey in Peru, Bolivia, Mexico, Kuwait, France, Australia and the United States of America. *J Keio Med Soc*, 52:353-361. (Japanese)
- Furuse K, T Sakurai, A Hirashima, M Katsuki, A Ando, and I Watanabe. 1978. Distribution of ribonucleic acid coliphages in South and East Asia. *Appl Environ Microbiol*, 35(6):995-1002.
- Gantzer C, J Henry and L Schwartzbrod. 2002. *Bacteroides fragilis* and *Escherichia coli* bacteriophages in human feces. *Int J Hyg Environ Health*, 205:325-328.
- Gerba, CP. 2006. Bacteriophage as pollution indicators. In: *The Bacteriophages* by S T Abedon. R L Calendar, ed. Oxford University Press. pp: 695-701.
- Gino E, J Starosvetsky and R Armon. 2007. Bacteriophage ecology in a small community sewer system related to their indicative role in sewage pollution of drinking water. *Environ Microbiol*, 9(10):2407-2416.
- Guzman C, L Moce-Llivina, F Lucena and J Jofre. 2008. Evaluation of *Escherichia coli* host strain CB390 for simultaneous detection of somatic and F-specific coliphages. *Appl Environ Microbiol*, 74(2):531-534.
- Goyal SM, CP Gerba and G Britton, eds. 1987. *Phage Ecology*. John Wiley and Sons, Inc. NYC, NY.
- Havelaar AH, and WM Hogeboom. 1984. A method for the enumeration of male-specific bacteriophages in sewage. *J Appl Bacteriol*, 56:439-447.
- Havelaar AH, WM Hogeboom, and R Pot. 1984. F-specific RNA bacteriophages in sewage: methodology and occurrence. *Wat Sci Tech*, 17:645-655.
- Havelaar AH and WM Pot-Hogeboom. 1988. F-specific RNA-bacteriophages as model viruses in water hygiene: ecological aspects. *Water Sci Tech*, 20:399-407.
- Havelaar AH, M van Olphen and YC Drost. 1993. F-specific RNA bacteriophages are adequate model organisms for enteric viruses in fresh water. *Appl Environ Microbiol*, 59(9):2956-2962.

- Hsu FC, YS Carol Shieh, J van Duin, MJ Beekwilder and MD Sobsey. 1995. Genotyping male-specific RNA coliphages by hybridization with oligonucleotide probes. *Appl Environ Microbiol*, 61(11):3960-3966.
- Jiang S, R Noble and W Chu. 2001. Human adenoviruses and coliphages in urban runoff-impacted coastal waters of southern California. *Appl Environ Micro* 76(1):179-184.
- Kirs M and DC Smith. 2007. Multiplex quantitative real-time reverse transcriptase PCR for F+-specific RNA coliphages: a method for use in microbial source tracking. *Appl Environ Microbiol*, 73(3):808-814.
- Lee JV, SR Dawson, S Ward, SB Surman and KR Neal. 1997. Bacteriophages are a better indicator of illness rates than bacteria amongst users of a white water course fed by a lowland river. *Wat Sci Tech* 35(11-12):165-170.
- Lee DY, K Shannon and LA Beaudette. 2006. Detection of bacterial pathogens in municipal wastewater using an oligonucleotide microarray and real-time quantitative PCR. *J Microbiol Meth*, 65:453-467.
- Leclerc H, S Edberg, V Pierzo and JM Delattre. 2000. Bacteriophages as indicators of enteric viruses and public health risk in groundwaters. A review. *J Appl Microbiol*, 88:5-21.
- Loeb T, and ND Zinder. 1961. A bacteriophage containing RNA. *Proc Nat Acad Sci*, 47:282-289.
- Long SC, SS El-Khoury, SJG Oudejans, MD Sobsey and J Vinjé. 2005. Assessment of sources and diversity of male-specific coliphages for source tracking. *Environ Eng Sci*, 22(3):367-377.
- Long SC, and MD Sobsey. 2004. A comparison of the survival of F+ RNA and F+ DNA coliphages in lake water microcosms. *J Water Health*, 2(1):15-22.
- Love DC and MD Sobsey. 2007. Simple and rapid F+ coliphage culture, latex agglutination, and typing assay to detect and source track fecal contamination. *Appl Environ Microbiol*, 73(13):4110-4118.
- Maynard C, F Berthiaume, K Lemarchand, J Harel, P Payment, P Bayardelle, L Masson and R Brousseau. 2005. Waterborne pathogen detection by use of oligonucleotide-based microarrays. *Appl Environ Microbiol*, 71(12):8548-8557.
- Meschke JS and MD Sobsey. 2003. Comparative reduction of Norwalk virus, poliovirus type 1, F+RNA coliphage MS2 and *Escherichia coli* in miniature soil columns. *Water Sci Tech* 47(3):85-90.

- Miyake T, K Furuse, T Shiba, T Aoi, T Sakurai and I Watanabe. 1971. Isolation and grouping of RNA phages in Taiwan. *J Keio Med Soc*, 48:25-34. (Japanese).
- Miyake T, K Furuse, T Shiba, T Aoi, T Sakurai and I Watanabe. 1973. Isolation and grouping of RNA phages. II. A survey in Brazil. *J Keio Med Soc*, 50:353-362. (Japanese).
- Miyake T, I Haruna, T Shiba, YH Itoh, K Yamane, and I Watanabe. 1971. Grouping of RNA phages based on the template specificity of their RNA replicases. *Proc Nat Acad Sci*, 68(9):2022-2024.
- Miyake T, T Shiba, T Sakurai, and I Watanabe. 1969. Isolation and properties of two new RNA phages SP and FI. *Japan J Micro*, 13(4), 375-382.
- Nelson D. 2004. Phage taxonomy: we agree to disagree. *J Bacteriol*, 186(21):7129-7031.
- O'Connell KP, JR Bucher, PE Anderson, CJ Cao, AS Khan, MV Gostomski and JJ Valdes. 2006. Real-time fluorogenic reverse transcription-PCR assays for detection of bacteriophage MS2. *Appl Environ Microbiol*, 72(1):478-483.
- Ogorzaly L and C Gantzer. 2006. Development of real-time RT-PCR methods for specific detection of F-specific RNA bacteriophage genogroups: application to urban raw wastewater. *J Virol Meth*, 138:131-139.
- Osawa S, K Furuse, MS Choi, A Ando, T Sakurai, and I Watanabe. 1981. Distribution of ribonucleic acid coliphages in Korea. *Appl Environ Microbiol*, 41(4):909-911.
- Osawa S, K Furuse, and I Watanabe. 1981. Distribution of ribonucleic acid coliphages in animals. *Appl Environ Microbiol*, 41(1):164-168.
- Paranchych, W. 1975. Attachment, ejection and penetration stages of the RNA phage infectious process. In: ND Zinder (ed). *RNA Phages*. Cold Spring Harbor Laboratory. pp. 85-111.
- Puig A, N Queralt, J Jofre and R Araujo. 1999. Diversity of *Bacteroides fragilis* strains in their capacity to recover phages from human and animal wastes from fecally polluted wastewater. *Appl Environ Microbiol*, 65(4):1772-1776.
- Rohwer F and R Edwards. 2002. The phage proteomic tree: a genome-based taxonomy for phage. *J Bacteriol*, 184(16):4529-4535.
- Rose JB, X Zhou, DW Griffin and JH Paul. 1997. Comparison of PCR and plaque assay for detection and enumeration of coliphage in polluted marine waters. *Appl Environ Microbiol*, 63(11):4564-4566.
- Rose JB, SR Farrah, VJ Harwood, AD Levine, J Lukasik, P Menendez and TR Scott. 2004.

- Reduction of pathogens, indicator bacteria, and alternate indicators by wastewater treatment and reclamation processes. Water Environment Research Foundation (WERF). Water for Reuse; Final Report. Co-published by IWA.
- Sakurai T, T Miyake, T Shiba and I Watanabe. 1988. Isolation of a possible fourth group of RNA phage. *Japan J Microbiol*, 12(4):544-546.
- Schaper M, J Jofre, M Uys, and WOK Grabow. 2002. Distribution of genotypes of F-specific RNA bacteriophages in human and non-human sources of faecal pollution in South Africa and Spain. *J Appl Microbiol*, 92:657-667.
- Sobsey. Coliphage tracking to identify sources of fecal contamination. <ftp://ftp.sccwrp.org/>
- Stewart JR, J Vinjé, SJG Oudejans, GI Scott and MD Sobsey. 2006. Sequence variation among group III F-specific RNA coliphages from water samples and swine lagoons. *Appl Environ Microbiol*, 72(2):1226-1230.
- Stewart-Pullaro, J, JW Daugomah, DE Chestnut, DA Graves, MD Sobsey and GI Scott. 2006. F+RNA coliphage typing for microbial source tracking in surface waters. *J Appl Microbiol*, 101:1015-1026.
- Stoeckel DM. 2005. Selection and application of microbial source tracking tools for water-quality investigations. In: *Techniques and Methods Book 2*, Chapter 3A.
- Sundram A, N Jumanial, and MM Ehlers. 2006. Genotyping of F-RNA coliphages isolated from wastewater and river water samples. *Water SA*, 32(1):65-70.
- Tartera C and J Jofre. 1987. Bacteriophage active against *Bacteroides fragilis* in sewage-polluted waters. *Appl Environ Microbiol*, 53:1632-1637.
- Tartera C, F Lucena and J Jofre. 1989. Human origin of *Bacteroides fragilis* bacteriophage present in the environment. *Appl Environ Microbiol*, 55:2696-2701.
- van Duin, J., 1998. The single-stranded RNA bacteriophages. in: Fraenkel Conrat, H., Wagner, R.R. (Eds). *The Bacteriophages*. Series *The Viruses*. Plenum Press, NY. pp. 117-167.
- Vinjé J, SJG Oudejans, JR Stewart, MD Sobsey and SC Long. 2004. Molecular detection and genotyping of male-specific coliphages by reverse transcription-PCR and reverse line blot hybridization. *Appl Environ Microbiol*, 70(10):5996-6004.
- Wade TJ, RL Calderon, E Sams, M Beach, KP Brenner, AH Williams and AP Dufour. 2006. Rapidly measured indicators of recreational water quality are predictive of

- swimming-associated gastrointestinal illness. *Environ Health Persp*, 114(1):21-28.
- Wade TJ, N Pai, JNS Eisenberg and JM Colford Jr. 2003. Do U.S. Environmental Protection Agency water quality guidelines for recreational waters prevent gastrointestinal illness? A systematic review and meta-analysis. *Environ Health Persp* 111(8):1102-1109.
- Yee, SYF, NY Fong, GT Fong, OJ Tak, GT Hui and YS Ming. 2006. Male-specific RNA coliphages detected by plaque assay and RT-PCR in tropical river waters and animal fecal matter. *Int J Environ Health Res*, 16(1):59-68.
- Zinder ND, RC Valentine, M Roger, and W Stoeckenius. 1963. F1, a rod-shaped male-specific bacteriophage that contains DNA. *Virology*, 20:638-640.
- Zinder ND. 1965. RNA phages. *Annu Rev Microbiol*, 19:455-473.

V. Gene Mapping and Phylogenetic Analysis of the Complete Genome of 30 ssRNA Male-Specific Coliphages of the Family *Leviviridae*

Abstract

An international collection of male-specific ssRNA (FRNA) coliphages comprising the *Leviviridae* family exists but the genetic diversity of these strains is poorly characterized. FRNA coliphages belonging to the family *Leviviridae* are genetically divided into two genera (*Levivirus* and *Allolevivirus*) which can be further divided into four different genogroups (I, II, III and IV).

To better characterize this virus family and its genetic genogroups, strains were collected from the United States, Japan, Europe, Brazil, Taiwan and/or Mexico. The complete genomic sequences of 19 new FRNA strains (10 *Levivirus* and 9 *Allolevivirus*) from diverse sources were determined and compared to the eleven known genome sequences available in GenBank, for a total of thirty FRNA genomes. Phylogenetic analyses demonstrated all strains clustered into two genera, *Levivirus* and *Allolevivirus*, and in four distinct genogroups, I, II, III and IV. Out of ten genogroup I strains, nine strains shared nucleotide sequence similarities ranging from 91.68% - 99% whereas genogroup I strain fr shared a 75.27-77.65% sequence identity with the other genogroup I phages. Genogroup II FRNA strains shared 83.30-93.84% nucleotide similarity. *Allolevivirus* strains shared a nucleotide similarity range of 69.77-95.69% and 74.90-95.03% for genogroups III and IV, respectively. An approximate 50% nucleotide sequence identity was shared between *Levivirus* groups I and II and between *Allolevivirus* groups III and IV. For all strains

genomic full-length nucleotide and individual protein phylogenetic trees were compared. Genogroup II lysis protein tree formed a unique branch that was not observed in the full-length nucleotide tree. Thus, both full-length nucleotide and individual proteins need to be evaluated when genotyping or phylogenetically clustering these FRNA coliphages. Data for amino acid composition, nucleotide similarities and replicase catalytic domain location contributed to phylogenetic branches or strain subclusters. Eight nucleotides at the 3' termini clearly distinguished between the *Levivirus*, 5' ACCACCCA 3' from the *Allolevivirus* 5' TCCTCCCA 3' genera. This evidence suggests that the sequence data is valid.

Introduction

Male-specific RNA (FRNA) coliphages are single-strand RNA (ssRNA) viruses possessing a positive sense genome ranging from 3.8 to 4.2 kb in size enclosed by a non-enveloped 26 nm icosahedral-shaped capsid ([Buchen-Osmond, 2003](#)). The bacterial host is restricted to the gram-negative bacteria of the genera *Escherichia*, *Pseudomonas*, *Caulobacter*, *Salmonella*, and *Vibrio* ([Loeb and Zinder, 1961](#)). For successful infection, the host must possess a fertility (F) sex-pilus, coded on the F-plasmid of *E. coli* ([Paranchych, 1975](#)), as infection occurs by attachment to this receptor site ([Crawford and Gesteland, 1964](#)). As the sex-pili are only expressed at temperatures greater than 30 ° C ([Grabow, 2001](#)), phage replication is restricted to environments such as the intestinal tract of warm-blooded animals. The inability to replicate outside the gut, environmental stability, high numbers in sewage and a strong correlation with water-borne pathogens are some of the properties which have

made FRNA phages attractive candidates as indicators of fecal contamination in water (Havelaar et al., 1993).

FRNA phages belong to the family *Leviviridae* and can be further subdivided into two genera (*Levivirus* and *Allolevivirus*). *Levivirus* are subdivided into genogroups I and II and *Allolevivirus* are subdivided into genogroups III and IV. Historically, these subgroups were based on serological properties (Sundram et al., 2006), sedimentation, density and molecular weight (van Duin, 1988). Recently, genomic data has provided an additional subgrouping tool (Stewart et al., 2006).

Based on a limited number of complete sequences four genes could be identified (reviewed by Bollback & Huelsenbeck, 2001). These genes code for an assembly or maturation protein, capsid protein, lysis protein and replicase protein in the *Leviviruses* whereas in *Alloleviviruses*, the lysis protein is replaced by a read-through protein. Each levivirus virion contains one molecule of positive sense ssRNA, 180 copies of the capsid or coat protein, one copy of the maturation protein and, in alloleviviruses, approximately 15 copies of the read-through protein (Weber & Konigsberg, 1975; van Duin, 2000; van Duin and Tsareva, 2006).

A few complete nucleotide sequences of *Leviviridae* strains are known, including prototype strains MS2, GA, Q β and FI, as only 11 FRNA phages have been fully sequenced over the past few decades. With rapid molecular advances, sequencing is now more affordable and feasible.

Purpose

- Generate a nucleotide (nt) sequence database of complete genomic sequences of representative strains and environmentally isolated strains for all four genogroups of FRNA coliphages.
-
- Determine phylogenetic profiles, nucleotide sequence similarity, amino acid composition, Open Reading Frame (ORF) positions and subsequent gene locations for a total of 30 FRNA sequences.
-

Approach

-
- Sequence nineteen FRNA strains and compare to eleven full-length genome sequences available in GenBank (GenBank/EMBL/DDBJ).
-
- Determine Open Reading Frames by locating Shine-Dalgarno regions and start codons to map each gene.
-
- Translate nucleotide sequences into amino acid compositions for each protein.
-
- Compare nucleotide and amino acid percent similarities among each genogroup.
-
- Determine and compare each protein family, protein motifs and domains by use of bioinformatics tools.
-
- Compare phylogenetic clustering for each full-length nucleotide genome and individual proteins in all genogroups.

Materials and Methods

FRNA Coliphage Strains and RNA Extraction

FRNA strains used in this study include prototype strains MS2 (genogroup I), GA (genogroup II), Q β (genogroup III), FI (genogroup IV) and SP (genogroup IV) were kindly provided by Dr. K. Furuse (Toaki University, Japan) and FRNA strains ST4, TW18, VK and BZ1 were a gift from Dr. J. van Duin (Leiden University, The Netherlands). Field-collected strains BR1, BR8 and BR12 were a gift from Brian Robinson (NOAA, Charleston, SC) and prototype strain fr was provided by Dr. A. Boehm (Stanford University, Stanford, CA). Strain R17 was purchased from Felix D'Herelle Reference Centre for Bacterial Viruses, Universite Laval, Quebec, Canada. In addition, field strains isolated from wastewater, surface waters, swine lagoons and chicken litter were analyzed in this study ([Table 7.1](#)). Preliminary subgrouping of phages was previously determined by reverse line-blot hybridization ([Vinjé et al., 2004](#)).

Each strain was plaque purified and further enriched using *Escherichia coli* HS(pFamp)R as host ([Vinjé et al., 2004](#)). Single plaques were enriched overnight in Tryptic Soy Broth (TSB) supplemented with streptomycin-sulfate (15 mg/L) and ampicillin (15 mg/L). Cultures were centrifuged (3,220 x g for 10 min) to pellet host cells and debris and the supernatant was chloroform extracted (1:1 V:V). Approximately 1-2 ml aliquots of the purified supernatant were frozen at -75 °C.

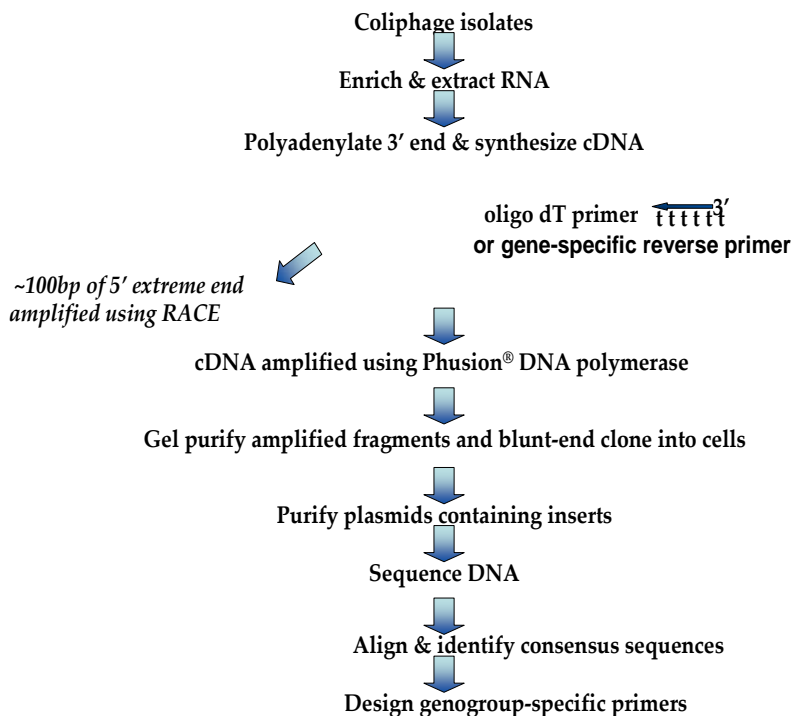
Coliphage titers were determined using a single agar layer procedure (SAL) ([US EPA Method 1602, 2001](#)). The procedure was as follows. A 1 ml of overnight *E. coli* Famp host culture was transferred into 50 ml trypticase soy broth with streptomycin and ampicillin

(TSB/strep/amp) and grown 4 hr to log phase. A 150 ml volume of trypticase soy agar, TSA (4.5 g TSB and 1.2 g agar), was autoclaved and placed into a water bath (47-55 ° C). A 300 ul aliquot of 500X strep/amp was added into the cooled 150 ml TSA. Serial 10-fold dilutions of the purified virus were prepared. For 10-fold stock dilution, 100 ul of stock was transferred into 900 ul phosphate buffered saline (PBS), mixed and this process repeated to a serial dilution 10^{-15} . To sterile 15 ml plastic tubes, labeled -1 to -15, 1 ml of the 4 hr *E. coli*, 100 ul of the appropriate virus dilution and 10 ml TSA were added. After quick manual mixing the tube contents were poured into labeled 20mm petri plates which were then cooled, allowed to dry, inverted and incubated overnight at 37 ° C. The coliphage titer was determined by counting the plates having a minimum of 5 to a maximum of 20 plaques per plate. Titer concentration was reported as PFU/ml.

Coliphage RNA was extracted from the purified viral supernatant using a QIAamp viral RNA mini kit (Qiagen, Valencia, CA) as follows. A 200 ul volume of purified virus stock was added into the 25 ul protease tube. Into the same tube, 200 ul of carrier RNA/Buffer AL mixture was added and the combined solutions were vortex mixed, incubated 56 ° C for 15 minute and clarified by a 1 minute centrifugation. To the same tube, 250 ul ethanol (EtOH) was added, vortex mixed and incubated 5 min at room temperature (RT). The lysate was applied into the QIAamp spin column and centrifuged 3-4 min at 6000 xg (8000 rpm) and the contents of the collection tube was discarded. To the column, 500 ul of Buffer AW2 was added, centrifuged 1 min and the contents of the collection tube were discarded. To the column, 500 ul EtOH was added, centrifuged 1 min and collection tube was discarded. The column was transferred to a clean collection tube and centrifuged for 1

minute. The column was again transferred a new, nuclease-free tube and 50 ul RNase free water was added onto column and incubated 5 min at RT. The final centrifugation, (14,000 rpm) for 1 min and the recovered purified RNA was frozen at -20°C .

Figure 5.1. Flow-chart of sequencing methods.



Generating cDNA from Polyadenylated RNA

For all procedures during cDNA synthesis, strain MS2 was used as a positive control. First, viral RNA was 3' polyadenylated with yeast PolymeraseA and 25 mM ATP in a 50 ul reaction volume (USB, Inc, Cleveland, OH). The reaction was prepared with 10 ul 5X Reaction Buffer, 10 ul RNA, 2 ul 25 mM ATP, 0.7 ul 600 U Poly(A)Polymerase and 27.3 ul nuclease-free water. The mixture was incubated at 37°C for 5 min and placed on ice for

enzymatic termination. Polyadenylated RNA was either immediately frozen or used as a template for cDNA.

Second, full-length cDNA was prepared using oligo-dT reverse primer supplied with the reverse transcriptase MonsterScript 1st Strand cDNA Synthesis Kit (EpiCentre, Madison, WI) or with a gene-specific reverse primer. To a 250 µl thin-walled PCR tube, the following reagents were added: 4 µl nuclease-free water, 10 µl polyadenylated RNA or RNA template and 1 µl of 10 µM PolyT primer or 1 µl of 10µM gene-specific primer. The mixture was heated for 1 min at 65 °C and chilled for 1 min on ice. To the same tube, 1 µl MonsterScript Reverse Transcriptase and 4 µl of 5X cDNA reaction buffer were added. The mixture was placed in a thermocycler with the following cycle regime: 37 °C for 5 min, 42 °C for 5 min, 60 °C for 40 min. The reaction was terminated by incubating at 90 °C for 5 min and chilled on ice for 1 min. The single-stranded cDNA was either frozen or used for PCR template (Fig 5.1). To verify the generation of full-length cDNA, a partial region of the the 5' end of MS2 was amplified using primers MS25 and MS23 (Lovmar *et al.*, 2003).

To amplify the 1 kb region between the replicase and the 3' end of the genome, strain-specific forward primers were designed based on a 200 nucleotide (nt) region of the replicase gene (Vinjé *et al.*, 2004). To amplify the 5' end of the genome, reverse primers were designed based on the replicase gene sequence of each strain and forward primers were designed based on available full-length sequences (GenBank) of each genogroup. As sequences were generated (Sequetech, Mountain View, CA), reverse primers were designed

to amplify overlapping sections of the genome. The majority of the genome was sequenced by “primer walking.”

5' Amplification of cDNA Ends

The nucleotide sequence of the 5' region was determined by rapid amplification of cDNA end (RACE) with the Smart Race cDNA Amplification Kit (Clontech, Mountain View, CA). First-strand cDNA synthesis was prepared on ice in a 250 ul thin-walled PCR tube by combining 3 ul RNA, 1 ul of 10uM gene-specific reverse primer and 1 ul Smart oligo (from kit). The 5 ul reaction volume was briefly centrifuged and the following components were added: 2 ul of 5X First Strand buffer, 1 ul of 20 mM DTT, 1 ul of 10 mM dNTP and 1 ul SuperScript II (Invitrogen, Carlsbad, CA). Following a brief centrifugation, the mixture was incubated for 90 min at 42 ° C. To dilute the first-strand cDNA, 20 ul of Tricine-EDTA buffer was added and heated for 7 min at 72 ° C. The reaction generated double stranded cDNA. The cDNA was frozen at - 20 ° C and used for subsequent PCR reactions. Concentration of cDNA was determined with a NanoDrop spectrophotometer (NanoDrop Technologies, Wilmington, DE).

Long Template PCR, Cloning and Sequencing

The cDNA was amplified by using Phusion DNA Polymerase (New England Biolabs, Ipswich, MA), with final concentrations of 1X of 5X Phusion Buffer, 0.2 mM dNTP, 1 ul of 10 uM forward primer, 1 ul of 10uM reverse primer, 3% DMSO, 2 ul cDNA and 0.5 ul Phusion Taq in a 50 ul reaction using the following cycle parameters: one round denaturation at 98 °C (1 min), 35 rounds at 98 °C (30 sec), 48 °C (1 min), 72 °C (3 min) followed by 10 min extension at 72 °C. For each reaction, positive controls were prepared using primers

MJV82 and JV81 for *Leviviruses* and MJV82 and JV41 with *Alloleviviruses* (Vinjé et al., 2004). A no-template negative control was included.

PCR products were separated by electrophoresis in a 1.5% agarose gel, stained with SYBR Gold Nucleic Acid Gel Stain (Molecular Probes, Carlsbad, CA) and visualized under blue light (Dark Reader Transilluminators, Clare Chemical Research, Dolores, CO).

Blunt-end PCR products were excised using a gel extraction tool (USA Scientific Plastics, Ocala, FL) and then purified (QuickClean 5M Gel Extraction Kit, GenScript Corporation, Piscataway, NJ). Excised bands were weighed in 1.5 ml microcentrifuge tubes and 3 volumes of Binding Solution II per gel slice were added. The gel solution was heated at 50 °C until melted. One volume of isopropanol was added and the mixture was transferred into the Genprep spin column and centrifuged for 1 min at 12,000 rpm. The column effluent was discarded and 500 µl of Wash Buffer was added, the column was centrifuged and the liquid waste discarded. The 500 µl wash was repeated and waste discarded. The column was placed into a clean 1.5 ml microcentrifuge tube, 30 µl of Elution Buffer was added and the column was incubated 1 min at room temperature. The tube was centrifuged 1 min at 12,000 rpm and the collected DNA eluate was transferred to a clean tube. Concentration of the DNA was determined using a NanoDrop spectrophotometer. The DNA was either cloned or the PCR product was sequenced.

Gel-purified DNA was cloned using a ZeroBlunt TOPO Cloning Kit, pCR-Blunt II TOPO plasmid kit (Invitrogen, Carlsbad, CA). To prepare a TOPO cloning reaction, 4 µl DNA, 1 µl salt solution and 1 µl pCR II Blunt TOPO were added to a nuclease-free tube and incubated 30 min at room temperature. To a vial of One Shot *E. coli* competent cells, 2 µl of

the TOPO cloning reaction were added and incubated on ice for 30 min. The cells were heat-shocked at 42 °C and immediately placed on ice. A 250 µl volume of SOC medium was added to the One Shot cells and shaken (200 rpm) for 1 hr at 37 °C. Fifty µl of transformed cells were plated onto pre-warmed LB agar plates containing 50 µg/ml kanamycin, the transformed cells were spread to isolate colonies and the plates were incubated overnight at 37 °C.

Colonies of transformed *E. coli* cells were screened for positive inserts using whole-cell PCR. Using aseptic techniques, individual colonies were selected with a sterile toothpick, the toothpick was briefly rinsed into a 50 µl Phusion master mix (as described above) then dropped into 10 ml LB/kan broth and incubated overnight at 37 °C. Whole-cell PCR was performed on individual colonies using Phusion DNA Polymerase and the same primers used to generate the pre-cloned amplicon with the following cycle modifications: one round denaturation at 98 °C (3 min), 35 rounds at 98 °C (10 sec), 57 °C (30 sec), 72 °C (30 sec) followed by 10 min extension at 72 °C. Amplicons were separated by electrophoresis in 1.5% agarose gel in 0.5X Tris-acetate-EDTA (TAE), stained with 20 µg/ml ethidium bromide and visualized under UV light (UVP, Upland, CA). Those clones with the appropriate size PCR amplicon were selected for plasmid purification.

Positive clones were plasmid-purified (QIAprep Spin Miniprep Kit, Qiagen, Valencia, CA). An *E. coli* colony that had been selected with a toothpick and incubated overnight was centrifuged 10 min at 8000 rpm (6800 xg). The supernatant was discarded and the cell pellet was resuspended and processed as follows. A 250 µl volume of Buffer P1 was added to the cell pellet, vortexed to mix and transferred to a clean 1.5 ml

microcentrifuge tube. A 350 ul volume of Buffer P2 was added to the resuspended pellet and mixed by inversion followed by addition of 350 ul Buffer N3. The buffer mixture was inverted 4-6 times and centrifuged 10 min at 13,000 rpm (17,900 xg). The supernatant was decanted into the QIAprep spin column and centrifuged 1 min at 13,000 rpm. The column effluent was discarded and the column was washed with 500 ul Buffer PB, the column was centrifuged 1 min and the wash was discarded. To the spin column, 750 ul Buffer PE was added, the column was centrifuged and the wash was discarded. The column was incubated with 50 ul Buffer EB to elute the plasmid. Following a 1 min incubation at room temperature, the column was centrifuged 1 min. Purified plasmid was shipped frozen for sequencing. Each cDNA PCR amplicon was cloned and sequenced in triplicate to obtain the consensus sequence. In some cases, PCR products were sequenced directly. To achieve publication quality data, both forward and reverse strands were sequenced (Sequetech, Mountain View, CA). This process was repeated until complete genomes were obtained.

To avoid contamination, a PCR hood (AirClean 600, AirClean Systems, Raleigh, NC) located in a designated room was used to prepare master mixes separate from template additions. PCR amplification, electrophoresis, template and/or viral preparations ([EPA, 2004](#)) were conducted in individual assigned rooms based on designated use.

Sequence Analyses

Raw sequences from three to five individual clones were imported and aligned using BioEdit v7.0.1 ([Hall, 1999](#)) followed by Basic Local Alignment Search Tool (BLAST, National Center for Biotechnology Information) analyses for sequence and phylogenetic confirmation. Completed sequences from all strains were aligned with full-length prototype

strains (GenBank) using BioEdit ClustalW application. Open Reading Frames (ORF) for each strain was determined using BioEdit.

Similarity analyses were evaluated using SimPlot v3.5.1 (Lole et al., 1999).

Relationships among aligned nucleotide sequences were depicted in similarity plots. The SimPlot program determines the percent identity between a reference sequence and the queried sequence. Percent similarity was calculated within a sliding window 160 bp wide with a step size of 10 bp between plots.

Amino Acid Analysis

Deduced amino acid sequences for each of the four genes were determined using a computer-generated DNA-to-protein translation tool, ExPASy (<http://ca.expasy.org/>).

Prediction of protein sequence motifs were identified by PROSITE (<http://ca.expasy.org/>) and protein families and domains were modeled in Pfam (<http://pfam.janelia.org>).

Genetic distance was calculated for each protein within their respective genogroup as follows. Protein amino acid composition was aligned using BioEdit ClustalW followed by protein distance matrix using Neighbor Joining analysis (BioEdit). Matrices values are the fraction of mismatches at aligned positions. Protdist (BioEdit) protein distance matrix compares the number of amino acid mismatches within each protein. Therefore, the smaller the distance value, the higher the amino acid similarity.

Phylogenetic Analysis

Sequence data were analyzed using BioNumerics Software v.3.5 (Applied Maths, Saint-Martens-Latem, Belgium). Phylogenetic trees were built by global cluster analysis performed on multiple aligned sequences and clustered by UPGMA using the Jukes and

Cantor correction ([Jukes & Cantor, 1969](#)). A bootstrap analysis, based on 10,000 substitutions, was used to measure cluster significance. The reliability of each cluster was expressed on a percentage basis.

Nucleotide Sequence Accession Numbers

The accession numbers of full-length leviviruses sequences available in GenBank were as follows: Genogroup I MS2 (NC_001417.1), M12 (AF195778), fr (NC_0011333.1); Genogroup II GA (NC_001426.1), KU1 (NC_002250.1); Genogroup III M11 (NC_004304.1), Qbeta (AY099114.1), MX1 (NX_001890.1); and Genogroup IV SP (X07489.1), NL95 (AF059243.1), FI (EF068134.1).

Results

Comparison of Full-length Genome Sequences

Full-length genome sequences of 19 FRNA strains were determined in this study and compared to 11 strains previously published in GenBank with respect to genome size and Open Reading Frame(s) locations ([Table 5.1](#)).

Nucleotide sequence similarities among the *Leviviridae* strains are shown in [Table 5.2](#). A total of 7 group I strains were sequenced, DL1, DL2, DL13, DL16, ST4, R17 and J20 and compared to GenBank group I strains MS2, M12 and fr. Group I strains DL2 and DL13 were omitted from this table as they were >99 % identical having only 4 nt single-point mismatches to DL16. MS2 and ST4 were 98.71% similar to each other. Sequence similarity among genogroup I strains ranged from 75.27 - 96.67 % with strain fr forming a separate subgroup ([Table 5.2](#), [Fig 5.2](#)).

Table 5.1 Open Reading Frames positions for *Leviviridae*.

Open Reading Frame Location (nt)						
Strain	Group	Full length nt	ORF1	ORF2	ORF3	ORF4
DL1	I	3570	130-1311	1335-1727	1678-1905	1761-3398
DL2	I	3491 ^b	130-1311	1335-1727	1678-1905	1761-3398
DL13	I	3491 ^b	130-1311	1335-1727	1678-1905	1761-3398
DL16	I	3569	130-1311	1335-1727	1678-1905	1761-3398
J20	I	3569	130-1311	1335-1727	1678-1905	1761-3398
ST4	I	3569	130-1311	1335-1727	1678-1905	1761-3398
R17	I	3569	130-1311	1335-1727	1678-1905	1761-3398
MS2 ^a	I	3569	130-1311	1335-1727	1678-1905	1761-3398
M12 ^a	I	3340 ^b	130-1311	1335-1727	1678-1905	1761-3340 ^b
fr ^a	I	3575	129-1310	1336-1728	1691-1906	1762-3399
GA ^a	II	3465	136-1308	1325-1717	1717-1908	1749-3347
KU1 ^a	II	3486	137-1309	1325-1717	1683-1929	1770-3368
T72	II	3393 ^b	137-1309	1325-1717	1683-1916	1770-3368
DL10	II	3376 ^b	136-1309	1326-1718	1715-1906	1747-3345
DL20	II	3458	137-1309	1326-1718	1718-1909	1750-3348
TW18	III	4218	62-1324	1345-1746	1345-2334	2344-4122
HL4-9	III	4221	62-1324	1345-1746	1345-2337	2347-4125
BR12	III	4218	62-1324	1345-1746	1345-2334	2344-4122
VK	III	4218	62-1324	1345-1746	1345-2334	2344-4122
BZI	III	4219	62-1324	1346-1747	1346-2335	2345-4123
QB ^a	III	4215 ^b	56-1319	1339-1741	1339-2329	2338-4117
M11 ^a	III	4217	57-1322	1344-1744	1344-2333	2352-4118
MX1 ^a	III	4215	56-1321	1343-1744	1343-2332	2351-4111
HP-P22	IV	4241	52-1374	1395-1793	1395-2387	2407-4137
HP-P24	IV	4243	53-1378	1397-1795	1397-2389	2409-4139
BR1	IV	4273	52-1404	1424-1822	1424-2419	2439-4169
BR8	IV	4273	52-1404	1424-1822	1424-2419	2439-4169
NL95 ^a	IV	4248	53-1318	1402-1800	1402-2394	2414-4144
SP ^a	IV	4276	55-1407	1427-1825	1427-2422	2442-4172
FI ^a	IV	4184 ^b	55-1371	1392-1791	1392-2391	2406-4167

nt = nucleotide; ^a previously published GenBank genomes; ^b nearly full-length genome

Three environmental group II strains, DL10, DL20 and T72 were sequenced and

compared to GenBank group II GA and KU1. Among group II strains nucleotide sequence similarity ranged from 83.30 to 93.84% with strains DL10, DL20 and GA having the highest sequence identities (93.43-93.67%) whereas strains T72 and KU1 formed a separate subcluster (Table 5.2, Fig 5.2). Strains in group I had only 50% sequence identity (range of 46.74-53.85%) with strains in group II (Table 5.2, Fig 5.3A). However, all *Levivirus* strains shared an eight nucleotide sequence at the 3' terminus, 5' ACCACCCA 3'.

Among *Allolevivirus* group III, two different subclusters were formed. The first subcluster was composed of strains VK, HL4-9, BR12, BZ1, TW18 and GenBank strain Q β having a nucleotide sequence similarity ranging from 91.87-95.69%. The second subcluster formed with GenBank group III strains MX1 and M11 having an 87% nucleotide similarity to each other. The nucleotide similarity of strains between the two group III subclusters ranged from 69.77-71.33%. Group III strains shared < 40% identity (29.73-39.06%) to *Levivirus* groups I and II (Table 5.2). *Allolevivirus* strains share the 3' terminus signature sequences 5' TCCTCCCA 3' (Table 5.5).

Genogroup IV environmental strains BR1, BR8, HB-P22 and HB-P24 were sequenced and compared to GenBank group IV strains SP, FI and NL95. Group IV *Allolevivirus* shared sequence identities ranging from 74.90-95.03 % with the closest identities being 95.03% between strains BR8 and BR1. Strain HB-P24 shared 90.22% identity with prototype strain NL95, whereas strains BR8 and BR1 shared a greater percent similarity with prototype strain SP (91.05-91.73%). Group IV sequence identity was 53.53-57.99% when compared to *Allolevivirus* group III (Table 5.2, Fig 5.3B) and < 40 % (31.81-

38.73%) when compared to *Levivirus* groups I and II.

Table 5.2 Leviviridae nucleotide percent similarity.

Number/ Strain/ Group	% Nucleotide sequence similarity with the indicated strain																											
	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28
1/ MS2/ I	100.00																											
2/ ST4/ I	98.71	100.00																										
3/ R17/ I	96.39	96.67	100.00																									
4/ M12/ I	92.78	92.93	92.93	100.00																								
5/ J20/ I	92.19	92.47	92.38	91.82	100.00																							
6/ DL1/ I	92.16	92.41	92.16	92.46	93.95	100.00																						
7/ DL16/ I*	91.68	92.07	91.96	92.00	94.73	94.79	100.00																					
8/ tr/ I	75.27	77.09	75.89	75.43	76.63	76.55	75.65	100.00																				
9/ DL10/ II	53.13	52.18	52.54	52.29	53.47	53.15	52.35	49.93	100.00																			
10/ DL20/ II	52.89	52.23	52.17	52.18	53.85	50.65	53.69	51.68	93.84	100.00																		
11/ T72/ II	52.37	52.35	52.49	50.41	51.42	49.99	49.93	46.74	85.97	85.17	100.00																	
12/ GA/ II	51.35	51.44	51.76	51.75	51.14	50.17	51.79	48.25	93.67	93.43	84.79	100.00																
13/ KU/ II	49.55	49.61	49.69	49.72	51.15	50.23	49.58	48.71	84.69	83.83	89.89	83.30	100.00															
14/ VK/ III	38.66	34.85	37.90	33.79	34.59	37.31	36.13	33.01	37.39	37.37	36.40	37.61	36.39	100.00														
15/ HL4-9/ III	37.55	36.25	37.05	36.19	35.25	32.75	36.68	36.13	34.16	38.11	36.51	34.38	39.25	91.97	100.00													
16/ QB/ III	36.87	36.33	36.94	36.15	38.27	34.07	37.88	32.05	38.02	38.44	36.58	34.85	36.62	92.09	94.76	100.00												
17/ BR1/ III	36.85	36.91	36.91	36.47	37.17	36.32	36.19	32.69	37.95	38.49	36.83	38.73	36.44	95.69	91.87	91.97	100.00											
18/ BZ1/ III	36.79	36.91	37.61	36.31	36.68	37.05	36.53	37.00	31.95	32.27	36.70	33.04	39.06	93.37	92.30	92.24	92.87	100.00										
19/ TW18/ III	36.39	36.73	36.39	35.97	35.28	32.71	36.19	31.71	37.63	38.22	36.13	34.73	36.31	92.25	95.90	95.58	92.06	92.65	100.00									
20/ SP/ IV	36.29	34.64	34.73	34.64	34.90	34.77	34.18	33.77	37.09	36.69	32.80	37.13	32.32	55.49	57.75	54.19	54.64	53.09	55.92	100.00								
21/ HB-P24/ IV	35.39	35.47	34.70	35.43	37.00	33.64	38.54	35.85	37.30	33.03	37.54	36.81	38.73	56.13	55.93	53.53	56.73	54.87	55.89	79.37	100.00							
22/ BR1/ IV	34.13	34.13	36.77	34.61	34.59	37.12	34.30	37.55	37.21	33.08	32.77	33.18	31.81	53.61	57.37	53.93	55.36	54.31	56.94	91.73	79.60	100.00						
23/ FI/ IV	34.05	33.96	32.90	37.03	33.81	37.93	37.63	33.20	31.99	33.41	33.07	33.30	32.53	54.43	56.67	55.65	54.85	55.98	56.15	76.63	76.85	74.90	100.00					
24/ BR8/ IV	34.04	36.61	34.19	33.43	34.70	36.68	34.76	33.66	36.66	36.19	36.83	36.91	36.92	55.27	56.64	53.17	55.33	55.49	54.82	91.05	78.87	95.03	74.57	100.00				
25/ M11/ III	33.47	31.11	33.49	30.54	35.66	32.79	32.95	36.14	33.59	33.47	31.82	33.75	30.57	71.33	70.43	70.45	71.05	70.66	71.23	53.67	55.84	52.33	56.03	51.61	100.00			
26/ NL5/ IV	33.10	37.11	32.64	32.55	36.94	32.75	32.87	33.46	36.05	36.04	37.76	37.58	36.99	55.99	56.39	55.81	55.80	54.64	53.92	78.50	90.22	78.47	76.07	78.32	55.89	100.00		
27/ MX1/ III	32.53	32.33	31.21	29.73	36.44	37.49	32.27	33.09	33.47	32.59	37.81	33.45	32.81	70.27	70.55	69.90	70.13	69.77	70.92	52.00	55.08	52.46	54.03	51.19	87.15	54.77	100.00	
28/ HB-P22/ IV	32.30	32.87	32.39	32.28	33.43	32.55	32.65	33.06	36.29	37.01	37.56	36.69	37.54	57.79	57.37	57.65	56.32	57.34	81.80	82.83	82.07	77.69	81.38	54.97	82.25	54.43	100.00	

*DL2 and DL13 were omitted. DL16 was used as a representative strain as all three strains differed by only 4 nucleotides.

Figure 5.2 Phylogenetic analysis of *Leviviridae* nucleotide sequences.

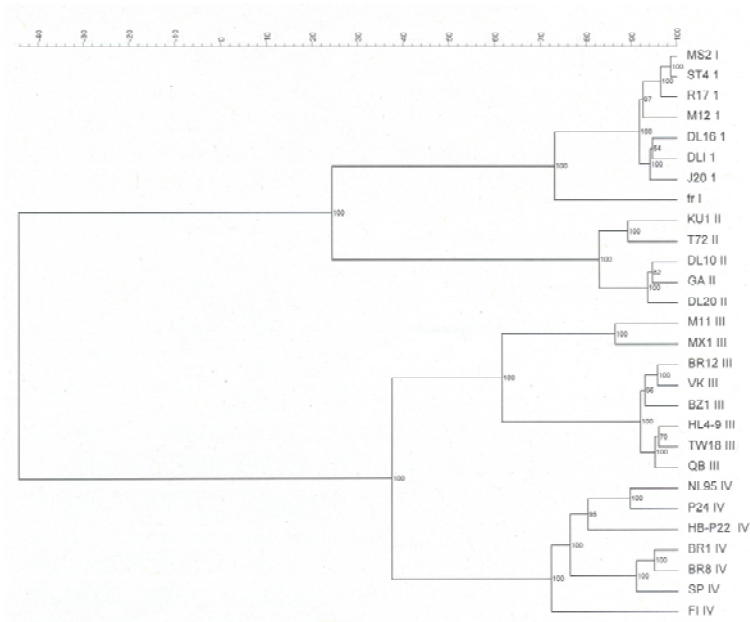
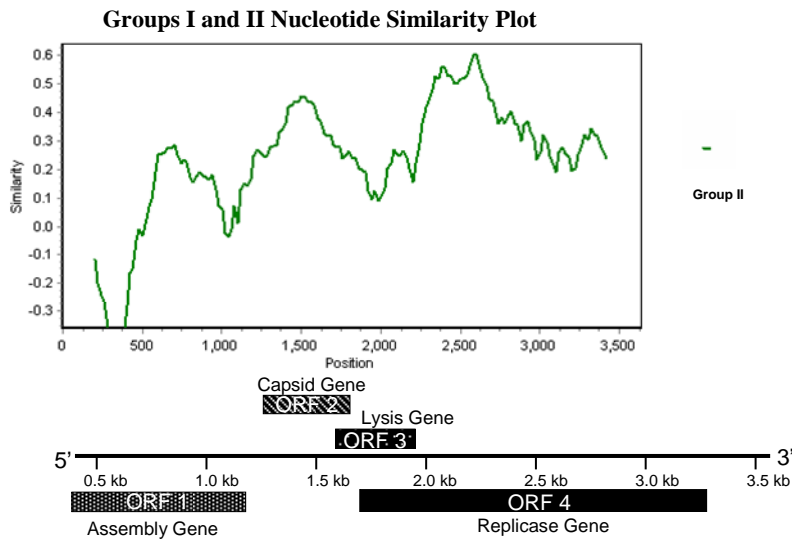


Figure 5.3 SimPlot nucleotide similarity and genome organization.



Open Reading Frame Analyses

Genome lengths, in nucleotides (nt), gene and Open Reading Frame (ORF) locations of all sequenced FRNA phages belonging to genus *Levivirus* and genus *Allolevivirus* are shown in [Table 5.1](#). Shine-Dalgarno sequences and start codons for all genes of these phages are presented in [Table 5.3](#).

Levivirus

For all group I strains except strain fr, the ORF start and stop codons were located at the same nucleotide (nt) positions as was previously reported for strain MS2 ([Fiers et al., 1976](#)). Four proteins and four ORFs were located at the following nt positions: maturation/assembly protein (130-1311), coat protein (1335-1727), lysis protein (1678-1905) and replicase (1761-3398) ([Table 5.1](#)). In contrast, the ORF positions for strain fr were as follows: maturation protein A (129-1310), coat protein (1336-1728), lysis protein (1691-1906) and replicase (1762-3398). The start codon for ORF1, ORF2, ORF3 and ORF4 of genogroup I were AUG with two exceptions (1) MS2, ST4 and fr start codon for ORF1 was GUG and (2) lysis gene start codon, ORF3, for strain fr was UUG ([Table 5.3](#)).

Group II lysis and replicase genes were more similar in nucleotide position among strains T72 and KU1 whereas strains DL10 and DL20 were similar to strain GA ([Table 5.1](#), [Table 5.3](#)). Group II start codons for all ORFs were AUG ([Table 5.3](#)). The four genes in group II differed from group I in their ORF nucleotide positions. Lengths of the 5' untranslated regions of groups I and II genomes were 128 to 129 nt and 135-136 nt length, respectively. Lengths of the 3' untranslated regions were 168-177 nt in group I and 135-140 nt length in group II ([Appendix A](#)).

Table 5.3 Start condons and Shine-Dalgarno sequences. Start codons (bold, underlined) and Shine-Dalgarno (underlined) sequences for each gene. (A) alignment of strains DL1, DL2, DL13, DL16, J20, ST4, R17 and strains M12 and MS2 from Group 1 Levivirus. Strain fr is shown separately. (B) alignment of strains T72, DL10, DL20, and strains KU1 and GA from Group II Levivirus, (C) alignment of strains TW18, HL4-9, BR12, BZ1, VK and strains QB, MX1 and M11 from Group III Allolevivirus, (D) alignment of strains HB-P22, HB-P24, BR1, BR8 and strains SP, NL95 and FI from Group IV Allolevivirus.

(A). Levivirus Group I.

<u>ORF</u>	<u>Gene</u>	<u>Sequences</u>
1	assembly strain fr	CCA <u>U</u> UCCU <u>AGGAGG</u> UUUGACYY <u>RUG</u> CGAGC GCU <u>AGGGAG</u> CCUCGU <u>GUG</u> CGAAAGU
2	capsid strain fr	AACCGGAGUUYGAAGC <u>AUGG</u> CUUCUAA CCGA <u>AGGGAG</u> AGCCAC <u>AUGG</u> CUUCG
3	lysis strain fr	RAUGCA <u>AGG</u> UCUCCURAAAG <u>AUG</u> GAAACCC AACUGGUAACCCAA <u>UUG</u> CAACAGC
4	replicase strain fr	CAUGAGGAUUACCC <u>AUG</u> UCGA ACAUGAGGA <u>AAUACCC</u> <u>AUG</u> UCAAAAU

(B) Levivirus Group II.

<u>ORF</u>	<u>Gene</u>	<u>Sequences</u>
1	assembly	AUACCGGAGGADCU <u>AUG</u> UUUCCGA
2	capsid	AAWWAYGGAGUUAGCCAY <u>AUGG</u> CAACUUUA
3	lysis strains T72, KU1 strains DL10, DL20, GA	GAUUGGGA <u>ACCCGG</u> UUGCUG <u>AUGC</u> CAUCUC CDCAGAGYGGCUUCUACGCGUA <u>AUGG</u> GCUCG
4	replicase strains T72, KU1 strains DL10, DL20, GA	CAUAAGGAAAACCU <u>AUG</u> UCCGAUUCA AAACAUAAGGAAAACCU <u>AUG</u> UCCGAUUCA

(C) Allolevivirus Group III.

<u>ORF</u>	<u>Gene</u>	<u>Sequences</u>
1	assembly	DRGAGGMMAY <u>AUG</u> CCWM
2/3	capsid/readthrough	UUGGGUCAAUUHGAUC <u>AUGG</u> CWAAA
4	replicase TW18, HL4-9, BR12, BZ1, VK, QB MX1 and M11	AGUAACUAAGGAUGAAAUGC <u>AUG</u> UCUAAG AGUAACURAAGGAGAUCUGC <u>AUG</u> UCWA

(D) Allolevivirus Group IV.

<u>ORF</u>	<u>Gene</u>	<u>Sequences</u>
1	assembly	CUACAGAGGAGAAUCU <u>AUG</u> CC
2/3	capsid/readthrough strains BR1, BR8, NL95, SP strains HB-P22, HB-P24, FI	CUUUGGGUCAAUUYGAUC <u>AUGG</u> CAA YUUUGGGUCAAUUYGAUC <u>AUG</u> GCWA
4	replicase strains BR1, BR8, NL95, SP	CUUAARRGAGRWAGC <u>AUG</u> YCAA

Allolevivirus

The *Allolevivirus* genome possesses four genes and 3 start codons as the capsid and readthrough genes share a single ORF (ORF2/ORF3).

For all group III strains, except MX1 and M11, the ORF alignment positions were very similar or identical. Although ORFs of strain Q β aligned perfectly with the other group III strains, the GenBank acquired Q β sequences were not complete. Thus, individually mapped ORF positions varied slightly (Table 5.1). MX1 and M11 ORF and Shine-Dalgarno positions were similar to the other group III strains for the assembly, capsid and readthrough genes, but differed for the replicase gene (Table 5.3).

Within group IV phages BR1, BR8, NL95 and SP had similar ORF positions whereas strains HB-P22, HB-P24 and FI ORFs had similar nt positions (Table 5.1, Table 5.3).

The 5' untranslated length in groups III and IV ranged from 56-62 nt and 50-53 nt, respectively. Groups III and IV 3' untranslated sequence length were 96-98 nt and 97-104 nt, respectively.

Shine-Dalgarno Sequences

Shine-Dalgarno sequences for *Levivivirus* groups I and II were located within 5-9 nt upstream from ORF1 and ORF2, 9-16 nt upstream from ORF3 and 7-8 nt upstream from ORF4. Shine-Dalgarno sequences for *Allolevivirus* groups III and IV were located 5-6 nt upstream from ORF1, 12 nt upstream from ORF2/ORF3, and 6-9 nt upstream from ORF4 (Table 5.3).

With respect to the entire *Leviviridae* family, the ORF positions for the coliphage genes were preceded by Shine-Dalgarno sequences located within 5-16 nt upstream from the ORF start codon(s).

Amino Acid Composition - *Levivirus*

The numerical amino acid (aa) positions are denoted as to their location within their respective protein. *Levivirus* group I demonstrated the most conservative amino acid composition when compared to groups II, III and IV ([Table 5.4, Appendix B](#)).

With the exception of strain fr, amino acid number was consistent in each of the four protein types in group I phages ([Table 5.4, Appendix B](#)). In the lysis protein, strain fr had 71 amino acids, a four codon deletion, when compared to 75 amino acids in the other group I strains. Alignment of the group I lysis protein, including strain fr, indicated 2 stretches of conserved amino acids; one region of 11 amino acids and another region having 13 amino acids. A 15 aa deletion was observed in group II lysis protein strains DL10, DL20, TL2 and GA when compared to T72 and KU1. No consensus sequence was noted in the lysis protein between groups I and II. The capsid protein of all strains in group I was 130 amino acids in length. *Levivirus* groups I and II capsid proteins shared a conserved region consisting of a 10 amino acid, FVLVDNGGTG, consensus sequence. Groups I and II maturation protein shared a consensus region RWLELQ at amino acid positions number 198-203. Groups I and II replicase shared a 24 amino acid conserved region at positions 198-221 and the YDGG sequence near amino acid position 340 ([Appendix B](#)).

Table 5.4 *Leviviridae* number of amino acids per gene. Male-specific coliphage family *Leviviridae*, genera *Levivirus* (groups I and II) and *Allolevivirus* (groups III and IV) table of amino acids. Proteins were determined by mapping of Open Reading Frames and translation of nucleotide sequences to amino acids using ExPASy (<http://ca.expasy.org/>) DNA-to-protein translation tool. The number of amino acids per protein is listed for each genogroup. If differences existed within groups, the amino acid numbers are listed for individual strains. (A) Group I, (B), Group II, (C) Group III and (D) Group IV.

	Maturation	Capsid	Lysis	Replicase
(A) Group I strains				
MS2, M12, DL1, DL2, DL13, DL16, J20, ST4, R17:	393	130	75	545
fr:	393	130	71	545
(B) Group II strains				
GA, DL10, DL20:	390	130	63	532
T72:	390	130	75	532
KU1:	390	130	79	532
(C) Group III strains	Maturation	Capsid	Read-thru	Replicase
QB, BR12, TW18, BZ1, VK:	420	133	328	592
HL4-9:	420	133	329	592
MX1:	421	133	328	586
M11:	421	133	328	588
(D) Group IV strains				
SP, BR1, BR8:	450	132	330	576
FI:	438	132	332	586
NL95:	442	132	331	576
HB-P24:	441	132	329	576
HB-P22:	440	132	329	576

Pfam - *Levivirus*

Not all of the four genes, as determined by ORF mapping, were grouped into both a protein domain and family membership by Pfam. A PfamA maturation protein search generated “phage_mat-A” domain and a total of 24 *Leviviridae* phages including all four genogroups and the additional species PRR1, PP7 and AP205. *Levivirus* capsid amino acid compositions were placed into the “Levi_coat” family and the replicase protein was placed into the “RNA replicase, beta-chain” domain. The group I lysis protein was not sorted to a family or domain in a PfamA search. A subsequent PfamB search for the group I lysis protein linked it to a lysis domain and the results generated *Levivirus* group I species fr, M12, MS2 and JP501.

For each protein, Pfam recognized *Leviviridae* ssRNA viral species. In some cases, such as the group I capsid protein, GenBank *Leviviridae* strains from groups I, II, III, IV and bacteriophage PRR1 were included in the Pfam species tree. In addition to the *Leviviridae* family, results of searches for the groups I and II replicase species tree added bacteriophages PRR1, ZR, BO1 and Acinetobacter phage AP205.

Protein sequence motifs - *Levivirus*

Predicted protein motifs, casein kinase II phosphorylation, cAMP and cGMP-dependent protein kinase phosphorylation, protein kinase C phosphorylation, N-myristoylation, N-glycosylation and tyrosine kinase phosphorylation, occurred frequently in the FRNA coliphages. Excluding strain fr, the maturation protein of groups I and II shared one amino acid motif with the casein kinase II phosphorylation protein and the replicase gene shared one amino acid position motif with the N-myristoylation protein.

No amino acid motif positions were shared between groups I and II in the capsid or lysis proteins. Unique to strain fr was the presence of a leucine zipper in the lysis protein and an amidation motif in the replicase region. The replicase gene RNA-dependent RNA polymerase catalytic domain occurred at amino acid positions 243-373 and 245-375 for groups I and II, respectively. Common to every group II strain was a prenyl group binding site (CAAX box) at amino acid positions 529-532 in the replicase region.

Genetic distances - *Levivirus*

Excluding strain fr, group I amino acid compositions were very similar as the genetic distance was very close among all four proteins. The capsid protein was most similar (distance of 0.0000 - 0.0411) followed by replicase (0.0033-0.0887), maturation protein (0.0046-0.0889) and lysis (0.0232-0.3416). Capsid protein was identical among strains DL1, DL2, DL13, DL16 and J20 (distance 0.0000). Amino acid composition of all four strain fr proteins showed the greatest distance from the other group I strains (0.2316-0.5685).

Closest to furthest genetic distance in the amino acid composition for the group II strains were as follows: capsid (0.0135-0.1116), maturation (0.0235-0.2036), replicase (0.0415-0.2160) and lysis (0.0500-4.6735).

Amino Acid Composition - *Allolevivirus*

The length of the maturation protein of groups III and IV varied from 420 aa to 450 aa ([Table 5.4](#)) and possessed a mutual conserved aa region LWLEFRYGL ([Appendix B](#)). The length of the capsid protein was 133 and 132 aa for groups III and IV, respectively, and conserved stretches of amino acids occurred in both groups. Groups

III and IV read-through protein was 328 to 332 aa in length and possessed conserved regions between aa positions 290-310. Consensus, however, was unique to each group. Compared to the other group III strains MX1 and M11 shared a three amino acid deletion at the 5' end. The group IV maturation protein in strains HB-P22, HB-P24, NL95 and FI had a 9 amino acid deletion when compared to the other group IV strains. The *Allolevivirus* replicase gene was 576-592 amino acids in length revealing the longest region of conserved amino acids ranging from amino acid positions 202-247 in group IV and positions 207-240 in group III. Groups III and IV replicase shared the sequence KAVTVPKNSKTDRCIAIEPGWNMFFQL in the 210-235 aa region and the YGDD sequence near amino acid position 360 ([Appendix B](#)).

Pfam - *Allolevivirus*

Identical results were obtained for groups III and IV PfamA and PfamB searches. Similar to the *Levivirus*, a maturation protein search generated “phage_mat-A” domain and a total of 24 *Leviviridae* phages including all four genogroups and the additional bacteriophage strains PRR1, PP7 and AP205. PfamA displayed the capsid protein in the family “Levi_coat” and linked to a 31-member *Leviviridae* species tree which included all four genogroups along with phages ZR, TH1, TL2, SD, f2 and BO1 plus bacteriophage strains PRR1. Strains PP7 and AP205 were not included in the capsid species tree.

Read-through proteins were grouped as “A1-protein coat readthrough” with PfamB generating a 5-member species tree of SP, QB, NL95, MX1 and M11. As with the *Levivirus*, the *Allolevivirus* replicase protein was sorted into the “RNA replicase, beta-

chain” family including a 24-member *Leviviridae* species tree with additional bacteriophages PRR1, ZR, PP7, BO1 and AP205.

Protein sequence motifs - *Allolevivirus*

As observed in the *Levivirus* genus, the most prevalent *Allolevivirus* protein motifs were casein kinase II phosphorylation, cAMP and cGMP-dependent protein kinase phosphorylation, protein kinase C phosphorylation, N-myristoylation, N-glycosylation and tyrosine kinase phosphorylation. With the exception of group III strains MX1 and M11 and group IV strain HB-P24, a cell attachment motif (RGD) in the maturation protein was present in the majority of group III and IV strains. Group IV strains SP, BR8, BR1 and HB-P22 had a cell attachment motif in the read-through protein.

The catalytic domain of the RNA-dependent RNA polymerase (replicase protein) was located at amino acid positions 262-394 in group III strains except for M11 and MX1. Their catalytic domain was located at amino acid positions 259-391. Group IV catalytic domain was located at amino acid positions 259-391.

Genetic distances - *Allolevivirus*

Group III amino acid genetic distances were most similar in the capsid protein (distance 0.0000-0.3734) followed by replicase (0.0278-0.6571), maturation protein (0.0347-0.8289) with the greatest genetic distance in the read-through protein (0.0444-0.5128). Strains BR12 and VK shared identical capsid proteins (distance of 0.0000).

In group IV, highly similar amino acid compositions were found in the replicase

(distance 0.0474-0.3382), followed by the capsid (0.0535-0.2569) and read-through protein (0.0555-0.5072). The greatest genetic distance was observed in the maturation protein (0.0607-0.5646) in group IV strains.

Phylogenetic Analyses

An algorithmic approach was selected to construct phylogenetic trees from the nucleotide sequence and amino acid data (Fig 5.2, Fig 5.4). Nucleotide sequences in the phylogenetic tree of *Levivirus* group I strains produced two branches, with 9 strains clustered as MS2-like and strain fr an individual branch (Fig 5.2). Within group II nucleotide sequences, strains KU1 and T72 formed one branch and strains DL10, DL20 and GA formed a second branch. *Allolevivirus* group III nucleotide sequences clustered into a MX1, M11 branch and a second branch with Q β -like strains BR12, VK, BZ1, HL4-9, TW18 and prototype Q β . Nucleotide sequence analysis formed three branches in group IV strains as follows: 1) HB-P24, HB-P22 and prototype NL95, 2) BR1, BR8 and prototype SP and 3) prototype FI (Fig 5.2).

Individual proteins were clustered into phylogenetic trees (Fig 5.4). In some cases, phylogenetic protein trees formed more subclusters or branches than the nucleotide trees. For example, *Levivirus* group II lysis protein formed a separate branch (strains T72, KU1) whereas the remaining genogroup II strains (DL20, TL2, DL10, GA) formed a branch off of the group I strains (Fig 5.4). Protein trees generated for *Allolevivirus* were similar to nucleotide phylogenetic clustering. Group III formed two branches, MX1 with M11 and QB-like (6 strains). Group IV generated four subclusters for each protein

tree with the least amount of variation, >90% similarity, noted for the capsid protein.

SimPlots provide a visual picture of regions of similarity when two or more strains are compared. When nucleotide genomes were compared for all Groups I and II strains, SimPlots showed that the replicase was most similar whereas the maturation gene shared the least amount of nucleotide regions (Fig 5.3A). When complete genome nucleotide sequences were compared between groups III and IV, SimPlot graphs showed similar nucleotide regions in the capsid and the 5' portion of the replicase, but least similar in the maturation and 3' region of the replicase (Fig 5.3B)

Table 5.5 Comparison of genomic traits. *Allolevivirus* and *Levivirus*.

	<u>Allolevivirus</u>	<u>Levivirus</u>
Genome size (nt)	4215-4276	3458-3575
Gene number	4	4
Proteins	4	4
Number of ORF initiation sites	3	4
Protein names	assembly/maturation capsid read-through replicase	assembly/maturation capsid lysis replicase
ORF initiation sites	same for 2 nd and 3 rd genes	no sharing of ORF initiation sites
replicase gene	single gene	Start codon occurs within lysis protein
3' terminus	5' TCCTCCCA 3'	5' ACCACCCA 3'

Figure 5.4 (A) Phylogenetic analysis of each protein: Levivirus I, II.

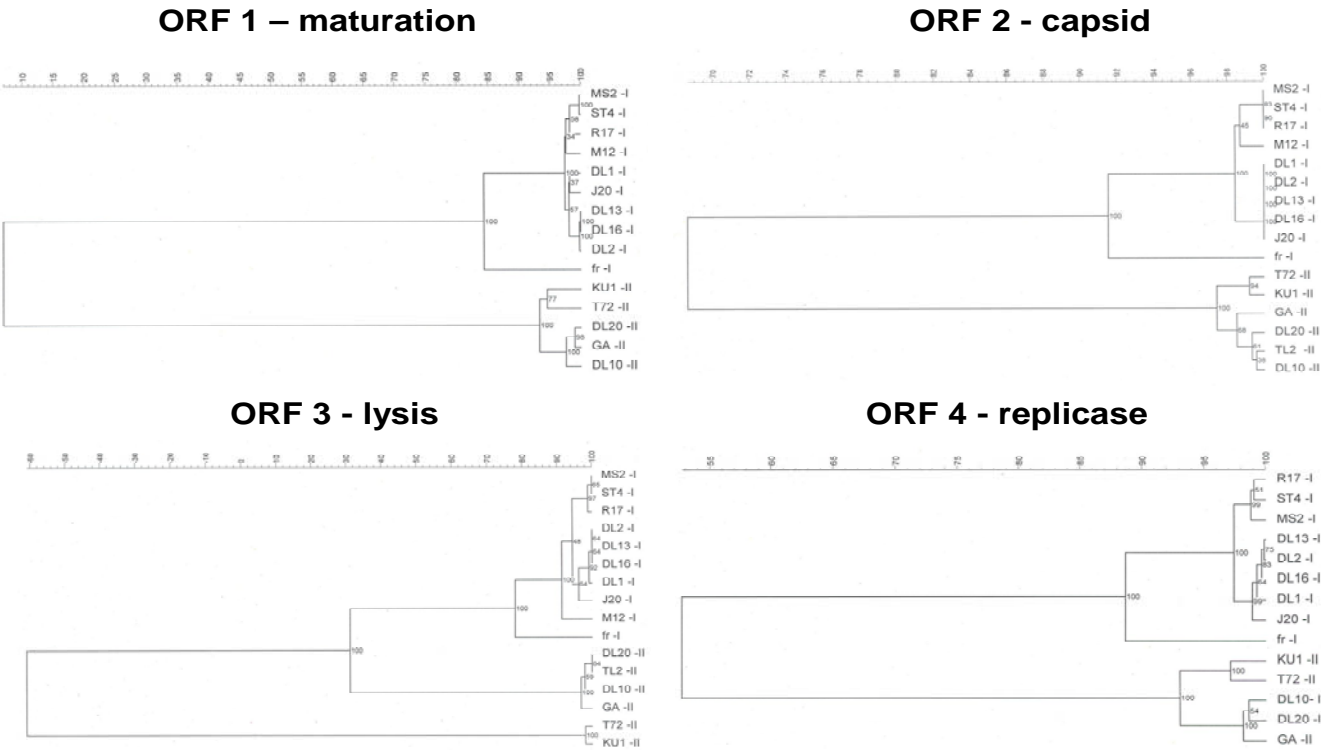
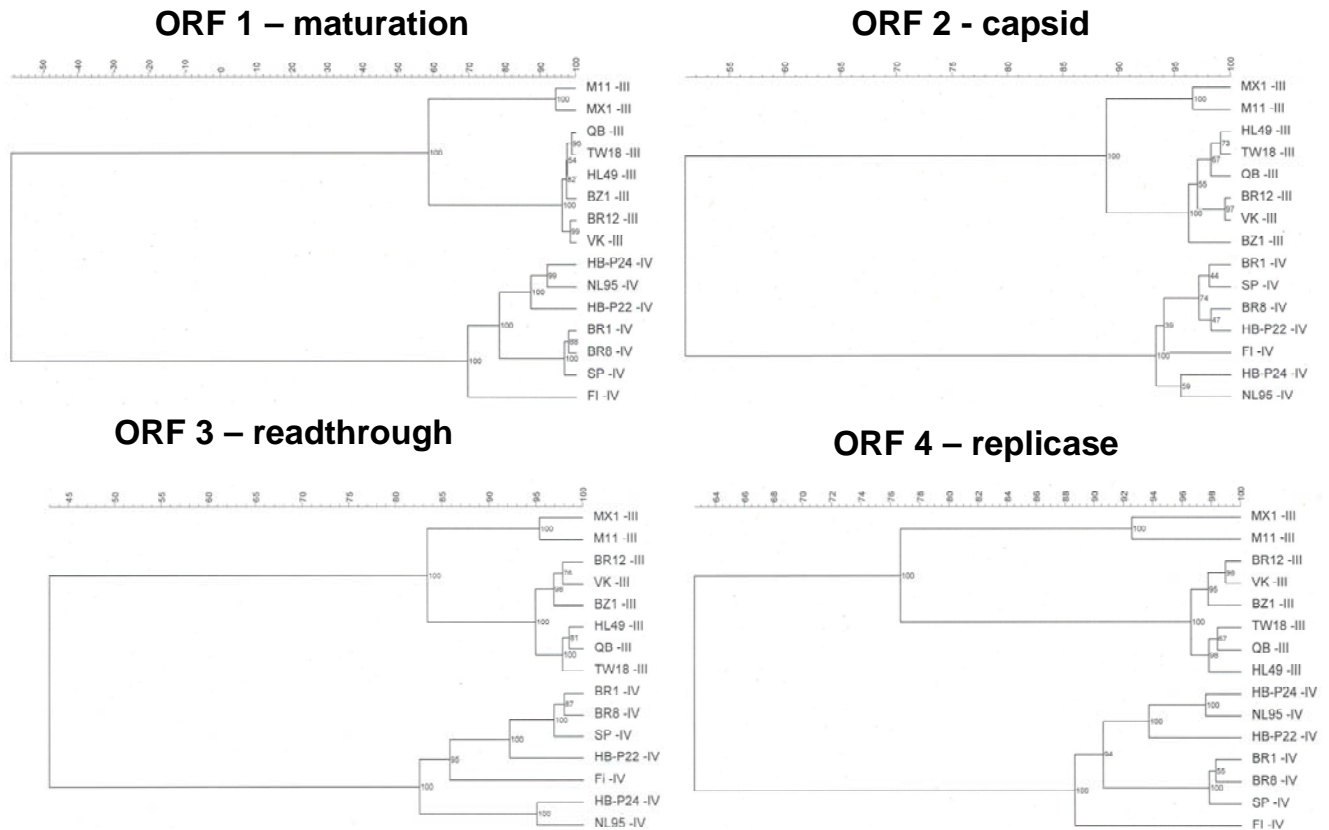


Figure 5.4 (B) Phylogenetic analysis of each protein: Allolevivirus III, IV.



Discussion

Geographically dispersed strains showed more than 90% sequence identity ([van Duin, 1998](#)). Our data supports this finding ([Fig 5.1, Table 5.2](#)). Although strains DL13, DL2 and DL16 were collected from widely different geographic locations ([Table 7.1](#)), sequence alignment among these three strains revealed only 4 nt single-point mismatches throughout the genome with a >99% sequence similarity ([Appendix A](#)). With the exception of strain fr, group I sequence identities were greater than 90% ([Table 5.2](#)). Strain fr shared a sequence identity no closer than 77.09 % to any of the other group I phages ([Table 5.2](#)).

Sequence analyses of the 11 FRNA GenBank strains showed uniformity throughout the *Leviviridae* family ([Klovins et al., 2002](#)). Data reported here supports and advances those findings. Capsid/coat protein amino acid length was highly conserved in the *Leviviridae* family ranging from 130-133aa ([Table 5.4](#)). Capsid amino acid length is apparently constant in ssRNA phages ([Klovins et al., 2002](#)) and may be related to restrictions by capsid infrastructure ([Nishihara et al., 2006](#)). In both the previously sequenced *Levivirus* groups I and II phages and in those sequenced in the present study the lysis gene was embedded out of frame as the initiation codon lies at the 3' end of the capsid gene and the termination codon was at the 5' region of the replicase gene. Subsequently, the replicase ORF4 initiation site begins at the 3' region of the lysis gene ([Fig 5.3 map, Appendix A](#)). A replicase secondary structure was reported as a conserved amino acid motif, YGDD, in all ssRNA replicases ([Olsthoorn et al., 1995](#)). Replicase amino acids from the 30 *Leviviridae* strains reported here and the bacteriophages PP7, AP205 and PRR1 shared the YGDD motif. Also, noteworthy from the sequence data was confirming the *Leviviridae* family conserved

sequence, CCCA_{OH}, at the 3' end of the genome (Klovins et al., 2002; van Duin and Tsareva, 2006). This observation was extended by identifying genus-specific *Allolevivirus* and *Levivirus* signatures of 8 nucleotides at the 3' termini that clearly distinguish *Allolevivirus*, 5' TCCTCCCA 3' from *Levivirus*, 5' ACCACCCA 3' (Table 5.5).

In all members of the *Leviviridae* family, the ORF positions for the coliphage genes were preceded by Shine-Dalgarno sequences located 5-16 nt upstream from the start codon(s) (Table 5.3). In prokaryotes, greater than 80% of Shine-Dalgarno sequences occur within 5-13 bases upstream (Ma et al., 2002). Thus, the positions reported here for these viral genomes are similar to the prokaryote criteria.

Stop codons, UAG, UAA, UGA serve as signals for peptide chain termination. During translation of the viral RNA coat protein cistron, the UGA stop codon can be read through, resulting in an additional translated product (Weiner and Weber, 1973). In *Allolevivirus*, a read-through protein resulted when a leaky UGA stop codon is misread as a tryptophan codon (UGG) (van Duin, 2006), influencing regulatory control and efficiency of gene expression (Beier and Grimm, 2001). Alignment of group III nucleotides revealed that the *Allolevivirus* maturation protein stop codon is also a UGA; however, in this instance, a non-leaky codon. This may occur because the 5' and 3' codons flanking the UGA influence translation termination efficiency (Namy et al., 2001, Bertram et al., 2001, Skuzeski et al., 1990). It was proposed that programmed read-through in strain Q β was regulated by the 3' nucleotides, specifically an A residue, flanking the stop codon in strain Q β (Engelberg-Kulka, 1981). However, alignment of group III and IV nucleotide data in the present study did not reveal a 3' flanking pattern following the UGA stop codon nor was a 3' residue

observed immediately following the stop codon in all group III strains. Noticeably, Q β -like strains did contain the 3' A residue but not MX1 or MX11. Interestingly, in all *Allolevivirus* sequences, a 5' pattern emerged at the read-through stop codon but was absent in the maturation UGA stop codon. Beginning 12 nt upstream from read-through UGA, the sequence AAY CCR GCR UAY STOP in group III and AAY CCW GCN UAC STOP in group IV was observed. Nucleotides common to both III and IV are underlined. These findings suggest the upstream sequences may reduce translation termination efficiency of the UGA read-through stop codon in *Allolevivirus* sp.

Representation of protein domain, family conservation and discrete amino acid sequence features, or motifs, were observed by Pfam and PROSITE patterns. Pfam domain groupings were more broadly defined. The replicase beta-chain and capsid domains were distributed throughout the *Leviviridae* family, along with a few additional bacteriophages. In contrast, the lysis and read-through domains were genogroup-specific.

As *Pseudomonas aeruginosa* ssRNA phage PP7 shared secondary regulatory RNA structures with *Leviviridae* it was classified into the genus *Levivirus* (van Duin and Tsareva, 2006) despite the lack of sequence similarity (Olsthorn et al., 1995, van Duin and Tsareva, 2006) and amino acid clustering (Ruokoranta et al., 2006). Pfam protein domain profile in this study supported the observations that the lysis and capsid proteins of phage PP7 failed to cluster to these same proteins in members of the *Levivirus* or the capsid and read-through proteins in members of the *Allolevivirus*. Data from the present study also indicated that phage PP7 replicase protein did not cluster to the replicase members of the *Levivirus* but did cluster to *Allolevivirus* replicase protein. Common to ssRNA phages, the replicase amino

acid motif, YGDD, was present in phage PP7 replicase. Phage PP7 maturation protein did cluster to this same protein in both *Levivirus* and *Allolevivirus*. Data from this and previous studies suggest that the ssRNA phage PP7 genetic map was structured similar to *Levivirus* groups I and II (Ruokoranta et al., 2006).

Phage PRR1 has a broad host range related to a plasmid IncP-encoded receptor, adsorbs to host pili and displays a genetic map similar to *Leviviridae* (Ruokoranta et al., 2006). PRR1 shared approximately 43-48% sequence identity to other ssRNA phages and clustered outside the *Levivirus* and *Allolevivirus* genera (Ruokoranta et al., 2006). PRR1 genetic map was similar to *Leviviridae* and subsequently phage PRR1 was grouped into the *Levivirus* genus (Ruokoranta et al., 2006). Pfam analysis presented here resulted in phage PRR1 sharing Pfam domains with *Leviviridae* maturation, capsid and replicase proteins. PRR1 displayed the ssRNA amino acid YGDD replicase motif but did not share the signature *Levivirus* 3' terminus ACCACCCA.

The ssRNA phage from *Acinetobacter*, AP205, shared Pfam domains in the *Levivirus* and *Allolevivirus* maturation and replicase proteins only. Amino acid compositions from AP205 and their corresponding coat, maturation, lysis and replicase proteins clustered outside the *Levivirus* and *Allolevivirus* tree (Ruokoranta et al., 2006). AP205 lacked significant sequence similarity but shared important structural features with *Leviviridae* (Klovins et al., 2002). As with phages PRR1 and PP7, phage AP205 did not share the 3' termini *Levivirus* or *Allolevivirus* signatures but had the replicase YGDD motif. NCBI GenBank classified bacteriophages PRR1, PP7 and AP205 into an “unclassified” category.

Using PROSITE, conserved amino acids and subsequent motifs identified in amino

acid sequences of the phage proteins provided insight to structural features. A cell-attachment motif (RGD) was identified in both the maturation and/or read-through proteins in the majority of *Allolevivirus* strains but was absent in *Levivirus* strains. The function of the RGD motif in *Leviviridae* coliphages has yet to be experimentally demonstrated but may be explained because in *Levivirus* strains the phage attaches to the host's pili via the maturation protein; in *Allolevivirus* strains both the maturation and read-through proteins are required for phage infection ([van Duin, 1999](#)). The Arg-Gly-Asp (RGD) motif was shown to be involved in cell-to-cell adhesion in the passaged foot-and-mouth disease virus ([Martinez et al., 1997](#)), in enterovirus, echovirus 9 strain Barty, coxsackievirus A9, echovirus 22 ([Nelsen-Salz et al., 1999](#)) and the blue-tongue virus ([Tan et al., 2001](#)). In nearly all *Astrovirus sp.* an RGD or similar integrin-recognition motif was observed ([van Hemert et al., 2007](#)). In contrast, a second RGD motif in a SAT1 foot-and-mouth virus was not necessary for cell-to-cell attachment ([Storey et al., 2007](#)).

As more Gram negative bacteriophage sequences are elucidated, the protein domains, phylogenetic relationships and novel virus groups will likely emerge and enrich the database.

Conclusion

The findings of this study agree with previously determined FRNA features and phylogenetic analyses which concluded that the *Leviviridae* contain two genera and four distinct genogroups consisting of two genogroups within each genus. Despite the fact that genogroup I strain fr, and genogroup III strains MX1 and M11 only shared between a 70-78% sequence identity with the rest of the strains in their respective genogroups, the analyses suggested that fr does belong in *Levivirus* group I, and MX1 and M11 belong in *Allolevivirus*

group III. Distinguished features such as amino acid and nucleotide similarity and catalytic domain location tend to sub-cluster the strains. For example, Q β -like strains clustered together when compared to MX1 and M11 in group III. Genogroups within each genera shared approximately 50% sequence identity whereas between the two genera, *Levivirus* and *Allolevivirus*, <40% nucleotide sequence identity was observed. Genome organization, amino acid conservation and identical or very similar nucleotide start and stop positions supported the genogroup designation. In addition, eight nucleotides on the 3' termini clearly distinguish the *Allolevivirus*, 5' TCCTCCCA 3', from the *Levivirus*, 5' ACCACCCA 3'.

Summary

- Alignment of nine group I environmental strains with reference strain MS2 showed identical ORF and start codon positions for all four *Levivirus* genes, indicating that the sequence data generated in the present study was valid.
- All strains for groups II, III and IV had similar results in that the sequenced genomes and gene maps showed identical or very similar nucleotide positions to the GenBank reference strains.
- Basic Local Alignment Search Tool (BLAST) of all environmental strains placed them into their respective genogroups.
- Amino acid composition was similar among FRNA coliphage genogroups, further validating the nucleotide sequences and the groupings based upon them.

References

- Beier, H, and M Grimm. 2001. Misreading of termination codons in eukaryotes by natural nonsense suppressor tRNAs. *Nucl Acids Res*, 29(23):4767-4782.
- Bertram G, S Innes, O Minella, JP Richardson, and I Stansfield. 2001. Endless possibilities: translation termination and stop codon recognition. *Microbiol*, 147:255-269.
- Bollback, J.P. and J.P. Huelsenbeck. 2001. Phylogeny, genome evolution, and host specificity of single-stranded RNA bacteriophage (family *Leviviridae*). *J Mol Evol* 52:117-128.
- Buchen-Osmond, C. (Ed.) 2004. Levivirus. In ICTVdB - *The Universal Virus Database*, version 3. ICTVdB Management, Columbia University. New York, NY.
- Crawford, EM, and RF Gesteland. 1964. The adsorption of bacteriophage R17. *Virol*, 22:165-167.
- EPA. 2004. Quality assurance/quality control guidance for laboratories performing PCR analyses on environmental samples. EPA 815-B-04-001. October 2004.
- Engelberg-Kulka, H. 1981. UGA suppression by normal tRNA^{Tp} in *Escherichia coli*: codon context effects. *Nucl Acids Res*, 9(4):983-991.
- Fiers W, R Contreras, F Duerinck, G Haegeman, D Iserentant, J Merregaert, W Min Jou, F Molemans, A Raeymaekers, A Van den Berghe, G Volckaert and M Ysebaert. 1976. Complete nucleotide sequence of bacteriophage MS2 RNA: primary and secondary structure of the replicase gene. *Nature*, 260:500-507.
- Grabow, W.O.K., 2001. Bacteriophages: update on applications as models for viruses in water. *Water S.A.* 27, 251-268.
- Havelaar, A. H., M. van Olphen, and Y. C. Drost. 1993. F-specific RNA bacteriophages are adequate model organisms for enteric viruses in fresh water. *Appl. Environ. Microbiol.* 59:2956–2962.
- Jukes, T. H., and C. R. Cantor. 1969. Evolution of protein molecules, p. 21-123. In H. N. Munro (ed.), *Mammalian Protein Metabolism*. Academic Press, New York.
- Karnik, S. and M. Billeter. 1983. The lysis function of RNA bacteriophage Q β is mediated by the maturation protein. *EMBO J.* 2:1521-1526.
- Klovins, J., G.P. Overbeek, S.H.E. van den Worm, H.-W. Ackermann and J. van Duin. 2002.

- Nucleotide sequence of a ssRNA phage from *Acinetobacter*: kinship to coliphages. *J Gen Virol*, 83, 1523-1533.
- Loeb, T. and N.D. Zinder. 1961. A bacteriophage containing RNA. *Proc. Natl. Acad. Sci. U.S.* 47:282-289.
- Lole, KS., RC Bollinger, RS Paranjape, D Gadkari, SS Kulkarni, NG Novak, R Ingersoll, HW Sheppard and SC Ray. 1999. Full-length human immunodeficiency virus type 1 genomes from subtype c-infected serconverters in India, with evidence of intersubtype recombination. *J Virol*, 73(1):152-160.
- Ma J, A Campbell and S Karlin. 2002. Correlations between Shine-Dalgarno sequences and gene features such as predicted expression levels and operon structures. *J Bact* 184(20):5733-5745.
- Martinez, MA, N Verdaguer, MG Mateu, and E Domingo. 1997. Evolution subverting essentiality: dispensability of the cell attachment Arg-Gly-Asp motif in multiply passaged foot-and-mouth disease virus. *Proc Natl Acad Sci*, 94:6798-6802.
- Murphy FA, Fauquet CM, Bishop DHL, Ghabrial SA, Jarvis AW, Martelli GP, Mayo MA, Summers MD (1995) *Virus taxonomy: The classification and nomenclature of viruses. The sixth report of the International Committee on Taxonomy of Viruses.* Springer-Verlag, Vienna
- Namy, O, I Hatin and JP Rousset. 2001. Impact of the six nucleotides downstream of the stop codon on translation termination. *EMBO Reports*, 2(91):787-793.
- Nelsen-Salz, B, HJ Eggers, and H Zimmermann. 1999. Integrin $\alpha_v\beta_3$ (vitronectin receptor) is a candidate receptor for the virulent echovirus 9 strain Barty. *J Gen Virol*, 80:2311-2313.
- Nishihara T., S. Fujisaki, Y. Nishimura, Y. Minami and T. Yubisui. 2006. Analysis of six new genes encoding lysis proteins and coat proteins in *Escherichia coli* Group A RNA phages. *Microbiol. Immunol.*, 50(1), 61-66.
- Olsthoorn, RCL, G Garde, T Dayhuff, JF Atkins, and J van Duin. 1995. Nucleotide sequence of a single-stranded RNA phage from *Pseudomonas aeruginosa*: kinship to coliphages and conservation of regulatory RNA structures. *Virol*, 206:611-625.
- Paranchych, W. 1975. Attachment, ejection and penetration stages of the RNA phage infectious process. In: ND Zinder (ed). *RNA Phages.* Cold Spring Harbor Laboratory. pp. 85-111.
- Ruokoranta, TM, AM Grahn, JJ Ravantti, MM Poranen and DH Bamford. 2006. Complete genome sequence of the broad host range single-stranded RNA phage PRR1 places it

- in the *Levivirus* genus with characteristics shared with Alloleviviruses. *J Virol*, 80(18):9326-9330.
- Skuzeski, JM, LM Nichols and RF Gesteland. 1990. Analysis of leaky viral translation termination codons in vivo by transient expression of improved beta-glucuronidase vectors. *Plant Mol Biol*, 15(1):65-79.
- Stewart, J.R., J. Vinjé, S.J.G. Oudejans and G.I. Scott., 2006. Sequence variation among group III F-specific RNA coliphages from water samples and swine lagoons. *Appl. Env. Micro*, 72, 1226-1230.
- Storey, P, J Theron, FF Maree and HG O'Neill. 2007. A second RGD motif in the 1D capsid protein of a SAT1 type foot-and-mouth disease virus field isolate is not essential for attachment to target cells. *Viral Res*, 124:184-192.
- Sundram A, N Jumanial, and MM Ehlers. 2006. Genotyping of F-RNA coliphages isolated from wastewater and river water samples. *Water SA*, 32(1):65-70.
- Tan, BH, E Nason, N Staeuber, W Jiang, K Monastyrskaya and P Roy. 2001. RGD tripeptide of bluetongue virus VP7 protein is responsible for core attachment to *Culicoides* cell. *J Virol*, 75(8):3937-3947.
- van Duin, J. 1988. The single-stranded RNA bacteriophages. In: Fraenkel Conrat H and Wagner RR. (eds). *The Bacteriophages*. Series *The Viruses*. Chapter 4. Plenum Press, NY. pp. 117-167.
- van Duin, J., 1998. The single-stranded RNA bacteriophages. In: Fraenkel Conrat, H., Wagner, R.R. (eds). *The Bacteriophages*. Series *The Viruses*. Plenum Press, NY. pp. 117-167.
- van Duin, J. 1999. Single-stranded RNA phages (*Leviviridae*). In: Granoff, A and R Webster (eds). *Encyclopedia of Virology*. Academic Press, London. pp. 1663-1668.
- van Duin, J. 2000. Family *Leviviridae*. In: M.H.V. van Regenmortel, C.M. Fauquet, D.H.L. Bishop, E.B. Carstens, M.K. Estes, S.M. Lemon, J. Maniloff, M.A. Mayo, D.J. McGeoch, C.R. Pringle, R.B. Wickner (eds). *Virus Taxonomy*. Seventh Report of the International Committee on Taxonomy of Viruses. Academic Press, San Diego. pp. 645-646.
- van Duin, J, and Tsareva, 2006. Single-stranded RNA phages. In: *The Bacteriophages* by S T Abedon. R L Calendar, ed. Oxford University Press. pp:175-196.
- Weber, K, and W Konigsberg. 1975. Proteins of the RNA phages. In: ND Zinder (ed).

- RNA Phages. Cold Spring Harbor. pp. 51-84.
- Weiner, AM and K Weber. 1973. A single UGA codon functions as a natural termination signal in the coliphage Q β coat protein cistron. *J Mol Biol*, 80:837-855.
- van Hemert, FJ, VV Lukashow and B Berkhout. 2007. Different rates of (non-) synonymous mutations in astrovirus genes; correlation with gene function. *Virology*, 4:25-37.
- Vinje, J., S.J.G. Oudejans, J.R. Stewart, M.D. Sobsey and S.C. Long., 2004. Molecular detection and genotyping of male-specific coliphages by reverse-transcription-PCR and reverse line blot hybridization. *Appl. Env. Micro.*, 70(10), 5996-6004.

VI. Genomic sequences of two novel *Levivirus* FRNA coliphages (*Leviviridae*): Evidence for recombination?

Abstract

Male-specific ssRNA coliphages, family *Leviviridae*, contain two genera and four distinct genogroups. The *Levivirus* genus is subdivided into genogroups I and II whereas the *Allolevivirus* genus consists of genogroups III and IV. During an environmental genotyping study of *Leviviridae* ssRNA coliphages, two novel strains became evident. Nucleotide sequences and phylogenetic analysis of a 189 bp amplicon in the replicase gene clustered the strains between *Levivirus* genogroups I and II, leading to a proposed genogroup I *Levivirus* subcluster, termed JS-like. Seventeen strains from genogroups I and II, including two JS strains, were used to examine the genomic and phylogenetic relationships among these *Levivirus* groups. The two JS strains were 96.73% similar in nucleotide sequences to each other and 80-84 % nucleotide sequence similarity was shared between the JS strains and nine MS2-like strains. Amino acid composition between JS strains and MS-2 like strains of the maturation, capsid and lysis proteins shared 99-100%, 98-100% and 95-100%, respectively. However, the replicase amino acid sequences of the JS strains shared only 84-85% amino acid similarity to nine MS2-like strains. *Levivirus* JS strains diverged from group I in the replicase gene downstream from the conserved catalytic site. Analysis of complete genome sequences, amino acid composition, phylogenetic relationships and unique clustering suggest the JS strains are recombinants.

Introduction

Vinje et al (2004) investigated the genetic diversity of male-specific (FRNA) phages. In that Vinje study, a phylogenetic analysis of 32 *Levivirus* (genogroups I and II) field strains using a 189 bp replicase gene fragment revealed three main clusters: genogroup I, genogroup II and a potential novel group, designated JS, which clustered between genogroup I and genogroup II. The putative JS group, represented by phages, WWTP1_50 and 2GI13, had a >40% sequence diversity in the 189 bp replicase gene sequence with strains from genogroups I and II. As these strains were isolated from geographically different regions (Massachusetts and South Carolina) Vinje et al (2004) proposed that JS may form a stable lineage and suggested that further genomic sequence and serological data were needed to confirm whether these strains form a novel genogroup or whether these strains are the result of recombination or rearrangement events (Chetverin, 1999). In the present study complete genome sequences of two additional phages, belonging to the putative JS group (Sobsey et al., 2006; Love et al., 2008), were determined allowing a phylogenetic study into the nature of this proposed subgroup.

Whether or not the putative JS subgroup represents a novel genogroup, recombinations (sequence exchange and rearrangements) between RNA molecules may have occurred in these viruses. Largely responsible for the diversity of RNA viruses (Lai, 1992) RNA-RNA recombination has been shown to occur in several positive-sense, ssRNA human and animal viral taxa including caliciviruses, coronaviruses, hepatitis, dengue, enteroviruses and astroviruses (Cristina and Colina, 2006; Pantin-Jackwood et al., 2006; Holmes et al., 1999;

[Oberste et al., 2004](#); [Banner and Lai, 1991](#); [Oprisan et al., 2002](#); [Walter et al., 2001](#); [Jiang et al., 1999](#); [Belliot et al., 1997](#)).

RNA recombination events occur, in some cases, when two or more strains infect the same host. Proposed models for the formation of novel RNA sequences include (i) cleavage and ligation in RNA molecules or RNA secondary structures ([Lutay et al., 2007](#)), (ii) replicative template switching whereby the RNA-dependent RNA polymerase (replicase) switches from one template to another RNA template, also known as copy choice ([Chetverin, 1999](#); [Chetverin et al., 2005](#)), and (iii) RNA transesterification reaction that occurs when the polymerase adds a separate RNA fragment to the 3' terminus of the original RNA template ([Chetverin, 1999](#))

The first indication of non- replicative RNA recombination in a male-specific FRNA phage was reported by [Munishkin et al., \(1988\)](#) who found small, nonhomologous, recombinant RNA molecules produced *in vitro* in a purified template-free Q β replicase molecule. These investigators noted similar RNA molecules were present in *E. coli* cells infected with phage Q β . [Chetverin et al \(2005\)](#), studied this phenomenon by observing the formation of novel sequences in a colony of RNA molecules, suggested that this recombination event occurred as a transesterification reaction catalyzed by a conformation acquired by Q β replicase during RNA synthesis ([Chetverin et al., 2005](#); [Chetverin, 1999](#)). Nucleotide sequences of recombined RNA molecules were non-homologous to the parent RNA and were formed in the absence of DNA intermediates, demonstrating an RNA

recombination mechanism in the presence of Q β replicase ([Chetverin, 1999](#)). Therefore, it is plausible to have recombination in environmental ssRNA male-specific coliphage (*Leviviridae*) isolates.

Two JS strains, DL52 and DL54, were isolated during an environmental genotyping study of *Leviviridae* FRNA phages ([Sobsey et al., 2006](#); [Love et al., 2008](#)). These phages were placed into the putative JS subgroup using the genotyping methods of Vinjé et al ([2004](#)). The objective of the present study was to determine whether the existence of a novel JS-like subgroup representing a third *Levivirus* cluster as proposed by Vinjé et al ([2004](#)) could be verified. The approach was to analyze recently generated nucleotide and amino acid sequence data from entire genomes of 10 levivirus group I strains, 5 levivirus group II strains and two JS group strains. Analysis of the novel JS strains should provide evidence as to whether or not these *Levivirus* strains were genogroup I, II, a combination of groups I and II (recombinant strain) or a unique genogroup. To further understand the composition of these JS strains, complete genomic sequencing, amino acid composition and phylogenetic analyses were examined.

Purpose

- Evaluate the genomic nucleotide and amino acid sequences to determine the uniqueness of JS strains and to clarify why JS strains do not hybridize to group I or group II reverse-line blot hybridization probes.

Approach

- Sequence the two JS-like strains, DL52 and DL54, to determine their taxonomic classification.
- Map Open Reading Frames and determine gene locations.
- Translate nucleotide sequences into amino acid compositions for each protein.
- Compare JS strains to *Levivirus* groups I and II.
- Analyze similarities and differences between JS and groups I and II using various bioinformatic methods.

Materials and Methods

Coliphage Isolates and Propagation

FRNA phage strains CICEET 29 and CICEET 24 were isolated and placed into the putative JS subgroup by Love et al., (2008; Sobsey et al., 2006) using the genotyping methods of (Vinjé et al., 2004). CICEET 29, renamed DL52, was isolated from estuarine waters in Rachel Carson W Reserve (Beaufort), NC, and CICEET 24, renamed DL54, and was isolated from Naragansett Bay, RI.

Sequencing was performed on plaque-purified coliphages which were enriched using *Escherichia coli* HS(pFamp)R as host (Vinjé et al., 2004). Overnight enrichments were centrifuged (3,220 X g for 10 min) to pellet host cells and debris. A 1:1 (V/V) chloroform to virus supernatant mixture was vortex mixed and centrifuged again. Approximately 1-2 ml aliquots of the supernatant containing the phage enrichments were stored frozen at -75°C .

Coliphage titers were determined using a single agar layer procedure (SAL) (US EPA Method 1602, 2001). The procedure was as follows. A 1 ml of overnight *E. coli* Famp host culture was transferred into 50 ml trypticase soy broth with streptomycin and ampicillin (TSB/strep/amp) and grown 4 hr to log phase. A 150 ml volume of trypticase soy agar, TSA (4.5 g TSB and 1.2 g agar), was autoclaved and placed into a water bath ($47-55^{\circ}\text{C}$). A 300 ul aliquot of 500X strep/amp was added into the cooled 150 ml TSA. Serial 10-fold dilutions of the purified virus were prepared. For 10-fold stock dilution, 100 ul of stock was transferred into 900 ul phosphate buffered saline (PBS), mixed and this process repeated to a serial dilution 10^{-15} . To sterile 15 ml plastic tubes, labeled -1 to -15, 1 ml of the 4 hr *E. coli*, 100 ul of the appropriate virus dilution and 10 ml TSA were added. After quick manual mixing the

tube contents were poured into labeled 20mm petri plates which were then cooled, allowed to dry, inverted and incubated overnight at 37 ° C. The coliphage titer was determined by counting the plates having a minum of 5 to a maximum of 20 plaques per plate. Titer concentration was reported as PFU/ml.

Coliphage RNA Isolation

Coliphage RNA was extracted from the purified viral supernatant using a QIAamp viral RNA mini kit (Qiagen, Valencia, CA) as follows. A 200 ul volume of purified virus stock was added into the 25 ul protease tube. Into the same tube, 200 ul of carrier RNA/Buffer AL mixture was added and the combined solutions were vortex mixed, incubated 56 ° C for 15 minute and clarified by a 1 minute centrifugation. To the same tube, 250 ul ethanol (EtOH) was added, vortex mixed and incubated 5 min at room temperature (RT). The lysate was applied into the QIAamp spin column and centrifuged 3-4 min at 6000 xg (8000 rpm) and the contents of the collection tube was discarded. To the column, 500 ul of Buffer AW2 was added, centrifuged 1 min and the contents of the collection tube was discarded. To the column, 500 ul EtOH was added, centrifuged 1 min and collection tube was discarded. The column was transferred to a clean collection tube and centrifuged for 1 minute. The column was again transfered a new, nuclease-free tube and 50 ul RNase free water was added onto column and incubated 5 min at RT. The final centrifugation, (14,000 rpm) for 1 min and the recovered purified RNA was frozen at -20 °C.

Generating cDNA from Polyadenylated RNA

Phage MS2 serves as a positive, procedural control. Viral RNA was 3' polyadenylated with yeast PolymeraseA and 25 mM ATP in a 50 ul reaction volume (USB, Inc, Cleveland,

OH). The reaction was prepared with 10 ul 5X Reaction Buffer, 10 ul RNA, 2 ul 25 mM ATP, 0.7 ul 600 U Poly(A)Polymerase and 27.3 ul nuclease-free water. The mixture was incubated at 37⁰ C for 5 min and placed on ice for enzymatic termination. Polyadenylated RNA was either immediately frozen or used as a template for cDNA.

Full-length cDNA was prepared using oligo-dT reverse primer supplied with the reverse transcriptase MonsterScript 1st Strand cDNA Synthesis Kit (EpiCentre, Madison, WI) or with a gene-specific reverse primer. To a 250 ul thin-walled PCR tube, the following reagents were added: 4 ul nuclease-free water, 10 ul polyadenylated RNA or RNA template and 1 ul of 10 uM PolyT primer or 1 ul of 10uM gene-specific primer. The mixture was heated for 1 min at 65 ° C and chilled for 1 min on ice. To the same tube, 1 ul MonsterScript Reverse Transcriptase and 4 ul of 5X cDNA reaction buffer were added. The mixture was placed in a thermocycler with the following cycle regime: 37 ° C for 5 min, 42 ° C for 5 min, 60 ° C for 40 min and 90 ° C for 5 min and chilled on ice for 1 min. The single-stranded cDNA was either frozen or used for PCR template.

5' Amplification of cDNA Ends

The nucleotide sequence of the 5' region was determined by rapid amplification of cDNA end (RACE) with the Smart Race cDNA Amplification Kit (Clontech, Mountain View, CA). First-strand cDNA synthesis was prepared on ice in a 250 ul thin-walled PCR tube by combining 3 ul RNA, 1 ul of 10uM gene-specific reverse primer and 1 ul Smart oligo (from kit). The 5 ul reaction volume was briefly centrifuged and the following components were added: 2 ul of 5X First Strand buffer, 1 ul of 20 mM DTT, 1 ul of 10 mM dNTP and 1 ul SuperScript II (Invitrogen, Carlsbad, CA). Following a brief centrifugation, the mixture was

incubated for 90 min at 42 °C. To dilute the first-strand cDNA, 20 ul of Tricine-EDTA buffer was added and heated for 7 min at 72 °C. The reaction generated double stranded cDNA. The cDNA was frozen at - 20 °C and used for subsequent PCR reactions.

Concentration of cDNA was determined with a NanoDrop spectrophotometer (NanoDrop Technologies, Wilmington, DE).

PCR, Cloning and Sequencing

The cDNA was amplified by using Phusion DNA Polymerase (New England Biolabs, Ipswich, MA), with final concentrations of 1X of 5X Phusion Buffer, 0.2 mM dNTP, 1 ul of 10 uM forward primer, 1 ul of 10uM reverse primer, 3% DMSO, 2 ul cDNA and 0.5 ul Phusion Taq in a 50 ul reaction using the following cycle parameters: one round denaturation at 98 °C (1 min), 35 rounds at 98 °C (30 sec), 48 °C (1 min), 72 °C (3 min) followed by 10 min extension at 72 °C. For each reaction, positive controls were prepared using primers MJV82 and JV81 for *Leviviruses* and MJV82 and JV41 with *Alloleviviruses*. A no-template negative control was included.

To verify the generation of full-length cDNA, the 5' end of phage MS2 was PCR amplified using primers MS25 and MS23 and the same protocol as stated above ([Lovmar *et al.*, 2003](#)).

PCR products were separated by electrophoresis in a 1.5% agarose gel, stained with SYBR Gold Nucleic Acid Gel Stain (Molecular Probes, Carlsbad, CA) and visualized under blue light (Dark Reader Transilluminators, Clare Chemical Research, Dolores, CO).

Blunt-end PCR products were excised using a gel extraction tool (USA Scientific Plastics, Ocala, FL) and then purified (QuickClean 5M Gel Extraction Kit, GenScript Corporation, Piscataway, NJ).

Excised bands were weighed in 1.5 ml microcentrifuge tubes and 3 volumes of Binding Solution II per gel slice were added. The gel solution was heated at 50 ° C until melted. One volume of isopropanol was added and the mixture was transferred into the Genprep spin column and centrifuged for 1 min at 12,000 rpm. The column effluent was discarded and 500 ul of Wash Buffer was added, the column was centrifuged and the liquid waste discarded. The 500 ul wash was repeated and waste discarded. The column was placed into a clean 1.5 ml microcentrifuge tube, 30 ul of Elution Buffer was added and the column was incubated 1 min at room temperature. The tube was centrifuged 1 min at 12,000 rpm and the collected DNA eluate was transferred to a clean tube. Concentration of the DNA was determined using a NanoDrop spectrophotometer. The DNA was either cloned or the PCR product was sequenced.

Gel-purified DNA was cloned using a ZeroBlunt TOPO Cloning Kit, pCR-Blunt II TOPO plasmid kit (Invitrogen, Carlsbad, CA). To prepare a TOPO cloning reaction, 4 ul DNA, 1 ul salt solution and 1 ul pCR II Blunt TOPO were added to a nuclease-free tube and incubated 30 min at room temperature. To a vial of One Shot *E. coli* competent cells, 2 ul of the TOPO cloning reaction were added and incubated on ice for 30 min. The cells were heat-shocked at 42 ° C and immediately placed on ice. A 250 ul volume of SOC medium was added to the One Shot cells and shaken (200 rpm) for 1 hr at 37 ° C. Fifty ul of transformed cells were plated onto pre-warmed LB agar plates containing 50 ug/ml kanamycin, the

transformed cells were spread to isolate colonies and the plates were incubated overnight at 37 ° C.

Colonies of transformed *E. coli* cells were screened for positive inserts using whole-cell PCR. Using aseptic techniques, individual colonies were selected with a sterile toothpick, the toothpick was briefly rinsed into a 50 ul Phusion master mix (as described above) then dropped into 10 ml LB/kan broth and incubated overnight at 37 ° C. Whole-cell PCR was performed on individual colonies using Phusion DNA Polymerase with the following cycle modifications: one round denaturation at 98 °C (3 min), 35 rounds at 98 °C (10 sec), 57 °C (30 sec), 72 °C (30 sec) followed by 10 min extension at 72 °C. Amplicons were separated by electrophoresis in 1.5% agarose gel in 0.5X Tris-acetate-EDTA (TAE), stained with 20 ug/ml ethidium bromide and visualized under UV light (UVP, Upland, CA). Those clones with the appropriate size PCR amplicon were selected for plasmid purification.

Positive clones were plasmid-purified (QIAprep Spin Miniprep Kit, Qiagen, Valencia, CA). An *E. coli* colony that had been selected with a toothpick and incubated overnight was centrifuged 10 min at 8000 rpm (6800 xg). The supernatant was discarded and the cell pellet was resuspended and processed as follows. A 250 ul volume of Buffer P1 was added to the cell pellet, vortexed to mix and transferred to a clean 1.5 ml microcentrifuge tube. A 350 ul volume of Buffer P2 was added to the resuspended pellet and mixed by inversion followed by addition of 350 ul Buffer N3. The buffer mixture was inverted 4-6 times and centrifuged 10 min at 13,000 rpm (17,900 xg). The supernatant was decanted into the QIAprep spin column and centrifuged 1 min at 13,000 rpm. The column effluent was discarded and the column was washed with 500 ul Buffer PB, the column was centrifuged 1 min and the wash was

discarded. To the spin column, 750 ul Buffer PE was added, the column was centrifuged and the wash was discarded. The column was incubated with 50 ul Buffer EB to elute the plasmid. Following a 1 min incubation at room temperature, the column was centrifuged 1 min. Purified plasmid was shipped frozen for sequencing (Sequetech, Mountain View, CA). Each cDNA PCR amplicon was cloned and sequenced in triplicate to account for errors. In some cases, the PCR product was sequenced directly. To achieve publication quality data, both forward and reverse strands were sequenced (Sequetech, Mountain View, CA). This process was repeated until complete genomes were obtained.

To avoid contamination, a PCR hood (AirClean 600, AirClean Systems, Raleigh, NC) located in a designated room was used to prepare master mixes separate from template additions. PCR amplification, electrophoresis, template and/or viral preparations ([EPA, 2004](#)) were conducted in individual assigned rooms based on designated use.

Primer Design

PCR amplicons were first generated from an approximate 200 nt region using degenerate primers (MJV82 and JV81 for *Leviviruses*) specific to the replicase region ([Vinjé et al., 2004](#)) and sequenced (UNC, NC). The combination of these primer sets was termed “generic PCR” as the primers were genus specific but not genogroup specific. A strain-specific forward primer was designed from this 200 nt region. A combination of the strain-specific forward primer and a PolyT reverse primer amplified an approximate 1 kb PCR fragment. The amplicon was cloned and sequenced. As sequences of the fragments were generated, reverse primers were designed to amplify overlapping sections of the genome. Forward primers were designed by alignment (BioEdit v7.0.1 Clustal W application; [Hall,](#)

1999) of representative group I (J20, M12) and II (KU1) strains sequenced in this study or available in GenBank.

Sequence Analyses and Open Reading Frames

JS sequences were compared to nucleotide and/or amino acid sequences from 10 group I strains (MS2, DL1, DL2, DL13, DL16, ST4, R17, J20, M12, fr) and 5 group II strains (T72, DL10, DL20, GA, KUI). Sequences and/or amino acids were aligned using BioEdit v7.0.1 ClustalW application (Hall, 1999). Basic Local Alignment and Search Tool (BLAST) finds regions of local similarity between sequences and is used to search similar matches in the National Center for Biotechnology Information (NCBI) genetic database. BLAST analyses for sequence and phylogenetic confirmation were performed on each individual FRNA clone or PCR fragment. Open Reading Frames (ORF) were determined using BioEdit (Hall, 1999).

Nucleotide percent similarity and dendograms were constructed using Bionumerics software v. 3.5 (Applied Maths, Saint-Martens-Latem, Belgium). Phylogenetic trees were built using the global cluster analysis performed on multiple aligned sequences and clustered by UPGMA using the Jukes and Cantor correction. A bootstrap analysis, based on 10,000 substitutions, was used to measure cluster significance.

Cluster analysis of Levivirus Groups I, II and JS phages were generated from pairwise similarities of the amino acid sequences of their replicase genes (Bionumerics). Standard deviations of the average similarities of the clusters were determined using Bionumerics. The resulting phylogenetic tree produces a cophenetic correlation which represents the faithfulness of the clusters expressed on a percentage basis (Bionumerics).

Amino Acid Analysis

Amino acid compositions for each of the four genes were determined using a computer-generated DNA-to-protein translation tool, ExPASy (<http://ca.expasy.org/>). Prediction of protein sequence motifs were identified by PROSITE (<http://ca.expasy.org/>).

Amino acid sequence data were analyzed using BioNumerics Software v.3.5 (Applied Maths, Saint-Martens-Latem, Belgium). Phylogenetic trees were built by global cluster analysis performed on multiple aligned sequences and clustered by UPGMA using the Jukes and Cantor correction (Jukes & Cantor, 1969). Cophenetic correlations and cluster Cutoff method were employed to measure faithfulness and relevancy of the clusters. Average similarities with standard deviations were calculated for the relevant clusters.

Relationships among aligned amino acid sequences were depicted in similarity plots generated by SimPlot, v3.5.1 (Lole et al., 1999). The SimPlot program determines the percent identity between a reference and the queried sequence. Percent similarity was calculated within a sliding window of 160 bp wide with a step size between plots of 10 bp.

Results

Degenerate replicase primers in an RT-PCR assay identified the two strains, DL52 and DL54, to the *Levivirus* genus. Nucleotide sequences of the 189 bp amplicon classified the strains as JS-like (Vinjé et al., 2004). Reverse-line blot hybridization failed to genotype the two strains into subgroups I or II.

A total of 17 strains (MS2, ST4, DL1, DL2, DL13, DL16, R17, M12, J20 and fr in genogroup I, DL52 and DL54 in the JS genogroup and T72, DL10, DL20, GA and KU1 from

genogroup II) were used to examine the relationships among nucleotides and amino acids in the *Levivirus* genus. The first 9 strains in genogroup I are referred to as “MS2-like.”

With respect to nucleotide sequences, the MS2-like strains shared 91.68-99% full-length genome nucleotide sequence similarity to each other (Fig 5.2) and the two JS strains, DL52 and DL54, were 96.73% similar in nucleotide sequences to each other (Table 6.2). The JS strains were more similar to MS2-like genogroup I FRNA coliphage (80-84 % sequence similarity) than the fr strain was to MS2-like phages (75.27-77.65% sequence identity). Despite their sequence similarities, genome lengths for JS strains (3525 nt) were shorter than the MS2-like group I (3569-3575 nt) (Table 6.1) but longer than genogroup II genomes (3458-3486 nt) (Table 5.1). Numerous deletions in the 3' untranslated region and a portion of ORF4 (replicase) in JS strains accounted for the decreased length (Appendix C).

Within genogroup I, the amino acid sequences of all four proteins of strain fr were distinctly different from the proteins of the MS2-like strains (Fig 5.4A, Fig 6.2). A different pattern, however, emerged when comparing the sequences of the four proteins of the JS strains to the protein sequences of the MS2-like genogroup I strains. Sequence similarities of the maturation, capsid and lysis proteins of the JS strains were very similar to those of the MS2-like group strains, sharing 99-100%, 98-100% and 95-100% sequence similarities, respectively (Table 6.3, Fig 6.1). However, the replicase protein sequences of the JS strains were quite dissimilar to the replicase protein sequences of the MS2-like genogroup I strains, displaying a similarity range of 84-85%. In contrast, a similarity of 97-99% was observed among the highly conserved replicases of the nine MS2-like strains. Strain fr shared an 80% replicase similarity to JS strains and approximately 88-90% similarity to MS2-like strains

(Table 6.3). Group II replicase was approximately 50-53% similar to JS strains and to the other genogroup I strains (Table 6.4).

Table 6.1 JS strains and genogroup I. Open Reading Frame positions and genome lengths of FRNA coliphage JS strains DL52 and DL54 compared to genogroup I strains.

		Open Reading Frame Location (nt)				
Strain	Group	Full length nt	ORF1	ORF2	ORF3	ORF4
DL1	I	3570	130-1311	1335-1727	1678-1905	1761-3398
DL2	I	3491 ^b	130-1311	1335-1727	1678-1905	1761-3398
DL13	I	3491 ^b	130-1311	1335-1727	1678-1905	1761-3398
DL16	I	3569	130-1311	1335-1727	1678-1905	1761-3398
J20	I	3569	130-1311	1335-1727	1678-1905	1761-3398
ST4	I	3569	130-1311	1335-1727	1678-1905	1761-3398
R17	I	3569	130-1311	1335-1727	1678-1905	1761-3398
MS2 ^a	I	3569	130-1311	1335-1727	1678-1905	1761-3398
DL52	JS	3525	130-1311	1335-1727	1678-1905	1761-3398 ^c
DL54	JS	3398 ^b	130-1311	1335-1727	1678-1905	1761-3398 ^c

^a GenBank prototype strain

^b nearly full-length

^c deletions and insertions in JS ORF4

nt - nucleotide

Table 6.2 Pairwise comparison JS strains and genogroup I. Pairwise comparisons of nucleotide percent similarity.

(A) JS strains and group I (B) JS strains and group II.

(A) Group I and JS strains.

	DL52	DL54
<u>Strain</u>		
DL52	100	
DL54	96.73	100
DL1	81.48	81.87
DL16	85.41	84.72
ST4	80.30	80.11
R17	80.55	80.53
J20	82.00	82.01
MS2	80.12	80.01
fr	69.18	69.06

(B) Group II and JS strains.

	DL52	DL54
<u>Strain</u>		
DL52	100	
DL54	96.73	100
T72	53.96	53.53
DL10	54.07	53.89
DL20	52.87	52.65
GA	52.44	52.29
KU1	52.94	52.66

The replicase protein of all genogroup I strains including the JS subgroup was shown to be 545 amino acids in length ([Appendix C](#)). The JS replicase protein was, however, distinctive from the MS2-like replicase protein as it possessed a single amino acid insertion at position 467 and a one amino acid deletion at the 3' termini of the stop codon ([Appendix C](#)). The catalytic domain of the replicase protein was in the same location, between amino acid positions 243-373. In the JS replicase proteins from amino acid position 455 and continuing to the 3' end, the sequences of the JS replicase protein diverge from the parental MS2-like strains and were unique in composition ([Fig 6.3](#), [Appendix C](#)) resulting from a frame shift having a two nucleotide insertion ([Fig 6.5](#)).

A nucleotide alignment revealed numerous deletions in the JS strains when compared to the other genogroup I strains. Beginning approx 40 nt downstream of the replicase ORF4 stop codon and continuing to the 3' termini, 53 nt deletions were observed in the JS strains. JS strains, however, share the 3' “signature”, ACCACCCA, sequence with groups I and II *Levivirus* ([Appendix C](#)).

A cluster analysis of the amino acids sequences from the replicase proteins of group I, JS subgroup and group II strains was performed. Cophenetic correlations showed the MS2-like strains including strain fr, the JS subgroup strains, and the group II strains all formed faithful clusters with correlations of 100, 90 and 98, respectively. The cluster cutoff method, however, showed only two relevant clusters, genogroup I strains, which included fr and JS, and genogroup II strains ([Fig 6.4](#)).

Table 6.3 Pairwise comparison of amino acids for JS and genogroup I. Percent similarity in amino acid

[illegible][illegible][illegible]

Table 6.4 Pairwise comparison of replicase protein JS and genogroup I. Percent similarity in amino

sequences between JS strains and genogroups I and II replicase protein. Pairwise alignments were performed in Bionumerics.

Group I

ST4	100.00
R17	99.06 100.00
MS2	99.03 98.57 100.00
DL2	97.51 97.13 97.53 100.00
DL13	97.39 97.01 97.41 99.89 100.00
DL16	97.22 96.84 97.25 99.73 99.61 100.00
DL1	97.99 97.61 97.79 99.41 99.29 99.13 100.00
J20	97.67 97.20 97.47 98.99 98.87 98.71 99.13 100.00
fr	88.51 88.38 88.17 89.04 88.90 88.71 88.89 88.81 100.00

Group JS

DL52 JS	84.40 83.86 83.95 85.41 85.38 85.18 85.33 84.81 79.70 100.00
DL54 JS	84.15 84.12 83.93 85.05 85.02 84.82 85.03 84.81 79.81 97.20 100.00

Group II

T72	51.90 51.54 51.09 52.61 52.61 52.29 52.64 52.69 52.75 52.85 53.11 100.00
KU 1	51.16 50.80 50.35 51.90 51.91 51.57 51.93 51.97 52.49 52.54 52.59 97.13 100.00
DL10	52.67 52.31 51.87 53.37 53.37 53.05 53.39 53.66 52.74 53.68 53.83 93.29 93.02 100.00
DL20	52.70 52.34 51.90 53.43 53.43 53.11 53.22 53.71 53.22 53.35 53.50 93.77 93.41 98.62 100.00
GA	51.27 50.90 50.45 52.01 52.01 51.68 51.80 52.30 51.80 52.47 52.62 92.43 92.27 97.73 98.57 100.00

Figure 6.1 SimPlot analysis of genome nucleotide profile strain MS2 and DL52.

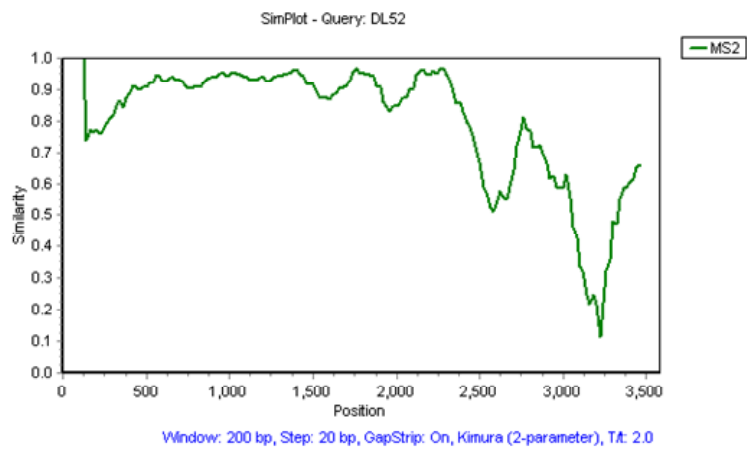


Figure 6.2 SimPlot analysis of genome nucleotide profile strain MS2 and fr.

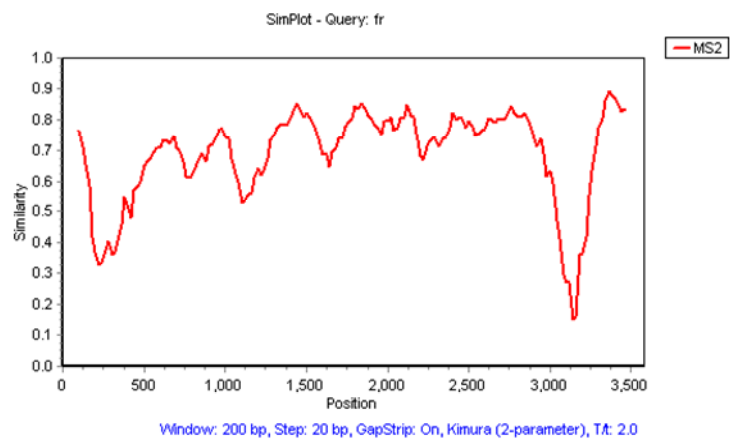


Figure 6.3 SimPlot analysis of replicase amino acid profile strain MS2 and DL52.

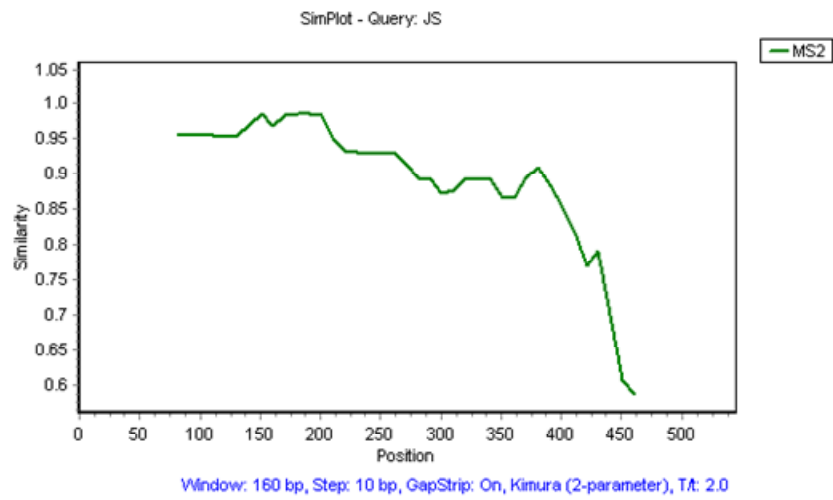
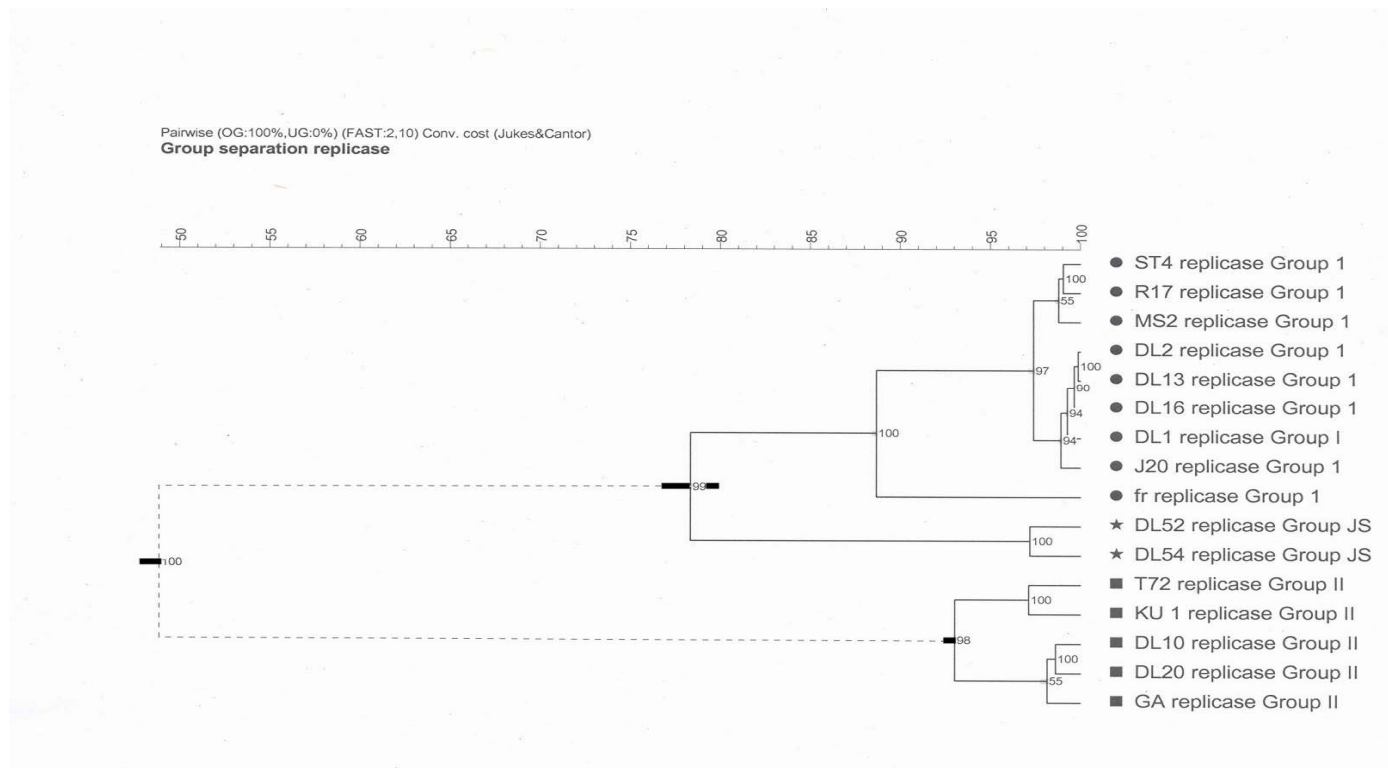
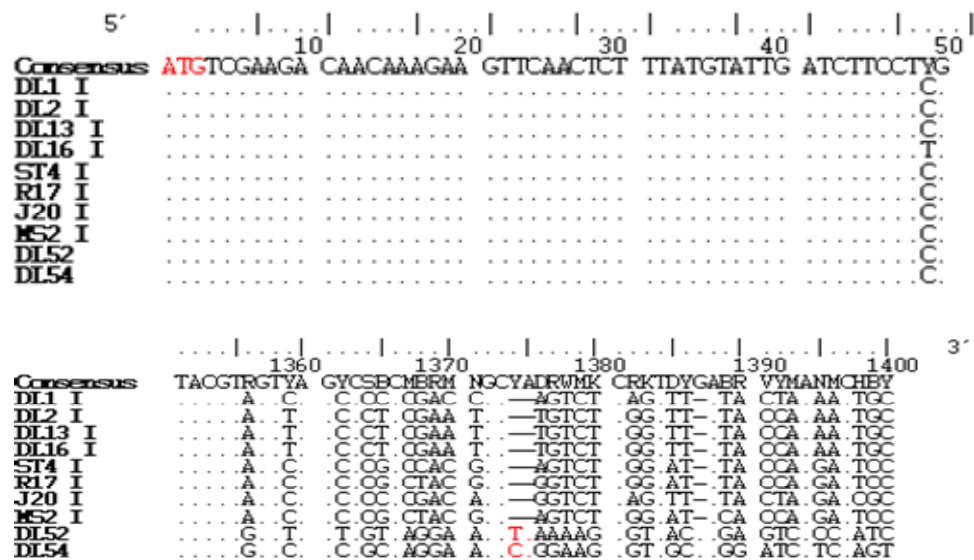


Figure 6.4 Phylogenetic analysis of replicase amino acids groups I, II and JS.

Cluster analysis of Levivirus Groups I and II phages generated from pairwise similarities of the amino acid sequences of their replicase genes. Horizontal bars at three of the branches show the standard deviations of the average similarities of the clusters. Numbers at each branch are the cophenetic correlations which represent the faithfulness of the clusters. Two relevant clusters, as determined by the cluster Cutoff method, are grouped to the right of the dashed lines.



Replicase frame shift in two JS strains when compared to genogroup I strains. Alignment (BioEdit v7.0.1) of the replicase nucleotide sequences from genogroup I strains DL1, DL2, DL13, DL16, ST4, J20, MS2 with JS strains DL52 and DL54. For clarity, only a portion of the alignment is shown. Alignment of each genogroup is depicted in discontinuous blocks to illustrate the nucleotide position. The numbers along the top are the nucleotide positions. Genome sequences read 5' - 3' direction. Dots indicate identity with the consensus sequence. Degenerate bases are noted in the standard IUB codes. The replicase start codon and two nucleotide insertions are highlighted in red.



Discussion

The JS strains and fr diverged from the MS2-like reference strains but in different ways. Across the entire genome strain fr consistently differed from the MS2-like strains (Fig 6.2). However, with JS strains, major differences were only observed downstream of the catalytic site in the 3' end of the replicase gene and the adjacent noncoding region, suggesting a specific genetic rearrangement or recombination event (Fig 6.3).

Cophenetic correlations strengthen the possibility that JS strains are recombinants as the JS is only a subgroup of genogroup I and not a novel genogroup. Throughout the *Leviviridae* family, subgroups emerge within genogroups, however, as with strain fr, subgroup strains differ in all four genes from the parent genogroup (Fig 5.4A, B).

Genetic exchange in ssRNA viruses was first demonstrated in polioviruses (Hirst, 1962; Ledinko, 1963). Subsequent experiments with ssRNA coliphage mutants failed to provide evidence for recombination. Horiuchi (1975) concluded that RNA phages would not undergo recombination. Those attempts to detect recombination occurred in the time when FRNA phages were thought to possess only three genes, not four. In all likelihood, laboratory-applied selective pressure failed to detect or generate a specific recombinant and may not necessarily reflect the lack of recombination or responsible mechanisms that could occur under conditions that better represent the natural history and ecology of these ssRNA coliphages. Eventually, ssRNA recombination was demonstrated in a *Leviviridae* coliphage Q β replicase (Munishkin et al., 1988).

Recombination events sometimes alter the RNA polymerase region. Human Noroviruses, a positive sense ssRNA virus with a genome length of 7400-8300 nt, are

considered to belong to a prototype strain if they share approximately 85% overall nucleotide sequence identity and a high amino acid sequence identity (> 95%) to the RNA polymerase gene ([Jiang et al., 1999](#)). The naturally occurring human Norovirus strain shared 95% amino acid sequence identity with the capsid sequences from a Mexico cluster and 95% amino acid identity to the RNA polymerase in a Lordsdale virus cluster. Sequences from the natural strain were obtained from one viral isolate. The combination of sequences in the one strain being complementary to two distinct human Norovirus clusters led to the proposition that this strain was a naturally occurring recombinant ([Jiang et al., 1999](#)).

Genetic recombination is known to occur in certain Enteroviruses, a positive ssRNA virus having an approximate 7500 nt genome. Poliovirus recombination occurs in vaccine-derived strains ([Kew et al., 2002](#)) in the human population as a single infected individual excretes a high proportion of recombinants ([Oprisan et al., 2002](#)). To determine if other enteroviruses undergo natural recombination, isolates of echoviruses were collected from a meningitis outbreak. Nucleotide sequences were clustered based on a capsid protein (VP1) and RNA polymerase (3D). Dendrogram relatedness of the echovirus strains grouped the VP1 sequences to the prototype strains. However, the RNA polymerase sequences did not cluster to the prototype strains, suggesting genetic recombination among the outbreak strains ([Oprisan et al., 2002](#)).

Human astroviruses are positive sense, ssRNA with a genome length of approximately 6,800 nucleotides ([Walter et al., 2001](#)) and a polyadenylated 3' tail ([Belliot et al., 1997](#)). Two sets of strains were investigated for recombination; one set was identified from a child care center in Houston, TX and the two other strains originated in stool samples from two children

in Mexico City. The pool of strains shared >97% nucleotide sequence similarity in two out of three genomic regions. The novel strain clustered to one group based on the capsid region. When the RNA polymerase gene was analyzed, the novel strain clustered to a separate human astrovirus group. The strains were identified as naturally occurring recombinants on the evidence of high sequence similarity to a few genes of one prototype and similarity to different genes in a second prototype. A total of 64 additional human astroviruses lacked these novel traits ([Walter et al., 2001](#)).

Evidence for recombination among positive ssRNA viruses exists within the RNA-dependent RNA polymerase. Turkey astrovirus is a non-enveloped, positive sense ssRNA virus with a polyadenylated 3' tailed genome of approximately 7kb. Astroviruses are associated with enteric disease ([Pantin-Jackwood et al., 2006](#)). The most conserved gene in the avian and mammalian astrovirus is the RNA-dependent RNA polymerase. Genetic analysis of capsid and polymerase sequences from twenty-three turkey astrovirus strains resulted in 8 clusters for the capsid gene and two phylogenetic clusters for the RNA polymerase gene. Computer-generated analyses identified polymerase gene recombination in strains of turkey astrovirus ([Pantin-Jackwood et al., 2006](#)).

Numerous reports of positive sense ssRNA viral recombinants are documented in the scientific literature ([Cristina and Colina, 2006](#); [Pantin-Jackwood et al., 2006](#); [Holmes et al., 1999](#); [Oberste et al., 2004](#); [Banner and Lai, 1991](#); [Oprisan et al., 2002](#); [Walter et al., 2001](#); [Jiang et al., 1999](#); [Belliot et al., 1997](#)). Virus strains are classified as natural recombinants when one virus strain is complementary to two different proteins or stretch of nucleotide sequences originating from two genetically distinct clusters or in other rearrangements events

(Chetverin, 1999). In this study, two JS strains shared >95% amino acid identity in three (maturation, capsid and lysis) *Levivirus* genes but only 84-85% amino acid identity to the otherwise highly conserved replicase protein. A nucleotide frame shift occurred downstream of the catalytic site in the replicase gene thereby accounting for the lack of nucleotide or amino acid similarity between the JS strains and the genogroup I replicase. Therefore, it is plausible to propose natural recombination in these two FRNA coliphages.

Conclusion

Phylogenetic tree analysis produced a cophenetic correlation which showed 1) ten genogroup I strains, including strain fr, 2) the JS subgroup strains, and 3) the genogroup II strains all formed faithful clusters with correlations of 100, 90 and 98, respectively. The cluster cutoff method, however, showed only two relevant clusters, 1) genogroup I strains, which included fr and JS, and 2) genogroup II strains (Fig 6.4). Therefore, the novel JS strains are not a unique *Levivirus* genogroup. The proposed classification of JS strains is a genogroup I subgroup “JS-like”.

The results of this study provide molecular genetic evidence indicative of recombination in two JS strains of FRNA coliphages. There was high nucleotide and amino acid identity in three genes, the maturation, capsid and lysis genes ($\geq 95\%$) but a lack of similarity in the replicase gene. A nucleotide frame shift occurred downstream of the replicase catalytic. Therefore, the catalytic site was conserved resulting in viable progeny.

Each JS strain was isolated from two different geographical states, North Carolina and Rhode Island, and the remaining group I strains were collected from other geographical locations across the USA and Germany (Table 7.1). Geographical distinctness does not play a

role in sequence variation within the *Leviviridae* family ([Table 7.1](#), [Appendix A](#)).

Recombination may explain why *Leviviridae* strains circulate as discrete sub-groups independent of geographical location.

Although the two JS strains were sequenced in the same laboratory, these strains were field-collected by different investigators and shipped to another location where they were plaque-purified and preliminarily classified. Therefore, the possibility that contamination resulted in false recombinants seems unlikely. This is the first description of possible recombinant strains from natural isolates in ssRNA *Leviviridae* bacteriophages.

Summary

- Two novel JS strains, DL52 and DL54, were initially isolated from North Carolina and Rhode Island, respectively.
- Three genes, maturation, capsid and lysis, shared > 95% amino acid similarity when JS strains were compared to nine MS2-like genogroup I strains.
- Among nine MS2-like strains, replicase amino acid similarity was highly conserved (97-99 % amino acid identity).
- In contrast, JS strains shared 85% replicase amino acid identity to group I MS2-like strains, 80% to group I fr and 53% to group II strains.
- High (> 95%) amino acid identity in three genes and a lack of similarity in the otherwise highly conserved replicase gene along with a statistical cluster cutoff analysis suggests the JS strains are recombinants.

References

- Banner LR and MMC Lai. 1991. Random nature of coronavirus RNA recombination in the absence of selection pressure. *Viol*, 185:441-445.
- Belliot G, H Laveran, and SS Monroe. 1997. Detection and genetic differentiation of human astroviruses: phylogenetic grouping varies by coding region. *Arch Virol*, 142:1323-1334.
- Chetverin AB. 1999. The puzzle of RNA recombination. Minireview. *FEBS Lett*, 460:1-5.
- Chetverin AB, DS Kopein, HV Chetverina, AA Demidenko and VI Ugarov. 2005. Viral RNA-directed RNA polymerases use diverse mechanisms to promote recombination between RNA molecules. *J Biol Chem*, 280,8748-8755.
- Cristina J and R Colina. 2006. Evidence of structural genomic region recombination in Hepatitis C virus. *Viol J*, 3:53
- EPA. 2004. Quality assurance/quality control guidance for laboratories performing PCR analyses on environmental samples. EPA 815-B-04-001. October 2004.
- Hall, T.A., 1999. BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucl. Acids Symp. Ser.* 41, 95-98.
- Hirst GK. 1962. Genetic recombination with Newcastle disease virus, polioviruses, and influenza. *Cold Spring Harb Symp Quant Biol*, 27:303-309.
- Holmes EC, M Worobey and A Rambaut. 1999. Phylogenetic evidence for recombination in Dengue virus. *Mol Biol Evol*, 16(3):405-409.
- Horiuchi K. 1975. Genetic studies of RNA phages. *In: RNA Phages*. N. Zinder, ed. Cold Spring Harbor Laboratory, Cold Spring Harbor, NY. pp:29-50.
- Jiang X, C Espul, WM Zhong, H Cuello and DO Matson. 1999. Characterization of a novel human calicivirus that may be a naturally occurring recombinant. *Arch Virol*, 144:2377-2387.
- Kew, O., V Morris-Glasgow, M Landaverde, C Burns, J Shaw, Z Garib, J André, E Blackman, CJ Freeman, J Jorba, R Sutter, G Tambini, L Venczel, C Pedreira, F Laender, H Shimizu, T Yoneyama, T Miyamura, H van Der Avoort, MS Oberste, D Kilpatrick, S Cochi, M Pallansch, C de Quadros. 2002. Outbreak of poliomyelitis in Hispaniola associated with circulating type 1 vaccine-derived poliovirus. *Science*, 296(5566):356-359.
- Lai, M.C. 1992. RNA recombination in animal and plant viruses. *Microbial. Rev* 56:61-79.

- Ledinko, N. 1963. Genetic recombination with poliovirus type 1 studies of crosses between a normal horse serum-resistant mutant and several guanidine-resistant mutants of the same strain. *Virology*, 20(1):107-119.
- Lole, KS., RC Bollinger, RS Paranjape, D Gadkari, SS Kulkarni, NG Novak, R Ingersoll, HW Sheppard and SC Ray. 1999. Full-length human immunodeficiency virus type 1 genomes from subtype c-infected seroconverters in India, with evidence of intersubtype recombination. *J Virol*, 73(1):152-160.
- Love, DC., J Vinjé, SM Khalil, J Murphy, GL Lovelace and MD Sobsey. 2008. Evaluation of RT-PCR and reverse line blot hybridization for detection and genotyping F+ RNA coliphages from estuarine waters and molluscan shellfish. *J Applied Microbiol*, 104:1203-1212.
- Lovmar, L., C. Fock, F Espinoza, F Bucardo, A-C Syvanen and K Bondeson. 2003. Microarrays for genotyping human Group A Rotavirus by multiplex capture and type-specific primer extension. *J Clin Microbiol* 41:5153-5158.
- Munishkin AV, LA Voronin, and AB Chetverin. 1988. An *in vivo* recombinant RNA capable of autocatalytic synthesis by Q β replicase. *Nature*, 333:473-475.
- Munishkin AV, LA Voronin, VI Ugarov, LA Bondareva, HV Chetverina and AB Chetverin. 1991. Efficient templates for QB replicase are formed by recombination from heterologous sequence. *J Mol Biol* 221:463-472.
- Oberste MS, K Maher and MA Pallansch. 2004. Evidence for frequent recombination within species *Human Enterovirus B* based on complete genomic sequences of all thirty-seven serotypes. *J Virol*, 78(2):855-867.
- Oprisan G, M Combiescu, S Guillot, V Caro, A Combiescu, F Delpeyroux and R Crainic. 2002. Natural genetic recombination between co-circulating heterotypic enteroviruses. *J Gen Virol*, 83:2193-2200.
- Pantin-Jackwood MJ, E Spackman and PR Woolcock. 2006. Phylogenetic analysis of turkey astroviruses reveals evidence of recombination. *Virus Genes*, 32:187-192.
- Sobsey MD, DC Love and GL Lovelace. 2006. F+RNA coliphages as source tracking viral indicators of fecal pollution. A final report submitted to the NOAA/UNH Cooperative Institute for Coastal and Estuarine Environmental Technology (CICEET).
- Vinjé J, SJG Oudejans, JR Stewart, MD Sobsey and SC Long. 2004. Molecular detection and genotyping of male-specific coliphages by reverse transcription-PCR and reverse

line blot hybridization. *Appl Environ Microbiol*, 70(10):5996-6004.

Walter JE, J Briggs, ML Guerrero, DO Matson, LK Pickering, G Ruiz-Palacios, T Berke and DK Mitchell. 2001. Molecular characterization of a novel recombinant strain of human astrovirus associated with gastroenteritis in children. *Arch Virol*, 146:2357-2367.

VII. A Reverse Transcription-PCR Assay to Distinguish the Four Genogroups of Male-Specific (F+) RNA Coliphages

Abstract

Identifying, managing and reducing exposure risks from fecal contamination in recreational, drinking, shellfishing and other waters and accurately assessing risk from exposure can best be attained if tools to distinguish between sources of pollution are available. The male-specific RNA coliphage (FRNA) genogroups exhibit some degree of source specificity at the human vs animal level. Reverse-transcription PCR (RT-PCR) can be effectively used for source identification if specific primer sets are designed to be capable of identifying all members within each genogroup. In this study methods were developed for a heat-release procedure that eliminated the need for RNA purification and an RT-PCR method employing genogroup-specific primers to genotype FRNA for potential use in source tracking fecal contamination. Each genogroup-specific primer set was designed from a minimum of 5 to a maximum of 10 strains of complete genome sequences per genogroup using a total genome database of 30 strains. The four genogroup-specific primer sets generated discrete PCR amplicon sizes from a variety of environmental phage strains. Cross-reactivity with strains from other genogroups was not observed.

Introduction

F-specific ssRNA (FRNA) coliphages, family *Leviviridae*, genera *Levivirus* and *Allolevivirus*, are recognized as promising indicators of both pathogenic enteric viruses and human sewage ([Grabow, 2001](#)). They generally meet the criteria for an effective indicator of sanitary quality of water ([Gerba, 1987](#); [Sobsey et al., 2005](#)). Host-specificity displayed by their four sub-groups also renders FRNA coliphages capable of providing information regarding sources of fecal pollution ([Griffin et al., 2000](#)). Group II and III phages generally originate from human waste, whereas groups I and IV are associated with animal waste ([Havelaar et al., 1986](#)). While data has not been invariably conclusive as to fecal source vs phage grouping, this trend occurs in a majority of cases ([Schaper et al., 2002](#); [Brion et al., 2002](#); [Cole et al., 2003](#); [Stewart-Pullaro et al., 2006](#)). Subgrouping was originally based on serological properties ([Sundram et al., 2006](#)). As serological typing was shown to sometimes yield inconclusive results ([Hsu et al., 1995](#); [Beekwilder et al., 1996](#)), genotyping techniques to group FRNA phage isolates were developed as an alternative approach for fecal source determination ([Hsu et al., 1995](#); [Beekwilder et al., 1996](#); [Griffin et al., 2000](#); [Vinjé et al., 2004](#)). Initially these techniques required the isolation of phages and subsequent RNA release and membrane hybridization. Recently, reverse transcription polymerase chain reaction (RT-PCR) ([Dryden et al., 2006](#)) and reverse quantitative (RTQPCR) techniques ([Kirs and Smith, 2007](#); [Ogorzaly and Gantzer, 2006](#)) for genotyping were developed. In these reports primer design was based on a limited number of complete or partial nucleotide sequences available in the National Center for Biotechnology Information (NCBI) GenBank. Thus, broad primer and probe genogroup specificity may not have been achieved due to the limited nucleotide

data from the few strains available in GenBank. In this study a design of highly precise, forward and reverse genogroup-specific, RT-PCR primers were based on a total of 30 FRNA phages of several strains from all four genogroups. Data were based upon full genomic sequences of nineteen newly sequenced FRNA coliphages along with eleven full-length FRNA coliphage sequences available in GenBank.

Purpose

- Design genogroup-specific primer sets and develop a one-step Reverse-Transcription PCR to be capable of identifying all members within each FRNA genogroup.

Approach

- Designed genogroup-specific primer sets based on alignment of full-length genome nucleotide sequences to a minimum of 5 strains/genogroup to a maximum of 10 strains/genogroup.
- Designed primer sets to produce discrete PCR amplicon sizes specific to each genogroup.
- Evaluated primers for hairpin loops, primer-dimers and melting profiles for RT-PCR optimization.
- Optimized the heat-release of purified FRNA viral stocks to serve as a template for RT-PCR.
- Optimized the one-step format and cycling temperatures for RT-PCR.
- Validated the one-step RT-PCR with 25 FRNA phages and tested for cross-reactivity.

Materials and methods

Coliphage Isolates and Propagation

FRNA phages were collected from water, sewage and various animals representative of diverse geographical locations ([Table 7.1](#)). Vinjé et al., (2004) designed two degenerate primer sets, MJV82 and JV81 for *Leviviruses* and MJV82 and JV41 for *Allolevivirus*, specific to the replicase region. The combination of these primer sets was termed “generic PCR” as the primers were genus specific but not genogroup specific. Preliminary subgrouping of phages was previously determined by generic RT-PCR and reverse line-blot hybridization ([Vinjé et al., 2004](#)). Environmental and prototype phages were provided from The University of North Carolina at Chapel Hill collection. Strain R17 was purchased from Felix D’Herelle Reference Centre for Bacterial Viruses, Université Laval, Quebec, Canada.

Table 7.1 Sources of *Leviviridae* strains.

Origin of <i>Leviviridae</i> Strains									
Group I					Group III				
Strain	Group	Source	Collected	Site collected					
DL1	I	water	Jan 2004	Tijuana River, CA	TW 18	III	sewage	1970	Changhua, Taiwan ^f
DL2	I	water	Feb 2004	Delaware Bay, DE	HL4-9	III	hog lagoon	May 2000	Duplin County, NC
DL13	I	oyster	Oct 2004	Whiskey Creek (Masonboro Is), NC	BR12	III	water	July 2005	New Market Creek Charleston, SC
DL16	I	water	Nov 2004	Great Bay (Nannie Is), NH					
J20	I	chicken litter	Sept 2000	South Carolina	VK	III	sewage	Oct 1963	Tokyo, Japan ^e
ST4	I	UNK	UNK	UNK	BZ1	III	sewage/feces	Oct-Nov 1971	Recife, Brazil ^g
R17	I	sewage	1962	Philadelphia, PA ^a	QB	III	human feces	June 1961	Kyoto, Japan ^e
MS2	I	sewage	Sept 1959	Berkeley, CA ^b	M11	III	UNK	UNK	Netherlands ^h
M12	I	UNK	UNK	Germany ^a	MX1	III	raw wastewater	1973	Campeche, Mexico ^{i,j}
fr	I	dung-hill	1963	Heidelberg, Germany ^d					
Group II					Group IV				
T72	II	bird	June 2002	Talbert Marsh sandflats, Huntington Beach, CA	HB-P22	IV	bird	April 2002	Talbert Marsh sandflats, Huntington Beach, CA
GA	II	sewage	Oct 1964	Ookayama, Japan ^e	HB-P24	IV	bird	April 2002	Talbert Marsh sandflats, Huntington Beach, CA
KU1	II	sewage	1973	Kuwait ⁱ	BR1	IV	water	Feb 2005	Guerin Creek (Charleston), SC
DL10	II	mussel	April 2005	Tijuana River, CA	BR8	IV	water	June 2005	Bull Creek (Charleston), SC
DL20	II	clam	May 2005	Naragansett Bay, RI	NL95	IV	calves	UNK	Netherlands ^h
					SP	IV	Siamang gibbon	1968	Tokyo, Japan ⁱ
					F1	IV	infants	1969	Hachioji, Japan ^h
					UNK = unknown				

^a Paranchych and Graham, 1962, ^b Dr. Alvin J. Clark, personal communication, ^c Zinder, 1965, ^d Marvin and Hoffman-Berling, 1963, ^e Watanabe et al., 1967, ^f Miyake et al., 1971, ^g Miyake et al., 1973, ^h Beekwilder et al., 1996, ⁱ Hirashima et al., 1983, ^j Sakurai et al., 1968, ^k Miyake et al., 1969, ^m Furuse et al., 1975.

UNK - unknown

Single-plaque coliphages were further purified and enriched using *Escherichia coli* HS(pFamp)R as host (Vinjé et al., 2004). Overnight enrichments were centrifuged (3,220 X g for 10 min) to pellet host cells and debris. A 1:1 chloroform to supernatant mixture was vortexed and centrifuged again. Approx 1-2 ml aliquots of the purified supernatant were frozen at -75°C .

Coliphage titers were determined using a single agar layer procedure (SAL) (US EPA Method 1602, 2001). The procedure was as follows. A 1 ml of overnight *E. coli* Famp host culture was transferred into 50 ml trypticase soy broth with streptomycin and ampicillin (TSB/strep/amp) and grown 4 hr to log phase. A 150 ml volume of trypticase soy agar, TSA (4.5 g TSB and 1.2 g agar), was autoclaved and placed into a water bath ($47\text{-}55^{\circ}\text{C}$). A 300 ul aliquot of 500X strep/amp was added into the cooled 150 ml TSA. Serial 10-fold dilutions of the purified virus were prepared. For 10-fold stock dilution, 100 ul of stock was transferred into 900 ul phosphate buffered saline (PBS), mixed and this process repeated to a serial dilution 10^{-15} . To sterile 15 ml plastic tubes, labeled -1 to -15, 1 ml of the 4 hr *E. coli*, 100 ul of the appropriate virus dilution and 10 ml TSA were added. After quick manual mixing the tube contents were poured into labeled 20mm petri plates which were then cooled, allowed to dry, inverted and incubated overnight at 37°C . The coliphage titer was determined by counting the plates having a minimum of 5 to a maximum of 20 plaques per plate. Titer concentration was reported as PFU/ml.

Coliphage RNA Isolation

Coliphage RNA was extracted from the purified viral supernatant using a QIAamp viral RNA mini kit (Qiagen, Valencia, CA) as follows. A 200 ul volume of purified virus

stock was added into the 25 ul protease tube. Into the same tube, 200 ul of carrier RNA/Buffer AL mixture was added and the combined solutions were vortex mixed, incubated 56 ° C for 15 minute and clarified by a 1 minute centrifugation. To the same tube, 250 ul ethanol (EtOH) was added, vortex mixed and incubated 5 min at room temperature (RT). The lysate was applied into the QIAamp spin column and centrifuged 3-4 min at 6000 xg (8000 rpm) and the contents of the collection tube was discarded. To the column, 500 ul of Buffer AW2 was added, centrifuged 1 min and the contents of the collection tube was discarded. To the column, 500 ul EtOH was added, centrifuged 1 min and collection tube was discarded. The column was transferred to a clean collection tube and centrifuged for 1 minute. The column was again transferred to a new, nuclease-free tube and 50 ul RNase free water was added onto column and incubated 5 min at RT. The final centrifugation, (14,000 rpm) for 1 min and the recovered purified RNA was frozen at -20 °C.

Heat Release of Viral RNA

A direct heat-release procedure was applied to aliquots of undiluted viral supernatant. Ten ul of viral supernatant was heated in thin-walled 250 ul size PCR tubes for 5 min at 98 °C and chilled on ice for 2 min ([Schwab et al., 1997](#); [Vinjé et al., 2004](#)). Aliquots of 5 ul were immediately placed into the RT-PCR mixture for amplification.

Reverse-Transcription PCR

A one-step single tube format, Qiagen One-Step RT-PCR kit (Qiagen, Valencia, CA), was used. The 50 ul reaction volume contained 24 ul RNase free water, 10 ul 5X Qiagen reaction buffer, 2 ul 10mM dNTP, 1 ul 10 uM forward primer, 1 ul 10 uM reverse primer and 2 ul Qiagen RT-PCR enzyme. To each 50 ul reaction volume, 5 ul of heat-released viral RNA

or 5 ul of purified RNA were used for reverse transcription. Thermal cycle (GeneAmp PCR System 9700, PE Applied Biosystems, Foster City, CA) conditions were as follows: 50 °C for 30 min, 95 °C for 15 min followed by 40 cycles of 94 °C for 1 min, 55 °C for 1 min, 72 °C for 1 min with a final extension of 72 °C for 10 min. Amplicons were separated by electrophoresis in 1.5% agarose gel in 0.5X Tris-acetate-EDTA (TAE), stained with 20 ug/ml ethidium bromide and visualized under UV light (UVP, Upland, CA).

A genus-specific RT-PCR assay with primers MJV82 forward and *Levivirus* JV41 reverse or *Allolevivirus* JV81 reverse were utilized as a positive control (Vinjé et al., 2004). For each reaction, a no-template RT-PCR negative control was prepared.

PCR master mixes were prepared in a PCR hood (AirClean 600, AirClean Systems, Raleigh, NC) separate from template additions. To avoid contamination, PCR master mixes, amplification, electrophoresis and template and/or viral preparations were conducted in separate assigned rooms based on designated use.

Primer Design

Sequences (Table 7.2) were aligned using BioEdit v7.0.1 Clustal W application (Hall, 1999). Genogroup-specific primers (Table 7.3) targeting each of the four genogroups were designed to produce discrete amplicon sizes (Fig 7.1) for rapid genogroup-positive visualization. Details of the aligned sequences from which the primers were derived are shown in Fig 7.2. Primer set FRNA I was designed by alignment of isolates DL1, DL2, DL13, DL16, ST4, R17, J20 and GenBank strains MS2, M12 and fr. Primer set FRNA II was designed by alignment of phages T72, DL10, DL20 and GenBank strains GA and KU1.

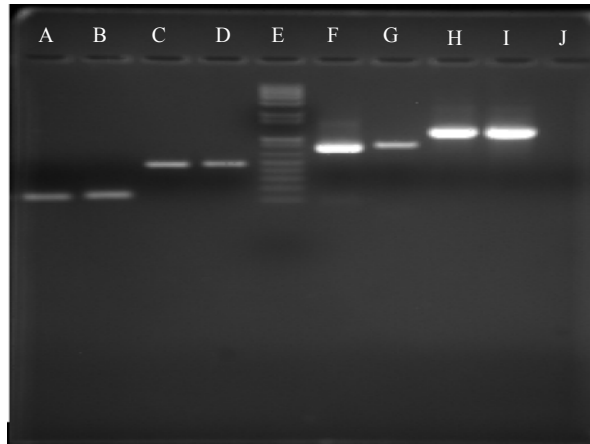
Primer set FRNA III was designed by alignment of TW18, HL4-9, BR12, BZ1, VK and GenBank strains QB, M11 and MX1. Primer set FRNA IV was designed by alignment of HB-P22, HB-P24, BR1, BR8 and GenBank strains SP, NL95 and FI. Primers were evaluated for primer-dimers and hairpin loops by NetPrimer freeware (Premier Biosoft International, Palto Alto, CA) www.premierbiosoft.com/netprimer/netprlaunch/netprlaunch.html).

Table 7.2 *Leviviridae* accession numbers. Accession numbers of *Leviviridae* male-specific ssRNA coliphages (FRNA) available and/or submitted to the GenBank/EMBL/DDBJ database.

<u>Strain</u>	<u>Genogroup</u>	<u>Accession number</u>
MS2	I	NC_001417.1
fr	I	NC_0011333.1
M12	I	AF195778
DL1	I	EU341815
DL16	I	EU341816
ST4	I	EU341817
R17	I	EU341818
J20	I	EU341819
GA	II	NC_001426.1
KU1	II	NC_002250.1
T72	II	EU372691
DL10	II	EU372692
DL20	II	EU372693
Qbeta	III	AY099114.1
M11	III	NC_004304.1
MX1	III	NX_001890.1
TW18	III	EU372694
HL4-9	III	EU372695
BZ1	III	EU372697
VK	III	EU372698
SP	IV	X07489.1
NL95	IV	AF059243.1
FI	IV	EF068134.1
HB-P22	IV	EU403427
HB-P24	IV	EU403428
BR1	IV	EU403429
BR8	IV	EU403430

Figure 7.1 Gel electrophoresis of FRNA phages using one-step RT-PCR.

Gel electrophoresis of heat-released FRNA coliphages following RT-PCR with genogroup-specific primers. (A, B) Genogroup I, 142 bp (C, D) Genogroup II, 471 bp (E) 1 kb Plus Track-It Ladder (F, G) Genogroup III, 795 bp (H, I) Genogroup IV, 1159 bp and (J) negative control.



Primer design based on alignment of genome sequences and nucleotide positions from each respective *Leviviridae* male-specific FRNA genogroup. Alignment for each genogroup is depicted in discontinuous blocks to illustrate the nucleotide position of the primers. The numbers along the top are the nucleotide positions. Primers are underlined. Genome sequences read 5' - 3' direction. Dots indicate identity with the consensus sequence. Degenerate bases are noted in the standard IUB codes. (A) Genogroup I. (B) Genogroup II. (C) Genogroup III. (D) Genogroup IV.

(B) Group II

Group II forward primer

	5'	3'
		90 100 110 120	
Consensus		<u>ATGCCGTTAG</u> <u>GTTTAGRTGA</u> <u>CGGTATRTTC</u>	
T72	G....A.-.	
DL10	A....A.-.	
DL20	A....A....	
GA	G....G.-.	
KU1	A....A.-.	

Group II reverse primer

	5'	3'
		540 550 560 570	
Consensus		TCAAT <u>TATTGG</u> <u>GGTTGCDATT</u> <u>GCD</u> GARGCYA	
T72	G....A..A..T.	
DL10	A....T..A..T.	
DL20	G....T..G..T.	
GA	A....T..A..C.	
KU1	T....G..A..C.	

(C) Group III

Group III forward primer

	5'	3'
		660 670 680 690 700	
Consensus		AARCCDRCTA <u>CTGCTGGTAA</u> <u>TCTCTGGCT</u> Y GARTTYMGKT AYGGVYTHRY	
TW18		..G..GA.....T..A..TC.T..T..CC.TAT	
HL4-9		..A..AA.....T..A..CC.T..T..TC.TAT	
BR12		..A..GA.....T..A..TC.T..T..CC.TAT	
BZ1		..A..GA.....T..A..TC.T..T..CC.TAT	
VK		..A..GA.....T..A..TC.T..T..CC.TAT	
QB		..A..GG.....T..A..TC.T..T..CC.TAT	
M11		..G..TG.....C..A..TC.T..T..CC.CGT	
MX1		..G..TG.....C..G..TA.G..C..TT.AAC	

Group III reverse primer

	5'	3'
		1430 1440 1450 1460	
Consensus		<u>GGTAAATCCC</u> <u>ACYAACGGYG</u> <u>TTG</u> CCCKCGCT	
TW18	T.....T.....G....	
HL4-9	T.....T.....G....	
BR12	T.....T.....T....	
BZ1	C.....T.....T....	
VK	T.....T.....T....	
QB	T.....C.....T....	
M11	T.....T.....G....	
MX1	C.....T.....G....	

(D) Group IV

Group IV forward primer

5'	3'
	2960 2970 2980 2990 3000	
Consensus	TCGTGGAAGC ATG CCTGTCC GCAGGATGT ACCA AACGYG CRYTVAAARTA	
BR1T.....C..AT.C..G..	
BR8T.....C..AC.C..G..	
HB-P22T.....C..AT.C..A..	
HB-P24A.....T..AC.G..A..	
SPT.....C..AT.C..G..	
NL95A.....C..AC.A..G..	
FIT.....C..GC.G..G..	

Group IV reverse primer

5'	3'
	4110 4120 4130 4140 4150	
Consensus	TDG CGTGGAT TCAGGACAGT GCCTT CATCC GGCCCCCKTA TTYNMTTAMG	
BR1	.G.....G..TCA..C.	
BR8	.A.....G..TCA..C.	
HB-P22	.G.....G..TTC..C.	
HB-P24	.G.....T..TTA..A.	
SP	.G.....G..TAA..C.	
NL95	.T.....T..TTA..A.	
FI	.G.....G..CG---C.	

Results

Detection of the Four Genogroups of FRNA Coliphages

Within the aligned genomic sequences of each FRNA coliphage genogroup, areas of genetic variability were observed ([Appendix A](#)). Robust, genogroup specific RT-PCR primers were designed to conserved sequences from a variety of strains (10 strains from group I, 5 strains from group II, 8 strains from group III and 7 strains from group IV) ([Table 7.1](#), [Fig 7.2](#)).

The four genogroup-specific primer sets generated discrete PCR amplicon sizes ([Fig 7.1](#)). The discrete amplicon sizes allowed rapid visualization of genogroup-positive FRNA strains. In addition, primer sets lacked primer-dimers or hairpin loop formations thereby allowing optimal PCR amplification. Primer set FRNA I ([Table 7.3](#)) produced 142 base pair (bp) amplicons from isolates DL1, DL2, DL13, DL16, ST4, R17, J20 and strain MS2. Primer set FRNA II produced 471 bp amplicons from isolates T72, DL10, DL20 and strain GA. Primer set III produced 795 bp amplicons from isolates TW18, HL4-9, BR12, BZ1, VK and strain QB. Primer set IV produced 1159 bp amplicons from isolates HB-P22, HB-P24, BR1, BR8 and strains SP and NL95.

Table 7.3 Genogroup-specific primer sets. Genogroup-specific primer sets designed to detect the FRNA coliphage (*Leviviridae*). Degenerate bases are highlighted in bold and written in the standard IUB code. UTR – untranslated region.

<u>Group</u>	<u>Primer</u>	<u>Sequence</u>	<u>Amplicon (bp)</u>	<u>Gene</u>
I	FRNA I F (forward)	5' CAAACCAGCATCCGTAGCC 3'	141	Replicase
I	FRNA I R (reverse)	5' CTTGTTCA GCGAACTTCT TR TA 3'		Replicase
II	FRNA II F (forward)	5' ATGCCGTTAGGTTTAG RT GAC 3'	471	5' UTR
II	FRNA II R (reverse)	5' GCAAT H GCAACCCCAATA 3'		Maturation
III	FRNA III F (forward)	5' CTA CTGCTGGTAATCTCTGGC 3'	795	Maturation
III	FRNA III R (reverse)	5'CAAC R CCGTT RT G GGGATTTA C 3'		Capsid
IV	FRNA IV F (forward)	5' CTGTCCGCAGGATGT W ACCA 3'	1159	Replicase
IV	FRNA IV R (reverse)	5' GGCACTGTCCTGAATCCACG 3'		Replicase

Detection of FRNA Coliphages Using Heat-released Viral RNA

For most coliphage strains, heat-released viral RNA (Schwab et al., 1997; Vinjé et al., 2004) produced the desired amplicon. If the amplicon was not observed, purified RNA was used to supplement the RT-PCR assay. Strains requiring purified RNA were J20, GA, QB and NL95. Use of RNA vs heat-release was not titer dependent as phage QB had a titer $> 10^{10}$ but this strain would not amplify with heat-release whereas strain SP amplified at a 10^3 titer.

Testing for Cross-reactivity of Primer Sets

Although BLAST (www.ncbi.nlm.nih.gov/BLAST) results did not indicate cross-reactivity in GenBank strains, each primer set was tested using a total of 25 environmental and prototype FRNA strains representing each genogroup. Cross-reactivity was not observed between groups I and II (genus *Levivirus*) or between groups III and IV (genus *Allolevivirus*). However, primer set IV produced faint non-specific amplification when *Levivirus* genogroups I and II strains were tested with RT-PCR (data not shown). Preliminary screening of environmental FRNA strains using generic primers as described (Vinjé et al., 2004) will determine the genus. Once the genus is determined by generic RT-PCR, subsequent genogroup-specific primers, I and II for *Levivirus* or III and IV for *Allolevivirus*, will eliminate non-specific amplification. However, the non-specific PCR amplification products were faint and amplicons were not the same molecular size or intensity as generated by each genogroup-specific primer set.

Discussion

In this study are reported the first broadly representative but specific RT-PCR primer

sets for each of the four separate FRNA coliphage genogroups designed from a large (30 strains) genetic sequence database. The robust genogroup-specific primers detected a variety of purified FRNA coliphages from various sources and samples collected around the world. Diverse FRNA coliphage sources were adequately represented, as those used for primer design came from a variety of water bodies, fecal waste sources and their animal hosts including birds, chickens, swine, oyster, mussel, clam, sewage, human feces, calves and lesser apes ([Table 7.1](#)). Each genogroup-specific primer set was designed from a minimum of 5 strains per group in which the complete genome was known. In addition, FRNA genogroup strains were isolated from the United States, Japan, Europe, Brazil, Taiwan and/or Mexico.

Previous investigations using real-time PCR or RT-PCR for detection and genotyping of FRNA coliphages used extracted viral RNA from either purified virus or environmental water samples ([O'Connell et al., 2006](#); [Dryden et al., 2006](#); [Kirs and Smith, 2007](#); [Ogorzaly and Gantzer, 2006](#)). In this study, coliphage RNA was made available for amplification by RT-PCR using a heat-release RNA technique directly applicable to culture enriched coliphage isolates ([Schwab et al., 1997](#); [Vinjé et al., 2004](#)). This heat-release procedure reduces RT-PCR preparation time by omitting initial coliphage isolation followed by RNA chemical purification steps. In field samples where low numbers of coliphages are present, it may be necessary to first enrich for FRNA coliphages ([Love and Sobsey, 2007](#)) followed by heat-release of enrichments for RT-PCR amplification.

Using primers designed from only a limited number of complete or partial nucleotide sequences available in the NCBI GenBank, Dryden et al., ([2006](#)), failed to detect FRNA coliphages in several environmental samples, despite the fact that coliphages were detected at

all sites by the single agar layer method (SAL). The authors acknowledged the possibility that their RT-PCR assay failed to detect unknown FRNA phages. Indeed, this may have been the case as their primer sets were designed to the limited FRNA sequences in GenBank, and, primers were not designed to genogroup III.

In previous studies, the sensitivity of lower detection limits appears to be generally consistent. Using purified RNA, RTQPCR for all four genogroups of FRNA phages detected 0.1 plaque-forming units (PFU) per 50 ul reaction in seawater and 0.5 PFU/50 ul reaction in stool samples ([Kirs and Smith, 2007](#)). These findings were comparable to a sensitivity of 0.1 PFU in laboratory-prepared MS2 in an RT-PCR assay using a single primer set that detects, but does not differentiate, the *Leviviridae* family ([Rose et al., 1997](#)). Lower detection limits for various FRNA phage strains with RTQPCR ranged from 1-10 PFU/ml for MS2, 0.01-0.1 PFU/ml for GA and SP and 0.1-1 PFU/ml for phage QB ([Ogorzaly and Gantzer, 2006](#)). The sensitivity of a real-time fluorogenic RT-PCR for FRNA strain MS2 detected 40-0.4 fg of RNA per 20 ul reaction ([O'Connell et al., 2006](#)). Sensitivity of detection based on a lower limit or dilution endpoint to extinction was not measured in this assay. The method was applied to phage enrichments that contained $>10^3$ PFU. Further studies are planned to determine the lower limit of FRNA coliphage detection and genotyping when the method is applied directly to enriched and serially diluted coliphages and coliphages in unenriched environmental samples such as feces, manure, biosolids and wastewater.

The new genogroup-specific primers, RNA preparation and RT-PCR amplification procedures described here should facilitate improved and more reliable genotyping of different FRNA coliphage isolates in environmental samples.

Summary

- Using genogroup-specific primers directed to highly conserved consensus sequences in each genogroup, a one-step RT-PCR was developed that detected all four genogroups when applied to 25 environmental and prototype FRNA strains.
- Primer sets produced PCR amplicon sizes of 142 bp, 471 bp, 795 bp and 1159 bp to genogroups I, II, III and IV, respectively.
- Discrete amplicon sizes allow easy visualization of genogroup-specific amplicons produced in the RT-PCR.
- The RT-PCR successfully amplified 25 FRNA coliphages when applied to direct heat-released viral RNA template, which reduced RNA sample preparation time by omitting the time-consuming and costly chemical RNA purification steps.
- These genogroup-specific primers sets can aid in source-tracking FRNA coliphages.

References

- Beekwilder, J., Nieuwenhuizen, R., Poot, R., van Duin, J., 1996. Secondary structure model for the first three domains of QB RNA. Control of A-protein synthesis. *J. Mol. Biol.* 256(1), 8-19.
- Brion, G.M., Meschke, J.S., Sobsey, M.D., 2002. F-specific RNA coliphages: occurrence, types, and survival in natural waters. *Water Res.* 36, 2419-2425.
- Cole, D., Long, S.C., Sobsey, M.D., 2003. Evaluation of F+ RNA and DNA coliphages as source-specific indicators of fecal contamination in surface waters. *Appl. Env. Micro.* 69, 6507-6514.
- Dryden, S.K., Ramaswami, B., Yuan, Z., Giammar, D.E., Angenent, L.T., 2006. A rapid reverse transcription-PCR assay for F+ RNA coliphages to trace fecal pollution in Table Rock Lake on the Arkansas-Missouri border. *Water Res.* 40, 3719-3724.
- Furuse, K., Ando, A., Watanabe, I. 1975. Isolation and grouping of RNA phages VII. A survey in Peru, Bolivia, Mexico, Kuwait, France, Australia and the United States of America. *J. Keio. Med. Soc.*, 52, 353-361.
- Gerba, C.P., 1987. Phage as indicators of fecal pollution, in: Goyal, S.M., Gerba, C.P., Bitton, G. (Eds), *Phage Ecology*. Wiley Interscience, New York, pp. 197-210.
- Grabow, W.O.K., 2001. Bacteriophages: update on applications as models for viruses in water. *Water S.A.* 27, 251-268.
- Griffin, D.W., Stokes, R., Rose, J.B., Paul, J.H. III., 2000. Bacterial indicator occurrence and the use of an F(+) specific RNA coliphage assay to identify fecal sources in Homosassa Springs, Florida. *Microbial. Ecol.* 39, 56-64.
- Hall, T.A., 1999. BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucl. Acids Symp. Ser.* 41, 95-98.
- Havelaar, A.H., Furuse, K., Hogeboom, W.M., 1986. Bacteriophages and indicator bacteria in human and animal faeces. *J. Appl. Bacteriol.* 60, 255-262.
- Hirashima A., Inokuchi, Y., Harigai, H., Furuse, K., Watanabe, I., 1983. Biochemical characterization of RNA coliphage MX1. *J. Gen. Virol.* 64(4), 939-946.
- Hsu, F.C., Shieh, Y.S., van Duin, J., Beekwilder, M.J., Sobsey, M.D., 1995. Genotyping male-specific RNA coliphages by hybridization with oligonucleotide probes. *Appl. Env. Micro.* 61(11), 3960-3966.
- Kirs, M., Smith, D.C., 2007. Multiplex quantitative real-time reverse transcriptase PCR for F+

- specific RNA coliphages: a method for use in microbial source tracking. *Appl. Env. Micro.* 73(3), 808-814.
- Love, D.C., Sobsey, M.D., 2007. Simple and rapid F+ coliphage culture, latex agglutination, and typing assay to detect and source track fecal contamination. *Appl. Env. Micro.* 73(13), 4110-4118.
- Marvin, D.A., Hoffman-Berling, H., 1963. Physical and chemical properties of two new small bacteriophages. *Nature.* 197(4866), 517-518.
- Miyake, T., Shiba, T., Sakurai, T., Watanabe, I., 1969. Isolation and properties of two new RNA phages SP and FI.. *Japan. J. Micro.* 13(4), 375-382.
- Miyake, T., Furuse, K., Shiba, T., Aoi, T., Sakurai, T., Watanabe, I., 1971. Isolation and grouping of RNA phages in Taiwan. *J. Keio Med. Soc.* 48, 25-34. (Japanese)
- Miyake, T., Furuse, K., Shiba, T., Aoi, T., Sakurai, T., Watanabe, I., 1973. Isolation and grouping of RNA phages. II. A survey in Brazil. *J. Keio Med. Soc.* 50, 353-362. (Japanese)
- O'Connell, K.P., Bucher, J.R., Anderson, P.E., Cao, C.J., Khan, A.S., Gostomaki, M.V., Valdes, J.J., 2006. Real-time fluorogenic reverse transcription-PCR assays for the detection of bacteriophage MS2. *Appl. Env. Micro.* 72(1), 478-483.
- Ogorzaly, L., Gantzer, C., 2006. Development of real-time RT-PCR methods for specific detection of F-specific RNA bacteriophage genogroups: application to urban raw wastewater. *J. Virol. Meth.* 138, 131-139.
- Paranchych, W., Graham, A.F., 1962. Isolation and properties of an RNA-containing bacteriophage. *J. Cell Comp. Physiol.* 60, 199-208.
- Rose, J.B., Zhou, X., Griffin, D.W., Paul, J.H., 1997. Comparison of PCR and plaque assay for detection and enumeration of coliphage in polluted marine waters. *Appl. Env. Micro.*, 63(11), 4564-4566.
- Sakurai, T., Miyake, T., Shiba, T., I. Watanabe, 1968. Isolation of a possible fourth group of RNA phage. *Japan. J. Micro.* 12, 544-546.
- Schaper, M., Jofre, J., Uys, M., Grabow, W.O., 2002. Distribution of genotypes of F-specific RNA bacteriophages in human and non-human sources of faecal pollution in South African and Spain. *J. Appl. Micro.* 92, 657-667.
- Schwab, K.J., Estes, M.K., Neill, F.H., Atmar, R.L., 1997. Use of heat release and an internal RNA standard control in reverse transcription-PCR detection of Norwalk virus from

- stool samples. J. Clin. Micro. 35(2), 511-514.
- Sobsey, M.D., Love, D., Lovelace, G., Stewart, J.R., Robinson, B., 2005. Methods to detect and genotype coliphages in water and shellfish. Biennial Meeting of the Interstate Shellfish Sanitation Conference.
<http://www.unc.edu/sobseylab/ISSCcoliphagedemo.pdf>
- Stewart, J.R., Daugomah, J.W., Chestnut, D.E., Graves, D.A., Sobsey, M.D., Scott, G.I., 2006. FRNA coliphage typing for microbial source tracking in surface waters. J. Appl. Micro. 101, 1015-1026.
- Stewart, J.R., Vinjé, J., Oudejans, S.J.G., Scott, G.I., 2006. Sequence variation among group III F-specific RNA coliphages from water samples and swine lagoons. Appl. Env. Micro. 72, 1226-1230.
- Sundram, A., Jumanial, N., Ehlers, M.M., 2006. Genotyping of F-RNA coliphages isolated from wastewater and river samples. Water S.A. 32(1), 65-70.
- US EPA., 2001. Method 1602: Male-specific (F+) and somatic coliphage in water by single agar layer (SAL) procedure. Office of Water, EPA 821-R-01-029. Washington, D.C., 20460. www.epa.gov/nerlcwww/1602ap01.pdf
- van Duin, J., 1998. The single-stranded RNA bacteriophages. in: Fraenkel Conrat, H., Wagner, R.R. (Eds). The Bacteriophages. Series *The Viruses*. Plenum Press, NY. pp. 117-167.
- Vinje, J., Oudejans, S.J.G., Stewart, J.R., Sobsey, M.D., Long, S.C., 2004. Molecular detection and genotyping of male-specific coliphages by reverse-transcription-PCR and reverse line blot hybridization. Appl. Env. Micro. 70(10), 5996-6004.
- Watanabe, I., Miyake, T., Sakurai, T., Shiba, T., Ohno.T., 1967. Isolation and grouping of RNA phages. Proc. Jap. Acad. 43(3), 204-209.
- Zinder, N.D., 1965. RNA phages. Ann. Rev. Microbiol. 19, 455-473.

VIII. Overall Discussion

Male-specific RNA coliphages, among the smallest autonomous viruses known, are positive sense, single-strand RNA (ssRNA) phages possessing a genome, 3.8 to 4.2 kb, enclosed within a non-enveloped 26 nm icosahedral-shaped capsid ([Buchen-Osmond, 2003](#)). These coliphages belonging to the family *Leviviridae* were initially grouped primarily according to their serological properties. The *Leviviridae* family contain two genera, *Levivirus* and *Allolevivirus* which are further subdivided into four major serogroups, I, II, III, IV and branched subgroups (a,b,c,d) according to serological cross-reactivity ([Sundram et al., 2006](#)). *Levivirus* are subdivided into genogroups I and II and *Allolevivirus* are subdivided into genogroups III and IV. It became apparent as early as the 1970s that the four genogroups of FRNA coliphages had somewhat different fecal source and geographic distributions. In the 1990s the development of genotyping methods based on synthetic oligonucleotide probes ([Hsu et al., 1995](#); [Beekwilder et al., 1996](#)) made it possible and convenient to genotype FRNA coliphages and better understand their ecology, their value as fecal and viral indicators, their ability to distinguish human from animal fecal wastes and the impacts of these different waste sources on ambient waters. By employing these methods it became apparent that more information regarding source of fecal pollution could be obtained by comparing full-length genomic sequences from FRNA coliphages collected from various animals and water bodies.

Male-specific coliphages have been suggested as a viral indicator for: (1) fecal contamination ([Osawa, 1981](#); [Furuse, 1983](#)), (2) enteric bacterial contamination ([Gerba, 1987](#)), (3) enteric viral contamination ([Grabow, 2001](#); [Leclerc et al, 2000](#)) and (4) risks of gastro-intestinal illness from recreational water exposures ([Colford et al., 2007](#)). FRNA coliphages are almost indistinguishable from most human enteric viruses ([Grabow, 2001](#)), occur in higher numbers in sewage and wastewater effluents than viral enteric pathogens ([Grabow, 2001](#)), their presence implies the presence of pathogenic viruses ([Grabow, 2001](#)), and, in a majority of cases, they display fecal-source specificity ([Vinjé et al, 2004](#); [Cole et al, 2003](#); [Furuse, 1987](#); [Schaper et al, 2002](#); [Scott et al, 2002](#); [Stewart, 2002](#); [Long et al, 2005](#)).

The focus of this study was to develop and validate a rapid, genogroup-specific molecular assay for the detection of FRNA coliphages as a potential viral indicator of fecal pollution. Before the molecular assay could be developed, a genetic sequence database was generated representing environmental and prototype FRNA coliphage strains from all four genogroups.

A ssRNA viral assay would need a large (at least 5 strains/genogroup) genetic sequence database. To develop a genetic database, 19 FRNA strains were sequenced and compared to the 11 FRNA full-length sequences available in the National Center for Biotechnology Information (NCBI) genetic database (GenBank) for a total of 30 FRNA strains. FRNA phages were collected from water, sewage, and various animals representative of diverse geographical locations ([Table 7.1](#)). The field-collected FRNA strains and prototype strains were represented by phages isolated from the United States, Japan, Europe, Brazil, Taiwan and/or Mexico ([Table 7.1](#)). The majority of groups II (80%) and III (75%)

strains were collected at municipal sewage sources or water bodies with the exception of one group II strain (collected from bird droppings), one group III strain (collected from swine lagoon) and one group III strain was from an unknown source. Two out of 10 group I strains (20%) were collected from sewage, 4 out of 10 group I strains (40%) were collected from ambient waters and/or sentinel organisms (oyster, clam, mussel), one strains' source was unknown and one strain was collected from a dung-hill. Four out of seven group IV strains (57%) were collected from animal sources (bird, gibbon, calves), two strains (29%) were collected from ambient water sources and one strain was obtained from an infant ([Table 7.1](#)).

Phages were sequenced by primer walking. A polyadenylated (Poly-A) tail was added to the 3' end of purified viral RNA, the poly-A RNA was reverse transcribed with a Poly-T reverse primer; the resulting cDNA was used as a PCR template. A gene-specific forward and a poly-T primer used in the PCR mixture produced an approximate one kilobase (kb) amplicon. PCR products were gel-purified (GenScript, Piscataway, NJ), cloned (Invitrogen, Carlsbad, CA) and sequenced (Sequetech, Mountain View, CA). This process was repeated until the majority (all sequences except 100-200 bp at the 5' end) of the genome was obtained. A rapid amplification of cDNA ends (RACE) Smart RACE kit (Clontech, Mountain View, CA) was used to amplify the 5' portion of the genome.

When full-length genome nucleotide sequences were aligned with the published GenBank strains within their respective genogroup, very similar or identical gene mapping, or Open Reading Frames (ORF), were observed for all four *Leviviridae* genes indicating the sequence data generated in this study was valid. Sequence similarity among genogroup I strains ranged from 75.27 - 96.67 % with strain fr forming a separate subgroup ([Table 5.2](#), [Fig](#)

5.2). Among group II strains, nucleotide sequence similarity ranged from 83.30 to 93.84% with strains DL10, DL20 and GA having the highest sequence identities (93.43-93.67%) whereas strains T72 and KU1 formed a separate subcluster (Table 5.2, Fig 5.2). Among *Allolevivirus* group III, two different subclusters were formed. The first subcluster was composed of strains VK, HL4-9, BR12, BZ1, TW18 and GenBank strain Q β having a nucleotide sequence similarity ranging from 91.87-95.69%. The second subcluster formed with GenBank group III strains MX1 and M11 having an 87% nucleotide similarity to each other. The nucleotide similarity of strains between the two group III subclusters ranged from 69.77-71.33% (Table 5.2, Fig 5.2). Group IV *Allolevivirus* shared sequence identities ranging from 74.90-95.03 % with the closest identities being 95.03% between strains BR8 and BR1.

Amino acid composition was similar among genogroups, further validating the nucleotide sequences. With the exception of strain fr, amino acid number was consistent in each of the four protein types in group I phages (Table 5.4, Appendix B). The capsid protein of all strains in group I was 130 amino acids in length. *Levivivirus* groups I and II capsid proteins shared a conserved region consisting of a 10 amino acid, FVLVDNNGGTG, consensus sequence. Groups I and II maturation protein shared a consensus region RWLELQ at amino acid (aa) positions number 198-203. The length of the maturation protein of groups III and IV varied from 420 aa to 450 aa (Table 5.4) and possessed a mutual conserved aa region LWLEFRYGL (Appendix B). The length of the capsid protein was 133 and 132 aa for groups III and IV, respectively, and conserved stretches of amino acids occurred in both groups.

An algorithmic approach was selected to construct phylogenetic trees from the nucleotide sequence and amino acid data (Fig 5.2, Fig 5.4). Nucleotide sequences in the

phylogenetic tree of *Levivirus* group I strains produced two branches, with 9 strains clustered as MS2-like and strain fr an individual branch (Fig 5.2). Within group II nucleotide sequences, strains KU1 and T72 formed one branch and strains DL10, DL20 and GA formed a second branch. *Allolevivirus* group III nucleotide sequences clustered into a MX1, M11 branch and a second branch with Q β -like strains BR12, VK, BZ1, HL4-9, TW18 and prototype Q β . Nucleotide sequence analysis formed three branches in group IV strains as follows: 1) HB-P24, HB-P22 and prototype NL95, 2) BR1, BR8 and prototype SP and 3) prototype FI (Fig 5.2). Individual proteins were clustered into phylogenetic trees. In some cases, phylogenetic protein trees formed more subclusters or branches than the nucleotide trees (Fig 5.4). Genome organization, amino acid conservation and identical or very similar nucleotide start and stop positions supported the *Leviviridae* genogroup designation. In addition, eight nucleotides on the 3' termini clearly distinguish the *Allolevivirus*, 5' TCCTCCCA 3', from the *Levivirus*, 5' ACCACCCA 3'.

In addition, two new undescribed *Levivirus* strains which did not hybridize to previously designed geno-specific hybridization probes (Vinjé et al., 2004) were sequenced. The two unique FRNA strains were collected from North Carolina and Rhode Island. Full-length genomic sequences from the novel strains were compared to nucleotide and/or amino acid sequences from 10 group I strains (MS2, DL1, DL2, DL13, DL16, ST4, R17, J20, M12, fr) and 5 group II strains (T72, DL10, DL20, GA, KUI). Based on full-length genome sequences and phylogenetic analyses, these novel strains were placed into a “JS” subcluster of genogroup I. Sequence similarities of the maturation, capsid and lysis proteins of the JS strains were very similar to those of the MS2-like group I strains, sharing 99-100%, 98-100%

and 95-100% sequence similarities, respectively (Table 6.3, Fig 6.1). However, the replicase protein sequences of the JS strains were quite dissimilar to the replicase protein sequences of the MS2-like genogroup I strains, displaying a similarity range of 84-85% and a frame shift resulting from a two nucleotide insertion (Fig 6.5). The results of this study provide molecular genetic evidence indicative of recombination in two JS strains of FRNA coliphages. The JS strains provided insight to phage ecology and recombination events in natural FRNA strains.

In this study, analyses of complete genomic sequences from 30 FRNA phages plus two novel strains support the known classification scheme. That is, the *Leviviridae* consist of two genera and four distinct genogroups. From this analysis an observation was made that to better define the sub-groupings, it may be more reasonable to assign an association to a specific strain name, i.e., Q β -like instead of genogroup III, subgroup a; MX1-like instead of genogroup III, subgroup b; and in group IV SP-like instead of genogroup IV, subgroup a; and FI-like instead of genogroup IV, subgroup b. Thus, a recommendation based on these findings would be to dismiss the alphabetical sub-grouping nomenclature.

Rose et al (1997) designed a one-step reverse transcription polymerase chain reaction (RT-PCR) using a single primer set that detects, but does not differentiate, FRNA coliphages. In this dissertation, a RT-PCR was designed to distinguish the four FRNA coliphage groups (I, II, III, IV) and ultimately, to distinguish human vs animal fecal sources. Primer sets were designed based on the complete genomic sequences of 30 FRNA strains. Genogroup specific RT-PCR primers were designed to conserved sequences from a variety of strains (10 strains from group I, 5 strains from group II, 8 strains from group III and 7 strains from group IV) (Table 7.1, Fig 7.2). Unique amplicon sizes were generated to allow rapid visualization of

each genogroup ([Fig 7.1](#)). The traditional one-step RT-PCR was developed, optimized for use with heat-released viral nucleic acid and tested for cross-reactivity. Rigorous validation to ensure lack of cross-reactivity was performed whereby each primer set was tested against 25 environmental and prototype strains. This assay was developed, in part, to allow laboratories lacking real-time equipment the ability to genotype FRNA isolates.

A limitation of molecular detection is that nucleic acid presence or persistence of the phage is detected and is not necessarily representative of the presence of infectious viruses. It has been suggested that if the virion capsid is disrupted, the RNA should degrade rapidly under environmental conditions. This position has not been supported by some lab and field studies which addressed long-term persistence of viral nucleic acids in environmental waters ([Kirs and Smith, 2007](#)). Another proposed approach is the discrimination between an intact but non-infectious FRNA phage based on degradation of accessible viral RNA by RNase to eliminate detection of free RNA from damaged (leaky) capsids.

The molecular FRNA phage assay developed in this study may be applicable to an accelerated turnaround time as the assay omits the RNA purification procedure by use of direct heat-release. The traditional primers developed not only allow genogroup identification but provide a comprehensive assessment as to the sanitary quality of the water. If any FRNA phages are detected, then fecal contamination has occurred. This approach utilized a non-cultivation library-independent method for differentiating between human and animal fecal contamination.

In a global phage genotyping assessment, genotypes were reported to be differentially distributed. For example, the FRNA phage from sewage samples in Brazil and West

Germany were from group I exclusively. However, it was unclear as to whether or not sewage treatment plants received slaughterhouse waste ([Furuse, 1987](#)). If, however, both slaughterhouse and domestic sewage were combined, and, if only groups II and III are specific to humans, then presumably at least genogroups I, II and III would have been detected in their study. Furuse argued that group I phages observed in raw sewage from treatment plants were most likely introduced from animal sources, and group II and group III phage were from human sources. This begs the question as to why their study only detected group I in sewage treatment facilities from Germany and Brazil. However, several explanations are possible for this result. Group I could have out-competed groups II and III or they could have slower inactivation rates. This trend of persistent group I FRNA coliphages should be apparent in other sampling stations if these explanations are correct. However, only limited genotyping from other studies are available for making such observations ([Osawa et al., 1981](#); [Miyake et al., 1971](#)). Certain FRNA genogroups may predominate in various human and/or animal populations and their occurrence may also be influenced by climate, diet, intestinal fauna, etc. Further study is needed in different geographical locations to better understand the ecology or natural history of the different FRNA coliphage genogroups; methods developed in the current study should contribute to this effort. Despite the lack of an absolute association between an FRNA genogroup and a unique source, these coliphages likely signal the presence of fecal contamination from either animal and/or human origin. Thus, an FRNA positive sample(s) warrants further investigation, intervention and under some circumstances a public notification alert.

The following three paragraphs will discuss governmental and organizational standards for sanitary quality of recreational water as they pertain to applications of this dissertation. Many of our nation's ambient water resources are impaired and fail to achieve US EPA implemented water-quality standards. In a report dated April 29, 2008, the number of national impaired waters was 39,918 and the leading impairment was pathogens, totaling 9191 of reported impaired waters, or 14.16% (iaspub.epa.gov/waters/national_rept.control). A water body is defined as impaired when the water body fails to maintain water quality standards even after applying effluent limits for point sources (Clean Water Act Section 303(d); www.epa.gov/waterscience/standards/303.htm). Water quality standards are based on water quality conditions and pollution sources, i.e., pathogens, nutrients, sediments, metals, habitat alteration and specific chemicals (www.epa.gov/waterscience/standards/about/).

The EPA Beaches Environmental Assessment and Coastal Health Program (BEACH) Act of 2000 was decreed to improve public health and recreational water quality. Fecal contaminated recreational waters may pose a potential health risk as bathers could contract a waterborne disease spread by fecal-oral route (www.epa.gov/waterscience/beaches/report/chapter02.pdf). Beach-goers who swim or bath in fecal-contaminated waters are at a greater risk than non-swimmers for contracting gastroenteritis. To protect public health, the use of fecal indicators as water quality standards were implemented (EPA, 1983; EPA, 1984). The BEACH Act (amended Section 303 of the Clean Water Act) requires states, tribes and territories to integrate EPA's water quality criteria, *E. coli* and/or enterococci, as their water quality standard (EPA, 2003). The focus of the BEACHs program is to strengthen beach testing and standards, provide faster testing

methods, predict pollution, to better define the criteria as to which fecal indicators and water quality standards are based, to invest in health and methods research and to inform the public (EPA, 2003). The BEACH Act requires states to adopt water quality standards that are “as protective of human health” as the federal criteria. The federal water quality criteria “Ambient Water Quality Criteria for Bacteria in Recreational Waters, 1986” was developed from US EPA epidemiology studies conducted over a period of years (1972 - 1978) at beaches located in New York, Louisiana, Massachusetts and Egypt (EPA, 1983; EPA, 1984). Epidemiological data supported the use of *E. coli* and enterococci as primary fecal indicators associated with statistically significant increased gastrointestinal illness rates to swimmers/bathers (EPA, 1983; EPA, 1984). A review of epidemiological studies and fecal indicators (Pruss, 1998) concluded the following: 1) an exposure-response relationship exists in recreational waters between bacterial indicator counts and gastrointestinal symptoms in exposed beach-goers (swimmers) and 2) there was no demonstrated relationship between bacterial indicator counts and non-gastrointestinal symptoms, i.e., rash, eyes, nose, ears.

The primary aim of the World Health Organization’s (WHO) “Guidelines for Safe Recreational Water Environments” (2003) is to protect the public health by addressing such issues as exposure to sewage-contaminated waters, exposure to freeliving pathogenic organisms such as *Vibrio*, *Aeromonas* sp. and *N. fowleri*, exposure to contaminated beach sand and other potential exposures encountered in recreational water use. Unlike the EPA epidemiology studies conducted in the United States, the WHO based their selection of fecal water quality indicators and health-risk outcomes on a series of randomized control trials conducted in the United Kingdom (REF). The selected bacterial fecal indicator for marine

waters was enterococci whereby a dose-response relationship between enterococci density and health outcome, i.e., gastrointestinal illness and acute febrile respiratory illness (AFRI), was demonstrated. However, the WHO document did not recommend or find a statistical salient

fecal indicator for freshwaters. Enterococci are also the EPA fecal indicator for monitoring marine recreational waters and *E. coli* is an indicator in freshwaters.

Obstacles to current fecal indicator methods include bacterial culture-based methods, or if a molecular assay is used, the protocols usually involve bacterial or viral RNA/DNA concentration and purification steps thereby increasing the time frame between sample collection, data analysis, public health intervention and protection. Most culture-based detection methods currently require at least a 24-48 hr time lag from sample collection to outcome and therefore, provide information that fecal contamination occurred within the past 24-48 hr. Public health intervention to protect bathers prior to exposure would necessitate sample analysis and data confirmation to occur within hours of sample collection, not days. This lag time between sample collection, completion of analysis and public notification causes a window of potential risk to bathers from exposure to pathogens. Clearly, real-time or short-term detection, with limited (1-4 hr) turnaround time from water sample collection to results to public notification are imperative for timely protection of bathers.

Although both the EPA and WHO have developed rigorous recreational water quality guidelines, one limitation is that the bacterial indicators have little or no correlation to the presence of pathogenic viruses ([Griffin et al., 2003](#)). Bacterial indicators may be an erroneous predictor of viral presence as their survival rates do not match those of viruses. Even when

current bacterial standards are met in recreational waters, risks to human health may be posed by viruses. For example, most illnesses contracted by swimmers appear to be of viral etiology (Griffin et al., 2003). Erroneous bacterial counts have been documented as these fecal indicator bacteria periodically occurred naturally in temperate climates (Hardina and Fujioka, 1991; Roll and Fujioka, 1997; Byappanahalli and Fujioka, 1998; Fujioka and Byappanahalli, 2000; Solo-Gabriele HM et al., 2000; Genthner et al., 2005). Elevated bacterial indicator counts exceeding EPA water-quality criteria were influenced by soil run-off and not a result of sewage input (Byappanahalli and Fujioka, 2004).

To date, a viral indicator has not been mandated for regulatory purposes in recreational waters. Additional gaps for determining fecal contamination are that these methods are not real-time nor do they provide information regarding source. To minimize risks to human health, resource managers and human health advisors need an early-warning indicator, an indicator that addresses fecal source and an indicator indicative of enteric viral presence.

This attribute as to the selection of a fecal indicator(s) is based on the relationship between the indicator densities in polluted waters to human-health risk. Few epidemiology studies exist correlating health risks with male-specific (F+) coliphage densities. A California beach study comparing the male-specific ssRNA phage (FRNA) and male-specific DNA phage (FDNA) densities and gastrointestinal illness rates, nausea, cough and fever suggested an association between F+ phages and illness (Colford et al., 2007). Meta-analysis of epidemiological freshwater studies reported an elevated gastrointestinal illness risk with elevated bacteriophage exposure (Wade et al., 2003). However, the meta-analysis did not

specify which type of bacteriophage was evaluated.

In conjunction with microbial source-tracking, a tiered approach to rapid and effective detection and management of pathogen risks from fecal contamination encompasses a “big picture” investigation of a contaminated area. For example, Boehm et al., (2003) applied the tiered approach to resolve a closure of a local beach populated by tourists in the following manner: (1) determine the contamination source, i.e., runoff or wastewater discharge (2) survey the potential source area, i.e., beach gull droppings, broken sewer pipes, stormwater drains and (3) apply microbial library-independent microbial source tracking to water samples. Results of the above study suggested that a suite of indicator organisms and an established FRNA threshold value with a concomitant tier could potentially yield a more accurate, precise and timely environmental site assessment.

Methods developed in this study could also be used to address waterborne transmission of infectious diseases. For example the Severe Acute Respiratory Syndrome (SARS) was disseminated through aerosolized droplets of sewage (WHO, 2003). Had a viral indicator such as FRNA been detected during surveillance studies perhaps the sewage transmission via fecal droplets may have been detected sooner and thereby prevented or minimized the outbreak. The proposed viral indicator of recreational water quality, FRNA, and the molecular assays developed here could improve 1) monitoring criteria for Total Maximum Daily Loads (TMDL), state and federal recreational water quality regulations, ambient water monitoring programs, 2) combined microbiological-epidemiological studies designed to improve water quality criteria and 3) monitoring systems for wastewaters, biosolids, beach (recreational) waters, drinking waters, irrigation waters and reuse waters.

This dissertation project concluded the following: (1) analysis of complete genomic sequences from 30 *Leviviridae* FRNA coliphage strains plus two novel JS strains support the current classification scheme of two genera, *Levivirus* and *Allolevivirus* and four distinct genogroups, I, II, III and IV (2) FRNA sequences generated in this study will triple the genetic information currently available in the national genetic database for *Leviviridae* viruses (3) this is the first report of evidence for recombination in FRNA coliphages and (4) the genogroup-specific primer sets and RT-PCR amplification procedures should facilitate improved and reliable genotyping of FRNA coliphages.

IX. Summary and Conclusions

Environmental pressures, genetic mutation rates, microbial cross-species plasmid exchange, episodic outbreaks, global health threats, i.e., SARS and Avian Flu, and natural disasters such as floods and hurricanes along with an increasing societal coastal population contribute to the need for better indicator species. Public health microbiologists are challenged to address those needs.

Various wastewater treatment processes, environmental stressors and predation may influence selection of one indicator versus another indicator species. Therefore, a suite of microbial indicator species would add confidence to water-quality public health assessment.

FRNA phages are valuable models as surrogates for enteric viruses for the following reasons: (i) structure - similar icosahedral structure (ii) size - virion diameter (iii) morphology - are almost indistinguishable, by electron microscopy, to Picornoviruses, i.e., poliovirus, enterovirus (iv) composition - most enterics contain ssRNA with the exception of Adenovirus (DNA) (v) site of replication - gastro-intestinal tract. FRNA and enteric viruses are both excreted by humans, FRNA are easily detectable and are approx 100X more abundant in wastewaters and raw sewage when compared to cytopathogenic enteric viruses (Grabow, 2001).

FRNA infect the host when bacteria are in log phase and under optimal conditions and temperatures whereas somatic phages infect a greater variety of bacteria genera, including

attachment to dead host cells. Somatic phages may replicate, under certain temperature and climate conditions, but it is very unlikely FRNA would replicate in the environment. Source-tracking of FDA phages is inconclusive although strain M13 occurs at approximately 77% in wastewaters. The presence of somatic, male-specific and *B. fragilis* phages in waters associated with animal and/or human wastes generally indicates the presence of enteric viruses and these phages potentially outnumber enteric viruses.

Rarely, if ever, does a direct correlation exist between the number of coliphages and enteric viruses at any given time. Enterics are excreted by infected humans whereas coliphages are excreted at all times. The incidence of human enterics in the form of outbreaks is seasonal and influenced by vaccination regimes. The excretion of coliphages is not affected by these occurrences.

Among various phages, FRNA are probably the best model for the presence of enteric viruses in the environment. Genogroup nucleotide and amino acid sequence alignment of FRNA phages originating from world-wide sources demonstrates the potential of FRNA to be applicable to various geographical locations and water sources.

FRNA sequences generated in this study tripled the genetic information currently available in the national genetic database for the *Leviviridae*. This additional genetic information may someday contribute to a better understanding of the basic molecular biology of these phages in terms of gene expression, control and regulation, recombination, mutations, virus-host interactions and phylogenetic relationships. In the more immediate future, this data can be applied to methods using FRNA coliphages as a fecal and viral indicator and as a source-tracking tool.

X. Recommendations for Future Research

Reverse-transcription PCR (RT-PCR) and real-time can be effectively used for source identification if specific primer sets are designed to be capable of identifying and distinguishing all members within each genogroup. Recently, reverse transcription polymerase chain reaction (RT-PCR) (Dryden et al., 2006) and reverse quantitative (RTQPCR) techniques (Kirs and Smith, 2007; Ogorzaly and Gantzer, 2006) for genotyping FRNA coliphages were developed. In these reports, primer design was based on the limited number of complete or partial nucleotide sequences available in GenBank. Thus, broad primer and probe genogroup specificity may not have been achieved due to the limited nucleotide data from the few strains available. To provide an advanced decision making tool for a comprehensive assessment of the sanitary quality of recreational water, a real-time RT-QPCR FRNA phage-based assay will be designed to allow for rapid detection and greater sensitivity. When developing real-time primers and TaqMan probes, guidelines for design included the absence of a G residue at the probe's 5' end, standard primer T_m should be 58-60 °C, probe should be 7-10 degrees higher than the primer set and the probe should be located close, just downstream from the forward primer. The large data set of 30 complete FRNA phage genomes will be used to align the members of all four genogroups. Therefore, optimal FRNA sequence target regions will be used to develop genogroup specific real-time primers and probes. Primer and probe sequences will be selected from the replicase gene for groups I

and IV and the capsid gene for groups II and III (Table 8.2). T_m 's of cleavage probes will range from 66-70 ° C, T_m 's for the primers from 58-61° C. Amplicons produced from all four primer sets will range from 99-153 bp in length. The method will incorporate a rapid heat-release procedure. To evaluate and control for false-negatives resulting from inhibition or other impurities leading to negative PCR outcomes, a non-competitive purified RNA (courtesy of Dr. Bill Burkhardt) will be incorporated as an internal RNA real-time control to account for false-negatives resulting from inhibition or other impurities leading to negative PCR outcomes. Variations in detection format, instrumentation, amplification efficiency, inhibition, various water matrix compositions, technical expertise and the step-wise process of real-time PCR will be accounted for by means of this internal control. This RNA molecule, approximately 300 nucleotides in length, possessed a unique pseudo-randomly generated ribonucleotide sequence. The primer set produces an amplicon of 149 bases. The probe will be labeled with Texas red with a Black Hole 1 quencher. Validation of this assay will begin in a phosphate buffer amended with raw sewage, representing a human source and/or feces from a variety of warm-blooded animals. Once validated with amended samples then water samples contaminated with known sources of fecal pollution will be used for the field validation.

In some recreational water samples FRNA coliphage densities would be below the limit of detection. When present in low numbers ($< 5 \times 10^2$ PFU/ml) phages need to be enriched *in vivo*. Thus, an enrichment procedure will bring the densities of these FRNA phages above the limit of detection. Briefly, 18 ml of a recreation water sample will be placed in a 37 ° C water bath. To the water sample, two ml of 10X TSB, streptomycin sulfate

(15 mg/L), ampicillin (15 mg/L) and 0.5 ml of an overnight culture of the host strain, *E. coli* HS (pFamp)R (DeBartolomeis and V.J. Cabelli, 1991) will be added. This suspension will be incubated for at least 90 min to allow the completion of one lytic cycle. Coliphage enrichment elevates the phage densities above the limit of detection as the burst size of each strain can range from between 2000 and 4500 PFU (Furuse, 1987). Therefore, the proportion of phages belonging to each genogroup may differ from what was present in the origin sample thereby introducing genogroup bias.

Although the one-step RT-PCR molecular assay designed in this study was based on full-length genomic sequences from 30 FRNA coliphages with a minimum of 5 to a maximum of 10 full-length genomic sequences/genogroup, more sequencing data is needed. In this study, phages were collected from ambient water sources, oysters, mussels, sewage and a few animal droppings (birds), swine lagoon or obtained from international collections. Future research would be to collect more phages from specific sources and, if possible, use a direct fecal swab or fresh voids. To prepare a large library of known FRNA coliphages and the respective source, collections need to be obtained directly from adults, children, pets and livestock and not mixtures of municipal sewage or livestock lagoons.

Limits of detection were not established in the traditional one-step RT-PCR, however, future plans are to perform limits of detection when developing the real-time PCR assay. To further develop the one-step RT-PCR, limits of detection will also be determined by first preparing log₁₀ serial dilutions in phosphate buffered saline (PBS) using known amounts of plaque-forming units/ml (PFU/ml) as established with titer plates. At least two different strains/genogroup, for a total of 16 FRNA strains, will be titered and serial dilutions prepared.

From the respective serial dilution, each PCR reaction will be prepared using heat-released viral RNA followed by RT-PCR. When a PCR amplicon is no longer detected the limits of the assay will have been obtained.

Future studies will be to conduct a collaborative or round-robin performance evaluation of the RT-PCR and QRT-PCR methods developed in this study. In these performance evaluations multiple laboratories and technical staff subject the coliphage molecular assays to environmental samples and also evaluate “blinded” (positive control or reference) specimens in order to verify that it is possible for the labs and their analysts to correctly detect the FRNA coliphage genotype. Collaborative studies would serve to standardize the methods and would reveal any possible deficiencies or discrepancies that may have gone unnoticed during the initial assay development.

Short-term enrichments possibly followed by centrifugation, sterile filtration, heat-release and the use of PCR thermal cycles designed for short cycling regimes could very well lead to a rapid turnaround time from sample collection to results outcome for decision making. The use of heat-release of viral RNA templates alleviates the need for cumbersome and costly RNA purification by chemical methods. Ultimately, the molecular coliphage assay may be applicable to the development and eventual use of a field-portable kit whereby all reagents would be available in one tube. The field staff would only need to add an aliquot of an enriched water sample, heat and place into a portable thermal cycler.

Epidemiology field studies such as those of the current EPA Beaches program (National Epidemiological and Environmental Assessment of Recreational Water) could readily implement the FRNA RT-PCR or QPCR into their microbiological monitoring. Data

gathered in the EPA Beaches program is analyzed to evaluate exposed-bather health-risk associations with microbial water quality parameters. If FRNA coliphage quantification and genotyping is incorporated into the Beaches study, the data would provide epidemiological information on bather health impacts in relation to the concentrations of different coliphage genotypes in water, making it possible to associate FRNA presence and concentration thresholds with bather health effects.

The FRNA genome sequences provided by this dissertation research project are applicable to the development of an indicator-based microarray or hand-held fluorogenic detector assay. A comprehensive microarray constructed from viral and bacteria indicators and pathogens could potentially detect multiple pathogens and indicators and pin-point their sources with minimal laboratory lag time. Pathogen detection kits, similar to the AgPath-ID One-Step RT-PCR (Ambion) could be designed to detect FRNA genogroup targets. Results from such assays are generated in about 1 hr using a single-tube, real-time RT-PCR. The availability of the FRNA genetic database in conjunction with the primer and probes developed and evaluated in this dissertation provides the foundation for an assortment of rapid molecular detection assays.

Future applications of molecular FRNA coliphage assays include surveillance, monitoring and quality verification of groundwater and surface waters, shellfish beds, produce, red meat and poultry, assessments of wastewater treatment plant performance and effluent quality, wastewater reuse and surveillance for contamination incidents harboring emerging waterborne pathogens in sewage and ambient water bodies, surrogate for biological warfare and surrogate for enteric virus presence and control measures. In addition to current

bacterial indicators, FRNA coliphages have the potential to serve as effective viral indicators of fecal pollution.

References

- Ashbolt NJ, WOK Grabow and M Snozzi. 2001. Indicators of microbial water quality. In: WHO Water Quality: Guidelines, Standards and Health.
- Boehm, AB, JA Fuhrman, RD Mrse and SB Grant. 2003. Tiered approach for identification of a human fecal pollution source at a recreational beach - case study at Avalon Bay, Catalina Island, CA. *Env Sci & Technol*, 37(4):673-680.
- Bosch A. 1998. Human enteric viruses in the water environment: a minireview. *Internatl Microbiol* 1:191-196.
- Byappanahalli MN and RS Fujioka. 1998. Evidence that tropical sould environment can support the growth of *Escherichia coli*. *Wat Sci Tech* 38(12):171-174.
- Cole D, SC Long and MD Sobsey. 2003. Evaluation of F+ RNA and DNA coliphages as source-specific indicators of fecal contamination in surface waters. *Appl Environ Micro* 69(11):6507-6514.
- Colford JM, Jr., TJ Wade, KC Schiff, CC Wright, JF Griffith, SK Sandhu, S Burns, M Sobsey, G Lovelace and SB Weisberg. 2007. Water quality indicators and the risk of illness at beaches with nonpoint sources of fecal contamination. *Epidemiology* 18(1):27-35.
- Dore WJ, K Henshilwood and DN Lees. 2000. Evaluation of F-specific RNA bacteriophage as a candidate human enteric virus indicator for bivalve molluscan shellfish. *Appl Environ Micro* 66(4):1280-1285.
- Durán AE, M Muniesa, L Mocé-Llivina, G Campos, J Jofre and F Lucena. 2003. Usefulness of different groups of bacteriophages as model micro-organisms for evaluating chlorination. *J Appl Microbiol*, 95:29-37.
- EPA. 1984. Health Effects Criteria for Fresh Recreational Waters. EPA-600/1-84-004.
- EPA. 1983. Health Effects Criteria for Marine Recreational Waters. EPA-600/1-80-031.
- EPA. 1986. Ambient Water Quality Criteria for Bacteria - 1986. EPA 440/5-84-002.
- EPA. 2003. Bacterial water quality standards for recreational waters (freshwater and marine waters) status report. EPA-823-R-03-008. June, 2003.
- Fujioka RS and MN Byappanahalli. 2000. Microbial ecology controls the establishment of

- fecal bacteria in tropical soil environment. Proc Int Symp of the Center of Excellence, March 2000, pp 99-108. University of Tokyo and Minister of Education, Science, Sports and Culture of Japan.
- Furuse K, S Osawa, J Kawashiro, R Tanaka, A Ozawa, S Sawamura, Y Yanagawa, T Nagao and I Watanabe. 1983. Bacteriophage distribution in human faeces: continuous survey of healthy subjects and patients with internal and leukaemic diseases. *J Gen Virol*, 64:2039-2043.
- Furuse K. 1987. Distribution of coliphages in the environment: general considerations. In: *Phage Ecology*. SM Goyal, CP Gerba and G Britton, eds. John Wiley & Sons, Inc., NY. pp 87-124.
- Genthner FJ, JB James, DF Yates and SD Friedman. 2005. Use of composite data sets for source-tracking enterococci in the water column and shoreline interstitial waters on Pensacola Beach, Florida. *Mar Poll Bull* 50:724-732.
- Grabow, W. 2001. Bacteriophages: Update on application as models for viruses in water. *Water*, SA 27: 251-268.
- Gerba, CP. 2000. Assessment of enteric pathogen shedding by bathers during recreational activity and its impact on water quality. *Quant Microbiol* (2):55-68.
- Gerba, CP. 1987. Phage as indicators of fecal pollution. In: *Phage Ecology*. SM. Goyal, CP. Gerba, and G. Britton (eds). John Wiley & Sons, Inc., NY. pp:197-209..
- Griffin D, CJ Gibson III, EK Lipp, K Riley, JH Paul, and JB Rose. 1999. Detection of viral pathogens by reverse transcriptase PCR and of microbial indicators by standard methods in the canals of the Florida Keys. *Appl Environ Micro* 65:4118-4125.
- Griffin DW, KA Donaldson, JH Paul and JB Rose. 2003. Pathogenic human viruses in coastal waters. *Clin Micro Rev* 16(1):129-143.
- Hardina CM and RS Fujioka. 1991. Soil: the environmental source of *Escherichia coli* and enterococci in Hawaii's streams. *Environ Tox Water Qual* 6:185-195.
- Leclerc H, S Edberg, V Pierzo, and JM Delattre. 2000. Bacteriophages as indicators of enteric viruses and public health risk in groundwater. *J Appl Microbiol*, 88(1):5-21.
- Long SC, SS El-Khoury, SJG Oudejans, MD Sobsey, and J Vinjé. 2005. Assessment of sources and diversity of male-specific coliphages for source tracking. *Env Eng Sci* 22(3):367-377.
- Nappier SP, MD Aiken and MD Sobsey. 2006. Male-specific coliphages as indicators of thermal inactivation of pathogens in biosolids. *Appl Environ Microbiol*, 72:2471-

- 2475.
- Osawa S, K Furuse, and I Watanabe. 1981. Distribution of ribonucleic acid coliphages in animals. *Appl Environ Microbiol* 41(1):164-168.
- Pruss A. 1998. Review of epidemiological studies on health effects from exposure to recreational water. *Int J Epidemiol*, 27:1-9.
- Roll BM and RS Fujioka. 1997. Sources of fecal indicator bacteria in a brackish tropical stream and their impact of recreational water quality. *Wat Sci Tech* 35:179-186.
- Schaper M, J Jofre, M Uys and WO Grabow. 2002. Distribution of genotypes of F-specific RNA bacteriophages in human and non-human sources of faecal pollution in South Africa and Spain. *J Appl Micro* 92(4):657-667.
- Scott TM, JB Rose, TM Jenkins, SR Farrah and J Lukasik. 2002. Microbial source tracking: current methodology and future directions. *Appl Environ Micro*, 68:5796-5803.
- Snow J. 1855. On the mode of communication of cholera. London: John Churchill, New Burlington Street, England.
- Solo-Gabriele HM, MA Wolfert, TR Desmarais and CJ Palmer. 2000. Sources of *Escherichia coli* in coastal subtropical environment. *Appl Environ Micro* 66(1):230-237.
- Stewart JR. 2002. Microbial source tracking using F+ RNA coliphage typing and *Escherichia coli* antibiotic resistance assays. Ph.D Dissertation. University of North Carolina at Chapel Hill, NC. School of Public Health, Department of Environmental Sciences and Engineering.
- Vinje J, SJG Oudejans, JR Stewart, MD Sobsey and SC Long. 2004. Molecular detection and genotyping of male-specific coliphages by reverse transcription-PCR and reverse line blot hybridization. *Appl Environ Micro* 70 (10):5996-6004.
- World Health Organization. 2003. Emerging issues in water and infectious disease. 22 pp.
- World Health Organization. 2003. Guidelines for safe recreational water environments. Vol 1. Coastal and Fresh Waters.

Alignment of Nucleotide Genomic Sequences

Appendix A1

Group I

Alignment: Levivirus Group I.

	10	20	30	40	50
Consensus	GGGTGGGACC	CCTTTTCGGGG	TCCTGCTCRA	CTTCCTGTCT	AGCTAAATGC	
DL1A.G	
DL2	-----	-----	-----	-----	-----	
DL13	-----	-----	-----	-----	-----	
DL16A.G-	
ST4A.G-	
R17A.G-	
J20A.G-	
MS2A.G-	
M12A.G-	
frG.A	

	60	70	80	90	100
Consensus	CATTTTAAAT	GTCTTTAGCG	AGACGCTACC	WTGGCTATCG	CTGTAGGTAG	
DL1	A.....	
DL2	-----	-----	-----	A.....	
DL13	-----	-----	-----	A.....	
DL16	A.....	
ST4	A.....	
R17	A.....	
J20	A.....	
MS2	A.....	
M12	T.....	
fr	A.....	

	110	120	130	140	150
				ORF1		
Consensus	CCGSAATTCC	ATTCTAGGR	RGYYTSRYBY	RYGMRAGYTY	WYASYRHYCK	
DL1	...G.....	...C....A	G.TT.GACTC	AT.CG..C.T	TC.GTGTC.T	
DL2	...G.....	...C....A	G.TT.GACTC	AT.CG..C.T	TT.GTGTC.T	
DL13	...G.....	...C....A	G.TT.GACTC	AT.CG..C.T	TT.GTGTC.T	
DL16	...G.....	...C....A	G.TT.GACTC	AT.CG..C.T	TT.GTGTC.T	
ST4	...G.....	...C....A	G.TT.GACCT	GT.CG..C.T	TT.GTACC.T	
R17	...G.....	...C....A	G.TT.GACCT	AT.CG..C.T	TT.GTGCC.T	
J20	...G.....	...C....A	G.TT.GACTC	AT.CG..C.T	TT.GTGTC.T	
MS2	...G.....	...C....A	G.TT.GACCT	GT.CG..C.T	TT.GTACC.T	
M12	...G.....	...C....A	G.TT.GACTC	AT.CG..C.T	TT.GTGTT.T	
fr	...C.....	...G....G	A.CC.CGTGT	GC.AA..T.C	AT.CCAAC.G	

	160	170	180	190	200
Consensus	WGAKMRKGAR	WVYRARASCY	WYGTSSYBHB	CRTYCGCRHH	TAYGCKRACG
DL1	T..TAAG..G	TCTG.G.C.T	TT..CCCGCT	.G.C...ACC	..T..TG...
DL2	T..TCAG..G	TCCG.G.C.T	TC..CCCTAG	.G.C...GTT	..T..GG...
DL13	T..TCAG..G	TCCG.G.C.T	TC..CCCTAG	.G.C...GTT	..T..GG...
DL16	T..TCAG..G	TCCG.G.C.T	TC..CCCTAG	.G.C...GTT	..T..GG...
ST4	T..TAGG..G	AACG.G.C.T	TC..CCCCTC	.G.T...GTT	..C..GG...
R17	T..TAAG..G	AGCA.G.C.T	TC..CCCTTC	.A.T...GTT	..C..GA...
J20	T..TAGG..A	TCCG.G.C.T	TC..CCCCTC	.G.T...GTT	..T..TG...
MS2	T..TAGG..G	AACG.G.C.T	TC..CCCCTC	.G.T...GTT	..C..GG...
M12	T..TCAG..G	AACG.A.C.T	TC..GCCCTC	.G.T...GTT	..T..GG...
fr	A..GAAT..G	--TA.G.G.C	AC..GGTCTC	.G.C...GAA	..T..TG...

	210	220	230	240	250
Consensus	GBSARVYYGA	RGATAACTCD	TTHYCBYTV	AAWTAYCGHTC	SAAYTGWCB
DL1	.CG.GGTT..	A.....G	..CT.CT.G	AA..C..T..	G..C...A.C
DL2	.GC.GGTC..	A.....G	..TT.CC.A	AA..C..T..	G..C...A.T
DL13	.GC.GGTC..	A.....G	..TT.CC.A	AA..C..C..	G..C...A.T
DL16	.GC.GGTC..	A.....G	..TT.CC.A	AA..C..C..	G..C...A.T
ST4	.TG.GACT..	A.....A	..CT.TT.A	AA..T..T..	G..C...A.T
R17	.TG.GACC..	A.....A	..CT.TT.A	AA..T..C..	G..C...A.T
J20	.GG.GGTC..	G.....A	..TT.CT.A	AA..C..T..	G..C...A.T
MS2	.TG.GACT..	A.....A	..CT.TT.A	AA..T..T..	G..C...A.T
M12	.CG.GACT..	A.....G	..TT.CC.C	AA..T..C..	G..C...A.C
fr	.GG.ACTC..	G.....T	..AC.GT.G	TT..C..A..	C..T...T.G

	260	270	280	290	300
Consensus	CCBGGYCRDT	WYAMHWSKAC	BGGKNCBMRM	ACRRADSART	GGCACTAYCC
DL1	..T..T.GT.	TT.ACTCG..	T..GG.TAGA	..GA.AC.G.T..
DL2	..C..T.GA.	TT.ATTCG..	C..GT.TAGA	..GA.AC.G.T..
DL13	..C..T.GA.	TT.ATTCG..	C..GT.TAGA	..GG.AC.G.T..
DL16	..C..T.GA.	TT.ATTCG..	C..GT.TAGA	..GG.AC.G.T..
ST4	..C..T.GT.	TT.ACTCG..	T..GG.CAAA	..GA.AC.G.C..
R17	..C..T.GT.	TT.ACTCG..	T..GG.CAGA	..GA.GC.G.T..
J20	..T..C.GA.	TT.ACTCG..	T..GA.CAGA	..GA.TC.G.T..
MS2	..C..T.GT.	TT.ACTCG..	T..GG.CAAA	..GA.AC.G.C..
M12	..G..T.GA.	TT.ACTCG..	T..GG.CAGA	..GA.AC.G.T..
fr	..G..C.AG.	AC.CAAGT..	G..TC.GCGC	..AA.GG.A.C..

	310	320	330	340	350
Consensus	VTCBYCBTAY	TCDMGDGGDG	CGHTNRRGHR	YAMDKCKVTN	GATCAAGGTD
DL1	G..CT.T..T	..TA.A..T.	..C.TA.CG.	C.CAT.GG.GG
DL2	G..CC.T..T	..TA.G..T.	..C.CA.TG.	C.CAT.GG.AG
DL13	G..CC.T..T	..TA.G..T.	..C.CA.TG.	C.CAT.GG.AG
DL16	G..CC.T..T	..TA.G..T.	..C.CA.TG.	C.CAT.GG.AG
ST4	C..TC.G..T	..AC.G..G.	..T.AA.TG.	C.CAT.GA.AG
R17	C..CC.G..T	..GC.G..G.	..T.AA.TG.	C.CGT.GA.AG
J20	G..CC.T..C	..TA.G..A.	..C.TA.TG.	C.CTT.GG.CT
MS2	C..TC.G..T	..AC.G..G.	..T.AA.TG.	C.CAT.GA.AG

M12 C..CC.T..C ..TC.G..A. ..T.GA.TG. T.CTG.GA.AG
fr A..GT.C..C ..AC.T..G. ..A.AG.AA. C.AGG.TC.TA

	360 370 380 390 400
Consensus	MBTAYRMR CG MWBKGGSWCR TCGTGGGGYC GYSMTGWCGA RGARMRWRCY
DL1	CT..TAAG.. CTCT..GT.AT. .CCC..A... G..GAAAG.C
DL2	CC..CAAG.. CTCT..GT.AT. .CCC..A... G..GAAAG.C
DL13	CC..CAAG.. CTCT..GT.AT. .CCC..A... G..GAAAG.C
DL16	CC..CAAG.. CTCT..GT.AT. .CCC..A... G..GAAAG.C
ST4	CC..CAAG.. AAGT..GT.AT. .CCC..A... G..GAAAG.C
R17	CC..TAAG.. CAGT..GT.AT. .CCC..A... G..GAAAG.C
J20	CC..CAAG.. CTCT..GT.AT. .TCC..A... G..GAAAG.T
MS2	CC..CAAG.. AAGT..GT.AT. .CCC..A... G..GAAAG.C
M12	CC..TAAG.. AAGT..GT.GT. .CCC..A... G..GAAAA.C
fr	AG..CGCA.. ATTG..CA.AC. .CGA..T... A..ACGTG.C

	410 420 430 440 450
Consensus	GGTTWYGGYW TSTCNMTCGA CGCACGYWSY TGYTAYAGCC TMTTCCCYGT
DL1AT..CT .C..AC....CTCC ..C..C.... .C.....T..
DL2TT..CT .C..AC....TTCC ..C..T.... .C.....T..
DL13TT..CT .C..AC....TTCC ..C..T.... .C.....T..
DL16TT..CT .C..AC....TTCC ..C..T.... .C.....T..
ST4TC..CT .C..CC....CTCC ..C..C.... .C.....T..
R17TT..CT .C..TC....CTCC ..C..C.... .C.....T..
J20TT..CT .C..GC....TTCC ..C..C.... .C.....T..
MS2TC..CT .C..CC....CTCC ..C..C.... .C.....T..
M12TT..TT .C..AC....CTCC ..C..C.... .C.....T..
frAT..CA .G..TA....TAGT ..T..T.... .A.....C..

	460 470 480 490 500
Consensus	HAGYCARAAY HTRACWTRSA THGAMGTRCC RMMGAACGTW GCKAAYCGSG
DL1	T..T..G..C C.G..T.AC. .T..A..G.. GCA.....T ..G..T..G.
DL2	T..T..G..T C.G..T.AC. .C..A..G.. GCA.....T ..G..C..G.
DL13	T..T..G..T C.G..T.AC. .C..A..G.. GCA.....T ..G..C..G.
DL16	T..T..G..T C.G..T.AC. .C..A..G.. GCA.....T ..G..C..G.
ST4	A..C..G..C T.G..T.AC. .C..A..G.. GCA.....T ..G..C..G.
R17	A..C..G..C T.G..T.AC. .C..A..G.. GCA.....T ..G..T..G.
J20	C..T..A..T A.G..T.AC. .C..A..G.. GCA.....T ..G..T..G.
MS2	A..C..A..C T.G..T.AC. .C..A..G.. GCA.....T ..G..C..G.
M12	A..C..G..C C.G..T.AC. .C..A..G.. GCA.....T ..G..C..G.
fr	T..T..A..C T.A..A.GG. .A..C..A.. AAC.....A ..T..T..C.

	510 520 530 540 550
Consensus	CBWCGACYGA RGTCYTRSRD AAGGTYACYC ARGGNAAYTT YAACCTTGGB
DL1	.GT....C.. A...C.GCAGC..C. .A..C..T.. C.....T
DL2	.GT....C.. A...C.GCAGC..T. .A..A..T.. C.....T
DL13	.GT....C.. A...C.GCAGC..T. .A..A..T.. C.....T
DL16	.GT....C.. A...C.GCAGC..T. .A..A..T.. C.....T
ST4	.GT....C.. A...C.GCAAC..C. .G..T..T.. T.....T
R17	.GT....C.. A...C.GCAAT..C. .A..T..T.. C.....T
J20	.GT....C.. A...C.GCAGT..C. .G..T..T.. T.....C
MS2	.GT....C.. A...C.GCAAC..C. .G..T..T.. T.....T

M12 .TA....C.. A...C.GCAGT..C. .A..G..C.. C.....G
fr .CA....T.. G...T.AGGTC..T. .A..T..T.. T.....C

	560 570 580 590 600
Consensus	GTNGCYITWG CWGARGCVMG RTCKACRGCC TCACAACSTK CGACGCAAAC
DL1	..A..CT.A. .A..G..GA. G..G..A... ..CG
DL2	..G..CC.A. .A..G..CA. G..G..A... ..CG
DL13	..G..CC.A. .A..G..CA. G..G..A... ..CG
DL16	..G..CC.A. .A..G..CA. G..G..A... ..CG
ST4	..T..TT.A. .A..G..CA. G..G..A... ..CG
R17	..C..CC.A. .A..G..CA. A..G..A... ..CG
J20	..A..TT.A. .A..G..CA. G..G..A... ..CG
MS2	..T..TT.A. .A..G..CA. G..G..A... ..CG
M12	..G..CC.T. .A..G..AA. G..G..A... ..CG
fr	..G..CC.T. .T..A..CC. G..T..G... ..GT

	610 620 630 640 650
Consensus	CATYGCYITS RTKAAGGCGT ACACYGCSGC TCGYCGCGGB AAYTGGCGCC
DL1	...T...C.C G.G..... ..T..C.. ...T....C ..C.....
DL2	...T...C.C G.G..... ..T..C.. ...T....C ..T.....
DL13	...T...C.C G.G..... ..T..C.. ...T....C ..T.....
DL16	...T...C.C G.G..... ..T..C.. ...T....C ..T.....
ST4	...T...C.C G.G..... ..T..C.. ...T....T ..T.....
R17	...T...C.C G.G..... ..T..C.. ...T....T ..T.....
J20	...T...C.C G.G..... ..T..C.. ...T....T ..T.....
MS2	...T...C.C G.G..... ..T..C.. ...T....T ..T.....
M12	...T...C.C G.G..... ..T..C.. ...T....T ..T.....
fr	...C...T.G A.T..... ..C..G.. ...C....G ..C.....

	660 670 680 690 700
Consensus	AGVCRSTCCG CTAYYTHGCS CTDAACGARR AYCGRARTT YMRDTCRAAR
DL1	..A.GC.... ..TC.C..C ..T....AG .C..A..A.. TCGG..A..A
DL2	..A.GC.... ..TC.T..C ..A....AG .T..G..G.. CCGA..G..A
DL13	..A.GC.... ..TC.T..C ..A....AG .T..G..G.. CCGA..G..A
DL16	..A.GC.... ..TC.T..C ..A....AG .T..G..G.. CCGA..G..A
ST4	..G.GC.... ..CC.T..C ..A....AG .T..A..G.. TCGA..A..A
R17	..G.GC.... ..CC.C..C ..A....AG .T..A..A.. TCGA..A..A
J20	..G.GC.... ..TC.T..C ..A....AG .T..G..A.. CCGG..A..A
MS2	..G.GC.... ..CC.T..C ..A....AG .T..A..G.. TCGA..A..A
M12	..C.GG.... ..CC.C..C ..A....AG .T..A..G.. TCGA..A..A
fr	..G.AC.... ..CT.A..G ..G....GA .T..A..A.. CAAT..G..G

	710 720 730 740 750
Consensus	YMCGTSGCVR GYAGRTGGYT GGAGTTGCAG TTCGGNTGGH TRCCRCTHMT
DL1	CA...G..GG .C..G...T.C...T .A..G..CA.
DL2	CA...G..AG .C..G...T.C...C .A..G..CA.
DL13	CA...G..AG .C..G...T.C...C .A..G..CA.
DL16	CA...G..AG .C..G...T.C...C .A..G..CA.
ST4	CA...G..CG .C..G...T.T...T .A..A..AA.
R17	CA...G..CG .C..G...T.A...T .A..A..AA.
J20	CA...G..AG .T..A...T.T...C .A..G..TA.
MS2	CA...G..CG .C..G...T.T...T .A..A..AA.

M12	CA...G..CG	.C..G...T.C...T	.A..A..TA.
fr	TC...C..AA	.C..G...C.G...A	.G..G..TC.

	760770780790800
Consensus	SAGYGATATC CARGGYGCRT AYGAGATGCT YACSAARGTK CAYCTTMARG
DL1	G..C..... ..G..C..A. .T..... T..G..G..T ..C...C.A.
DL2	G..C..... ..G..C..A. .T..... T..G..G..T ..C...C.A.
DL13	G..C..... ..G..C..A. .T..... T..G..G..T ..C...C.A.
DL16	G..C..... ..G..C..A. .T..... T..G..G..T ..C...C.A.
ST4	G..T..... ..G..T..A. .T..... T..G..G..T ..C...C.A.
R17	G..T..... ..A..T..A. .T..... T..G..G..T ..C...C.A.
J20	G..C..... ..A..T..A. .T..... T..G..G..T ..T...C.A.
MS2	G..T..... ..G..T..A. .T..... T..G..G..T ..C...C.A.
M12	G..T..... ..G..C..A. .C..... T..G..G..T ..C...C.A.
fr	C..C..... ..A..T..G. .T..... C..C..A..G ..T...A.G.

	810820830840850
Consensus	MRTTTMTBCC TATGMGDGCC GTRMGNCARG TNGGHMMWAA CRTYARKTTR
DL1	AG...C.C..A.A... ..GC.C..G. .A..CACT.. .G.C.AG..A
DL2	AG...C.T..A.G... ..AC.G..A. .G..CACT.. .A.T.AG..A
DL13	AG...C.T..A.G... ..AC.G..A. .G..CACT.. .A.T.AG..A
DL16	AG...C.T..A.G... ..AC.G..A. .G..CACT.. .A.T.AG..A
ST4	AG...C.T..A.A... ..AC.T..G. .C..TACT.. .A.C.AG..A
R17	AG...C.T..A.A... ..AC.T..A. .T..TACT.. .A.T.AG..A
J20	AG...C.T..A.A... ..AC.A..G. .G..TACT.. .G.T.AG..A
MS2	AG...C.T..A.A... ..AC.T..G. .C..TACT.. .A.C.AG..A
M12	AG...C.C..A.A... ..AC.T..G. .C..CACT.. .A.T.AG..A
fr	CA...A.G..C.T... ..GA.G..A. .C..ACAA.. .G.C.GT..G

	860870880890900
Consensus	DMTGGCCGBY TBDCKTMTCC RGCTGCAARC TWYMARWCWA CGTGCAACAT
DL1	GA.....CC .GT.G.A... A.....A. .ACC.AA.T.
DL2	GA.....TT .GT.G.A... A.....A. .ACC.GA.T.
DL13	GA.....TT .GT.G.A... A.....A. .ACC.GA.T.
DL16	GA.....TT .GT.G.A... A.....A. .ACC.GA.T.
ST4	AA.....TC .GT.G.A... A.....A. .TCC.GA.A.
R17	GA.....CT .GG.G.A... A.....A. .TCC.GA.A.
J20	GA.....CT .GT.G.A... A.....A. .ACC.GA.T.
MS2	GA.....TC .GT.G.A... A.....A. .TCC.GA.A.
M12	GA.....CC .TT.G.A... A.....A. .ACC.GA.A.
fr	TC.....GC .CA.T.C... G.....G. .ATA.GT.T.

	910920930940950
Consensus	ATCRCGACGH ATYGTGATAT GGTTTTACAT AAACGATGCA CGWYTGGCHT
DL1	...G.....T ..C..... ..G..... ..TT....T.
DL2	...A.....A ..C..... ..G..... ..TT....C.
DL13	...A.....A ..C..... ..G..... ..TT....C.
DL16	...A.....A ..C..... ..G..... ..TT....C.
ST4	...G.....T ..C..... ..G..... ..TT....A.
R17	...A.....T ..C..... ..G..... ..TT....A.
J20	...G.....A ..C..... ..G..... ..TT....C.
MS2	...G.....T ..C..... ..G..... ..TT....A.

M12	...G.....T ..C.....AT....C.
fr	...A.....C ..T.....TC....T.

	960 970 980 990 1000
Consensus	GGYTGTCTC YYTRGGKATY TTGAACCCRC TAGGWATAGT GTGGGAAAAG
DL1	..T....G.. TC.A..T..CA.T.....
DL2	..T....G.. TC.A..T..CA.T.....
DL13	..T....G.. TC.A..T..CA.T.....
DL16	..T....G.. TC.A..T..CA.T.....
ST4	..T....G.. TC.A..T..CA.T.....
R17	..T....G.. TC.A..T..CA.T.....
J20	..T....G.. TC.A..T..CA.T.....
MS2	..T....G.. TC.A..T..CA.T.....
M12	..T....G.. TC.G..T..CA.T.....
fr	..C....C.. CT.A..G..TG.A.....

	1010 1020 1030 1040 1050
Consensus	GTSCCBTTCT CWTTCSTKGT CGAYTGGYTS CTDCCKGTDG GDAACATGCT
DL1	..G..T.... .A...G.T.. ...C...C.C ..T..T..A. .A.....
DL2	..G..T.... .A...G.T.. ...C...C.C ..T..T..T. .G.....
DL13	..G..T.... .A...G.T.. ...C...C.C ..T..T..T. .G.....
DL16	..G..T.... .A...G.T.. ...C...C.C ..T..T..T. .G.....
ST4	..G..T.... .A...G.T.. ...C...C.C ..A..T..A. .T.....
R17	..G..T.... .A...G.T.. ...C...C.C ..A..T..A. .T.....
J20	..G..T.... .A...G.T.. ...C...C.C ..T..T..A. .A.....
MS2	..G..T.... .A...G.T.. ...C...C.C ..A..T..A. .T.....
M12	..G..C.... .A...G.T.. ...C...C.C ..G..T..G. .G.....
fr	..C..G.... .T...C.G.. ...T...T.G ..G..G..T. .G.....

	1060 1070 1080 1090 1100
Consensus	HGAGGGSCTH ACVGCYCCVR TDGGMTGYTC BTAYMWRTCD GGRACMGTTWA
DL1	C.....C..A ..G..C..CG .A..A..T.. T..CATG..G ..A..A..T.
DL2	C.....C..C ..A..C..CG .A..A..C.. C..CATG..T ..A..A..T.
DL13	C.....C..C ..A..C..CG .A..A..C.. C..CATG..T ..A..A..T.
DL16	C.....C..C ..A..C..CG .A..A..C.. C..CATG..T ..A..A..T.
ST4	C.....C..T ..G..C..CG .G..A..C.. C..CATG..A ..A..A..T.
R17	C.....C..T ..G..T..CG .T..A..C.. C..CATG..A ..A..A..T.
J20	C.....C..T ..A..C..CG .A..A..C.. C..CATG..T ..A..A..T.
MS2	C.....C..T ..G..C..CG .G..A..C.. C..CATG..A ..A..A..T.
M12	A.....C..T ..G..T..AG .A..A..T.. T..CATG..A ..G..A..T.
fr	T.....G..T ..C..C..GA .A..C..T.. G..TCAA..G ..A..C..A.

	1110 1120 1130 1140 1150
Consensus	CYGACGTAAT AWCRRGGWGAG TCSAYMATAA SCGYYGAYGM YMYCTAYGGK
DL1	.T..... .A.G..T... ..C.TC.... G..TT..C.C TCC...T..G
DL2	.T..... .A.G..T... ..C.TC.... G..TC..C.C TCC...T..G
DL13	.T..... .A.G..T... ..C.TC.... G..TC..C.C TCC...T..G
DL16	.T..... .A.G..T... ..C.TC.... G..TC..C.C TCC...T..G
ST4	.T..... .A.G..T... ..C.TC.... G..TT..C.C TCC...C..G
R17	.T..... .A.G..T... ..C.TC.... G..TT..C.C TCC...T..G
J20	.T..... .A.G..T... ..C.TC.... G..TT..C.C TCC...T..G
MS2	.T..... .A.G..T... ..C.TC.... G..TT..C.C TCC...C..G

M12 .T..... .A.G..T... ..C.TC.... G..TT..C.C TCC...C..G
fr .C..... .T.A..A... ..G.CA.... C..CC..T.A CAT...T..T

	1160 1170 1180 1190 1200
Consensus	TGGRMDRYRG WKMGACMKGS MACYGCTAAG GYSCADRTYW SDGCVRTSCA
DL1	...ACAGTG. AGA...AG.G C..T..... .CC..GA.CT CA..CA.G..
DL2	...ACGGTG. ATA...AG.G C..T..... .CC..AG.CT CA..CA.G..
DL13	...ACGGTG. ATA...AG.G C..T..... .CC..AG.CT CA..CA.G..
DL16	...ACGGTG. ATA...AG.G C..T..... .CC..AG.CT CA..CA.G..
ST4	...ACTGTG. AGA...AG.G C..T..... .CC..AA.CT CA..CA.G..
R17	...ACTGTG. AGA...AG.G C..T..... .CC..TG.TT CA..CA.G..
J20	...ACTGTG. AGA...AG.G C..T..... .CC..GA.CT CG..CA.G..
MS2	...ACTGTG. AGA...AG.G C..T..... .CC..AA.CT CA..CA.G..
M12	...ACTGTG. AGA...AG.G C..T..... .CC..AG.CT CA..CA.G..
fr	...GATACA. TGC...CT.C A..C..... .TG..AA.CA GT..TG.C..

	1210 1220 1230 1240 1250
Consensus	YCGRGGGGTR CARWSCGTRT GSCCMACWAC KGGCGYRTAC GTDAARTCWC
DL1	T..A.....A ..GTC...A. .G..A..A.. T....TA... ..A..G..A.
DL2	T..A.....G ..ATC...A. .G..A..A.. T....TA... ..G..G..A.
DL13	T..A.....G ..ATC...A. .G..A..A.. T....TA... ..G..G..A.
DL16	T..A.....G ..ATC...A. .G..A..A.. T....TA... ..G..G..A.
ST4	T..A.....A ..ATC...A. .G..A..A.. T....CG... ..A..G..T.
R17	T..A.....A ..ATC...A. .G..A..A.. T....CA... ..A..G..T.
J20	T..A.....G ..ATC...A. .G..A..A.. T....TA... ..G..A..A.
MS2	T..A.....A ..ATC...A. .G..A..A.. T....CG... ..A..G..T.
M12	T..A.....A ..ATC...A. .C..A..T.. T....TA... ..A..G..T.
fr	C..G.....A ..AAG...G. .G..C..A.. G....TA... ..T..G..A.

	1260 1270 1280 1290 1300
Consensus	CYTTYTCGAT KGTCCAYACY TTAGAYGCST TRGCAYTWWT CAGGCAACGS
DL1	.T..C..... G.....C..TT..G. .G...T.AA.G
DL2	.T..C..... G.....T..CC..G. .G...T.AA.G
DL13	.T..C..... G.....T..CC..G. .G...T.AA.G
DL16	.T..C..... G.....T..CC..G. .G...T.AA.G
ST4	.T..C..... G.....T..CT..G. .A...T.AA.G
R17	.T..C..... G.....T..CT..G. .A...T.AA.G
J20	.T..C..... T.....T..CT..G. .A...T.AA.G
MS2	.T..C..... G.....T..CT..G. .A...T.AA.G
M12	.C..T..... G.....T..CT..G. .G...T.AA.G
fr	.T..C..... G.....T..CT..C. .G...C.TT.C

		STOP 1		ORF2	
				
	1310 1320 1330 1340 1350				
Consensus	CTCTBDARAT AGAGSCCYYA ACCGRAGKKH GARCCRCATG GCTTCKAACT				
DL1CA.G..C..TC.G..TTC ..A--G....T....				
DL2TA.G..C..TC.G..TTC ..A--G....T....				
DL13TA.G..C..TC.G..TTC ..A--G....T....				
DL16TA.G..C..TC.G..TTC ..A--G....T....				
ST4CT.G..C..TC.G..TTT ..A--G....T....				
R17CT.A..C..TC.G..TTT ..A--G....T....				
J20CA.G..C..TC.G..TTT ..A--G....T....				
MS2CT.G..C..TC.G..TTT ..A--G....T....				
M12CT.A..C..TC.G..TTT ..A--G....T....				

fr GG . A G . . CT A . . GGA . . G . . A G

	13601370138013901400
Consensus	TTRMWSAGTT YGTTCTCGTC GACAATGGCG GAACBGGHGA YGTRAMWGTC
DL1	..ACTC.... C.....
DL2	..ACTC.... C.....
DL13	..ACTC.... C.....
DL16	..ACTC.... C.....
ST4	..ACTC.... C.....
R17	..ACTC.... T.....
J20	..ACTC.... T.....
MS2	..ACTC.... C.....
M12	..ACTC.... T.....
fr	..GAAG.... C.....

	14101420143014401450
Consensus	GCBCCRAGCA ACTTCGCTAA CGGGGTGCGW GAATGGATYA GCTCBAACTC
DL1	..C..A....
DL2	..T..A....
DL13	..T..A....
DL16	..T..A....
ST4	..C..A....
R17	..T..A....
J20	..T..A....
MS2	..C..A....
M12	N.G..A....
fr	..T..G....

	14601470148014901500
Consensus	VCGYTCWCAR GCTTACAAAG TRACYTGTAG YGTKCGTCAG AGCTCTGCGM
DL1	G..C..T..G
DL2	A..T..T..G
DL13	A..T..T..G
DL16	A..T..T..G
ST4	G..T..A..G
R17	G..C..A..G
J20	A..C..T..G
MS2	G..T..A..G
M12	C..C..A..A
fr	A..T..T..G

	15101520153015401550
Consensus	ASAAYCGSAA RTACACYRTY AARGTYGARG TRCCDAARGT GGCWACYCAR
DL1	.G..T..C.. G.....TA.C ..G..C..A. .G..G..A.. ...T..C..G
DL2	.G..C..C.. G.....CA.T ..G..T..G. .A..A..A.. ...T..C..A
DL13	.G..C..C.. G.....CA.T ..G..T..G. .A..A..A.. ...T..C..A
DL16	.G..C..C.. G.....CA.T ..G..T..G. .A..A..A.. ...T..C..A
ST4	.G..T..C.. A.....CA.C ..A..C..G. .G..T..A.. ...A..C..G
R17	.G..T..C.. A.....CA.T ..A..C..G. .G..T..G.. ...A..T..G
J20	.G..C..C.. G.....CA.C ..A..C..A. .G..A..A.. ...T..T..G
MS2	.G..T..C.. A.....CA.C ..A..C..G. .G..T..A.. ...A..C..G

M12 .G..C..C.. A.....CA.C ..G..C..G. .G..G..A.. ...A..C..A
fr .C..T..G.. A.....CG.C ..G..C..G. .G..G..A.. ...A..T..G

	15601570158015901600
Consensus	RYYSWHGGYG GYGTWSAGCT TCCTGTWGCs GCRTGGCGYT CGTAYHTRAA
DL1	ACCGTT..T. .C..AC....A..C ..G.....T.TC.G..
DL2	ACCGTC..T. .T..AC....A..C ..A.....T.CT.G..
DL13	ACCGTC..T. .T..AC....A..C ..A.....T.CT.G..
DL16	ACCGTC..T. .T..AC....A..C ..A.....T.CT.G..
ST4	ACTGTT..T. .T..AG....A..C ..A.....T.CT.A..
R17	ACTGTT..T. .T..AG....A..C ..A.....T.CT.A..
J20	ACCGTC..T. .C..AC....A..C ..A.....T.CT.A..
MS2	ACTGTT..T. .T..AG....A..C ..A.....T.CT.A..
M12	ACCGTT..C. .T..AG....A..C ..A.....T.CC.G..
fr	GTCCAA..C. .C..TG....T..G ..G.....C.CA.G..

	16101620163016401650
Consensus	TATGGAAYTR ACYATTCCDR THHTCGCDAC GAAYKMCGAY TGYGMSYTWA
DL1T.G ..T.....AA .C.....A.. ...CGA...C ..C.CGC.A.
DL2T.A ..T.....TA .T.....A.. ...CGA...T ..C.CGC.T.
DL13T.A ..T.....TA .T.....A.. ...CGA...T ..C.CGC.T.
DL16T.A ..T.....TA .C.....A.. ...CGA...T ..C.CGC.T.
ST4C.A ..C.....AA .T.....T.. ...TTC...C ..C.AGC.T.
R17T.A ..T.....AA .T.....T.. ...CTC...T ..C.AGC.T.
J20T.A ..T.....AA .T.....G.. ...CGA...T ..C.CGC.T.
MS2C.A ..C.....AA .T.....T.. ...TTC...C ..C.AGC.T.
M12T.A ..T.....GA .T.....T.. ...CTC...T ..C.CGC.T.
frT.A ..T.....GG .A.....G.. ...CGA...C ..T.CCT.A.

ORF3

	16601670168016901700
Consensus	TYGTYAARGC RWTGCAAGGY MYCYTDAAAR MTGWAACCC RATYSCHWCR
DL1	.T..T..G.. GA.....T CT.C.A...G A...A..... G..CC.CT.A
DL2	.T..T..G.. GA.....T CT.C.A...G A...A..... G..CC.CT.A
DL13	.T..T..G.. GA.....T CT.C.A...G A...A..... G..CC.CT.A
DL16	.T..T..G.. GA.....T CT.C.A...G A...A..... G..CC.CT.A
ST4	.T..T..G.. AA.....T CT.C.A...G A...A..... G..TC.CT.A
R17	.T..T..G.. AA.....T CT.C.A...G A...A..... G..TC.CT.A
J20	.T..T..G.. GA.....T CT.C.A...G A...A..... A..CC.CT.A
MS2	.T..T..G.. AA.....T CT.C.A...G A...A..... G..TC.CT.A
M12	.T..C..A.. AA.....T CT.C.G...G A...A..... G..TC.TT.G
fr	.C..T..G.. AT.....C AC.T.T...A C...T..... A..TG.AA.A

STOP 2

	17101720173017401750
Consensus	GCMATCGCAG CMAACTCSGG MATCTAYTAA KARAYNYSKK CCATTCMAAC
DL1	..A..... .A.....C.. A.....C... T.G.TTTCGGA...
DL2	..A..... .A.....C.. A.....C... T.G.TTCCGTC...
DL13	..A..... .A.....C.. A.....C... T.G.TTCCGTC...
DL16	..A..... .A.....C.. A.....C... T.G.TTCCGTC...
ST4	..A..... .A.....C.. C.....C... T.G.CGCCGGA...
R17	..A..... .A.....C.. C.....C... T.G.TGCCGGA...
J20	..A..... .C.....C.. A.....T... T.G.TTCCGGC...
MS2	..A..... .A.....C.. C.....C... T.G.CGCCGGA...
M12	..A..... .A.....C.. C.....C... T.G.TACCGGA...

fr ..C..... .C.....G.. A.....C... G.A.CCCGTGC...

ORF4					
				
	1760	1770	1780	1790	1800
Consensus	ATGAGGAWTA	CCC ATG TCRA	ARWCAACAAA	GAAGTTCAAC	TCTTTATGTA
DL1T..G.	.GA.....
DL2T..G.	.GA.....
DL13T..G.	.GA.....
DL16T..G.	.GA.....
ST4T..G.	.GA.....
R17T..G.	.GA.....
J20T..G.	.GA.....
MS2T..G.	.GA.....
M12T..G.	..A.....
frA..A.	.AT.....
				
	1810	1820	1830	1840	1850
Consensus	TTGATYTKYC	TYGCGATCTT	TCTCTCGAAR	TTTACCAATC	AATTGCTWSY
DL1C.TC.	.C.....ATCT
DL2C.TC.	.C.....ATCT
DL13C.TC.	.C.....ATCT
DL16C.TC.	.T.....ATCT
ST4C.TC.	.C.....ATCT
R17C.TC.	.C.....ATCT
J20C.TC.	.C.....ATCT
MS2C.TC.	.C.....ATCT
M12C.TC.	.C.....ATCT
frT.GT.	.C.....GAGC
				
	1860	1870	1880	1890	1900
Consensus	GTCGCTACTG	GAWSYKSTRA	TCCGCAYAGT	RRMGACTTTW	CRGCAATTGC
DL1AGCGG.G.C...	AGA.....A	.A.....
DL2AGCGG.G.C...	AGA.....A	.A.....
DL13AGCGG.G.C...	AGA.....A	.A.....
DL16AGCGG.G.C...	AGA.....A	.A.....
ST4AGCGG.G.C...	GAC.....A	.A.....
R17AGCGG.A.C...	GAC.....A	.A.....
J20AGCGG.G.C...	AGA.....A	.G.....
MS2AGCGG.G.C...	GAC.....A	.A.....
M12TGCGG.A.C...	GAC.....T	.A.....
frTCTTC.G.T...	GAC.....A	.A.....
STOP 3					
				
	1910	1920	1930	1940	1950
Consensus	TTACY TAA GR	GACGARYTKY	THACWAAGCA	YCCSWMBYTA	GGNDMYGGTA
DL1C....GGT.GC	.A..T....	T..CTCCT..	..CAAT....
DL2C....AAT.GC	.A..T....	T..GTCCT..	..AAAT....
DL13C....AAT.GC	.A..T....	T..GTCCT..	..AAAT....
DL16C....AAT.GC	.A..T....	T..GTCCT..	..AAAT....
ST4T....GAT.GC	.C..A....	T..GACCT..	..TTCT....
R17C....GAT.GC	.T..A....	T..GACTT..	..CTCT....
J20C....AGT.GC	.C..A....	T..GTCTT..	..AAAT....
MS2T....GAT.GC	.C..A....	T..GACCT..	..TTCT....
M12C....GGT.GC	.T..T....	C..GACGT..	..TTCC....

fr C G GC.TT .A..A. T..GAACC.. ..GGAT. . . .

	19601970198019902000
Consensus	AYGACGARGC RACCCGYMGN RSYTAGCTA TYGCTAAGCT NCKGGAGGCG
DL1	.T.....G.. G.....CC.A GCTT..... .T..... T.G.....
DL2	.T.....G.. G.....CC.G GCTC..... .C..... C.G.....
DL13	.T.....G.. G.....CC.G GCTC..... .C..... C.G.....
DL16	.T.....G.. G.....CC.G GCTC..... .C..... C.G.....
ST4	.T.....G.. G.....TC.T ACCT..... .C..... A.G.....
R17	.T.....G.. G.....TC.C ACCT..... .T..... G.G.....
J20	.T.....G.. G.....CC.G GCTT..... .C..... C.G.....
MS2	.T.....G.. G.....TC.T ACCT..... .C..... A.G.....
M12	.T.....G.. G.....CC.T GCTT..... .T..... A.G.....
fr	.C.....A.. A.....CA.A AGCC..... .C..... G.T.....

	20102020203020402050
Consensus	AATGRNSRDY GYGGYCAGAT HAAYAGRGAD GGTTTCTTAC AYGAYRMAWC
DL1AACGAT .T..C..... C..T..A..GT..CAA.T.
DL2AGCGGT .C..C..... T..T..G..AC..TAA.T.
DL13AGCGGT .C..C..... T..T..G..AC..TAA.T.
DL16AGCGGT .C..C..... T..T..G..AC..TAA.T.
ST4ATCGGT .C..T..... A..T..A..AT..CAA.T.
R17ATCGGT .C..C..... A..T..A..AT..CAA.T.
J20AACGAT .T..C..... T..C..A..AT..CAA.T.
MS2GTGATC .C..T..... A..T..A..AT..CAA.T.
M12ATCGAT .T..C..... T..C..A..AT..CAA.T.
frACCGTT .T..T..... A..T..G..TT..CGC.A.

	20602070208020902100
Consensus	CKYRTCRTGG GATCCGGATG TTTTACAAAC CAGCATCCGT AGCCTWATHG
DL1	.TTG..A...T..T.
DL2	.TTA..G...T..T.
DL13	.TTA..G...T..T.
DL16	.TTA..G...T..T.
ST4	.TTG..A...T..T.
R17	.TTG..A...T..C.
J20	.TTG..G...T..T.
MS2	.TTG..A...T..T.
M12	.TTG..G... A..A.
fr	.GCG..G...T..T.

	21102120213021402150
Consensus	GYAAYCTYCT YTCTGGYTAY MRHWSKYMGT TGTTTRGRCA MTGYACRTTY
DL1	.C..C..T.. C.....C..T CGTTCGTC..G.G.. A..C..G..C
DL2	.C..C..T.. C.....C..C CGCTCGTC..G.G.. A..C..G..C
DL13	.C..C..T.. C.....C..C CGCTCGTC..G.G.. A..C..G..C
DL16	.C..C..T.. C.....C..C CGCTCGTC..G.G.. A..C..G..C
ST4	.C..C..C.. C.....C..C CGATCGTC..G.G.. A..C..G..C
R17	.C..T..T.. C.....C..C CAATCGTC..G.G.. A..C..G..C
J20	.C..C..T.. C.....C..T CGTTCGTC..G.G.. A..C..G..T
MS2	.C..C..C.. C.....C..C CGATCGTC..G.G.. A..C..G..C

M12 .C..C..C.. C.....T..C CGATCGTC..G.G.. A..C..G..C
fr .T..C..C.. T.....C..T AGCAGTCA..A.A.. C..T..A..C

	21602170218021902200
Consensus	TCMAACGGTG CYYCDATGGG GCACAAGTTG CAGGATGCAG CGCCTTAYAA
DL1	..C..... .CT.G.....
DL2	..C..... .CT.A.....
DL13	..C..... .CT.A.....
DL16	..C..... .CT.A.....
ST4	..C..... .CT.T.....
R17	..C..... .CT.T.....
J20	..C..... .CT.T.....
MS2	..C..... .TC.T.....
M12	..C..... .CT.G.....
fr	..A..... .CT.T.....

	22102220223022402250
Consensus	GAAGTTCGCT GAACAAGCAA CCGTKACSCC SMGSGCKYTR ARAGCGGCNB
DL1
DL2
DL13
DL16
ST4
R17
J20
MS2
M12
fr

	22602270228022902300
Consensus	TRYTGGTGTCMR AGAYCARTGY RBKCCSTGGA TYMGWCAKCK GSWCSDCTWY
DL1	.AC.....CG ...C..G..T GTG..G.... .TA.A...G. .GT.CG..AC
DL2	.AC.....CG ...C..G..T GTG..G.... .CA.A...G. .GT.CG..AC
DL13	.AC.....CG ...C..G..T GTG..G.... .CA.A...G. .GT.CG..AC
DL16	.AC.....CG ...C..G..T GTG..G.... .CA.A...G. .GT.CG..AC
ST4	.AT.....CG ...C..A..T GCG..G.... .CA.A...G. .GT.CG..AT
R17	.AT.....CG ...C..A..T GCG..G.... .CA.A...G. .GT.CA..AT
J20	.AC.....AG ...C..G..T GCG..G.... .CA.A...G. .GT.CG..AT
MS2	.AT.....CG ...C..A..T GCG..G.... .CA.A...G. .GT.CG..AT
M12	.GC.....AG ...C..G..T GCG..G.... .TA.A...G. .GT.CG..AC
fr	.GC.....AA ...T..G..C AGT..C.... .CC.T...T. .CA.GT..TC

	23102320233023402350
Consensus	MMYGARTCAT ATRMATTTAG RCTCGTBGKA GGSAAACGGWG TGTTYACWGT
DL1	AAT..G.... ..GA..... G.....T.T. ..G.....A.T..A..
DL2	AAC..G.... ..GA..... G.....C.T. ..G.....A.T..A..
DL13	AAC..G.... ..GA..... G.....C.T. ..G.....A.T..A..
DL16	AAC..G.... ..GA..... G.....C.T. ..G.....A.T..A..
ST4	AAC..G.... ..GA..... G.....T.T. ..G.....A.T..A..
R17	AAC..G.... ..GA..... G.....T.T. ..G.....A.T..A..
J20	AAC..A.... ..AA..... A.....T.T. ..G.....A.T..A..
MS2	AAC..G.... ..GA..... G.....T.T. ..G.....A.T..A..

M12	AAC..G.....	..GA.....	G.....T.T.	..G.....A.C..A..
fr	CCC..G.....	..AC.....	G.....G.G.	..C.....T.T..T..

	23602370238023902400
Consensus	TCCGAAGAAY AATAAAATAG ATCGGGCTGC YTGYAARGAG CCYGATATGA
DL1T.....C.....
DL2T.....C.....
DL13T.....C.....
DL16T.....C.....
ST4T.....T.....
R17T.....T.....
J20T.....C.....
MS2T.....C.....
M12T.....T..C..A...
frC.....T..C..A...

	24102420243024402450
Consensus	ATATGTACYT HCAGAAAGGR GTHGGHGSYT TYATHMGDCG YCGBCTYARR
DL1C. T.....G ..T..C.CC. .C..TA.A.. C..C..C.AA
DL2C. C.....A ..T..C.CC. .C..AA.G.. T..C..C.AA
DL13C. C.....A ..T..C.CC. .C..AA.G.. T..C..C.AA
DL16C. C.....A ..T..C.CC. .C..AA.G.. T..C..C.AA
ST4C. C.....G ..C..T.CC. .T..CA.A.. C..G..C.AA
R17C. C.....G ..C..C.CT. .T..TA.A.. C..G..C.AA
J20C. T.....A ..T..C.CT. .T..AC.G.. T..T..C.GA
MS2C. C.....G ..C..T.CT. .C..CA.A.. C..G..C.AA
M12C. T.....A ..A..A.CC. .C..CA.G.. C..T..C.AA
frT. A.....G ..C..C.GT. .C..CC.T.. C..C..T.AG

	24602470248024902500
Consensus	WCYGTBGGTA TAGAYYTGAA YGATCARWCG ATCAAYCARC KYITRGCTCA
DL1	T.T..C.... ..CC.... T....AA..T..G. GTC.A.....
DL2	T.T..C.... ..CC.... T....AA..T..G. GTT.A.....
DL13	T.T..C.... ..CC.... T....AA..T..G. GTT.A.....
DL16	T.T..C.... ..CC.... T....AA..T..G. GTT.A.....
ST4	T.C..T.... ..TC.... T....AT..C..G. TTC.G.....
R17	T.C..T.... ..CC.... T....GT..C..G. GTC.A.....
J20	T.C..T.... ..CC.... T....AA..C..G. GTC.G.....
MS2	T.C..T.... ..CC.... T....AT..C..G. GTC.G.....
M12	T.C..T.... ..CC.... T....AA..C..G. GTC.G.....
fr	A.T..G.... ..CT.... C....AA..T..A. GCC.G.....

	25102520253025402550
Consensus	RCARGGYAGY VBHGATGGNT CDYTDGCRAC KATAGAYYTA TCGTCNGCNT
DL1	A..G..C..C ATA....T. .TT.A..G.. G....CC..A..T.
DL2	A..G..C..C ATA....C. .TT.A..A.. G....TT..G..T.
DL13	A..G..C..C ATA....C. .TT.A..A.. G....TT..G..T.
DL16	A..G..C..C ATA....C. .TT.A..A.. G....TT..G..T.
ST4	G..G..C..C GTA....T. .GC.T..G.. G....CT..T..A.
R17	G..A..C..C GCA....T. .GC.T..G.. G....CT..C..G.
J20	A..A..C..C GTC....C. .AT.G..G.. G....CT..G..C.
MS2	G..G..C..C GTA....T. .GC.T..G.. G....CT..T..A.

M12 A..A..C..T ATA.....A. .GT.A..G.. T.....CT..T..A.
fr A..A..T..C CGT.....G. .TT.G..G.. G.....CT..T..T.

	25602570258025902600
Consensus	CBGAYTCYAT YWSYGAYCGC CTRGTGTGGA RYTTYCTYCC ACCTGAGYTA
DL1	.G..T..C.. CTCT..C... ..G..... GC..T..C..C..
DL2	.T..T..T.. CTCT..C... ..G..... GT..C..C..T..
DL13	.T..T..T.. CTCT..C... ..G..... GT..C..C..T..
DL16	.T..T..T.. CTCT..C... ..G..... GT..C..C..T..
ST4	.C..T..C.. CTCC..T... ..G..... GT..T..C..C..
R17	.C..T..C.. TTCC..C... ..A..... AT..C..T..C..
J20	.T..C..C.. CTCT..C... ..G..... GT..C..C..C..
MS2	.C..T..C.. CTCC..T... ..G..... GT..T..C..C..
M12	.C..T..C.. CTCT..C... ..G..... GT..C..C..C..
fr	.T..C..C.. CAGC..C... ..A..... GT..T..C..C..

	26102620263026402650
Consensus	TAYTCATATC TYGAYMKKAT YCGMWSVCAC TAYGGWWWCR TARATGGNRA
DL1	..T..... .C..TCGT.. C..CTCC... ..C..AAT.G ..G....CG.
DL2	..T..... .C..TCGT.. C..CTCC... ..C..AAT.G ..G....GG.
DL13	..T..... .C..TCGT.. C..CTCC... ..C..AAT.G ..G....GG.
DL16	..T..... .C..TCGT.. C..CTCC... ..C..AAT.G ..G....GG.
ST4	..T..... .C..TCGT.. C..CTCA... ..C..AAT.G ..G....CG.
R17	..T..... .T..TCGT.. C..CTCG... ..C..AAT.G ..G....CG.
J20	..T..... .C..TCGT.. C..CTCC... ..T..AAT.A ..G....AG.
MS2	..T..... .C..TCGT.. C..CTCA... ..C..AAT.G ..G....CG.
M12	..C..... .C..TCGT.. C..CTCC... ..C..AAT.G ..G....TG.
fr	..T..... .C..CATG.. T..AAGC... ..C..TTA.G ..A....CA.

	26602670268026902700
Consensus	GAYGATWCGD TGGGAACATAT TTTCACRAT GGGWAAYGGG TTYACNTTYG
DL1	..C...A..GC..A.. ...A..T... ..C..T..C.
DL2	..C...A..AC..A.. ...T..T... ..C..T..C.
DL13	..C...A..AC..A.. ...T..T... ..C..T..C.
DL16	..C...A..AC..A.. ...T..T... ..C..T..C.
ST4	..C...A..AC..A.. ...A..T... ..C..G..T.
R17	..C...A..AC..G.. ...A..T... ..T..A..T.
J20	..C...A..AC..A.. ...A..T... ..C..T..T.
MS2	..C...A..AC..A.. ...A..T... ..C..A..T.
M12	..C...A..AC..A.. ...A..C... ..C..T..T.
fr	..T...T..TG..G.. ...T..T... ..C..C..T.

	27102720273027402750
Consensus	ARCTAGAGTC CATGATHTTY TGGGCWATAG TVARRGCDAC YCARATCCAT
DL1	.A..... ..A..TT.... .G.AA..G.. C..G.....
DL2	.A..... ..C..TT.... .G.AA..A.. C..G.....
DL13	.A..... ..C..TT.... .G.AA..A.. C..G.....
DL16	.A..... ..C..TT.... .G.AA..A.. C..G.....
ST4	.G..... ..A..CA.... .C.AA..G.. C..A.....
R17	.G..... ..A..CA.... .C.AA..A.. C..A.....
J20	.A..... ..A..TT.... .A.AA..A.. C..G.....
MS2	.G..... ..A..CA.... .C.AA..G.. C..A.....

M12	.G.....A..TT....	.C.AA..G..	C..A.....
fr	.A.....T..CT....	.C.GG..T..	T..G.....

	27602770278027902800
Consensus	TTTSGTAACR CCGGAACCAT WGGCATCTAY GGGGACGATA TYATATGYCC
DL1	...G....G A.....CT....T..
DL2	...G....G A.....CT....T..
DL13	...G....G A.....CT....T..
DL16	...G....G A.....CT....T..
ST4	...G....G A.....CT....T..
R17	...G....G A.....CC....T..
J20	...G....G A.....CT....T..
MS2	...G....G A.....CT....T..
M12	...G....G A.....CT....T..
fr	...C....A T.....TT....C..

	28102820283028402850
Consensus	CASWGAGATT GCACCYCGYG TGCTRGARGC RCTDRSCTWC TACGGTTTTYA
DL1	..GT.....C..T.A..G.. A..TGC..A.T.
DL2	..GT.....C..T.A..G.. A..TGC..A.C.
DL13	..GT.....C..T.A..G.. A..TGC..A.C.
DL16	..GT.....C..T.A..G.. A..TGC..A.C.
ST4	..GT.....C..T.G..G.. A..TGC..A.C.
R17	..GT.....C..T.A..G.. A..TGC..A.T.
J20	..GT.....C..T.A..G.. G..AGC..A.C.
MS2	..GT.....C..T.A..G.. A..TGC..A.T.
M12	..GT.....C..T.A..G.. A..TGC..A.T.
fr	..CA.....T..C.G..A.. A..GAG..T.C.

	28602870288028902900
Consensus	AACCGAATCT HCGWAARACG TTCRYGTCVG GSYYTTCG CGAGAGCTGY
DL1 T..T..A... ...GT...G. .GCTC.....C
DL2 T..T..A... ...GT...A. .GCTC.....T
DL13 T..T..A... ...GT...A. .GCTC.....T
DL16 T..T..A... ...GT...A. .GCTC.....T
ST4 C..T..A... ...GT...C. .GCTC.....C
R17 T..T..A... ...GT...C. .GCTC.....C
J20 T..T..A... ...GT...A. .GCTC.....T
MS2 T..T..A... ...GT...C. .GCTC.....C
M12 T..T..A... ...GT...C. .GCTC.....C
fr A..A..G... ...AC...C. .CTCT.....C

	29102920293029402950
Consensus	RGCGCGCACT WTTWCCGTGG TGTCGATGTY AAACCRTTYT AYATCARGAA
DL1	G..... T..A.....CG..T. .C....A...
DL2	G..... T..A.....CG..C. .C....A...
DL13	G..... T..A.....CG..C. .C....A...
DL16	G..... T..A.....CG..C. .C....A...
ST4	G..... T..A.....CG..T. .C....A...
R17	A..... T..A.....CG..T. .C....A...
J20	G..... T..A.....CG..T. .C....G...
MS2	G..... T..A.....CG..T. .C....A...

M12	G..... T..A..... ..C.....G..C. .C....A...
fr	G..... A..T..... ..T.....A..T. .T....A...

	2960 2970 2980 2990 3000
Consensus	ACCWRTYRMY RAYCTMTTYK CCCTDATGCT KATMHTDAAY CGKMTWMGSG
DL1	...AG.TGAC A.C..C..CTT..... G..AA.G..T ..GC.TA.G.
DL2	...AG.TGAC A.T..C..CTT..... G..CA.G..T ..GC.TA.G.
DL13	...AG.TGAC A.T..C..CTT..... G..CA.G..T ..GC.TA.G.
DL16	...AG.TGAC A.T..C..CTT..... G..CA.G..T ..GC.TA.G.
ST4	...TG.TGAC A.T..C..CGT..... G..AT.G..T ..GC.AC.G.
R17	...TG.TGAC A.C..C..TGG..... G..AT.A..T ..GC.AC.G.
J20	...AG.CGAC A.T..C..CTT..... G..CA.G..T ..GC.TA.G.
MS2	...TG.TGAC A.T..C..CGG..... G..AT.A..T ..GC.AC.G.
M12	...AG.TGAC A.C..C..TTT..... G..AC.G..T ..GC.AC.G.
fr	...AA.CACT G.C..A..CTA..... T..AC.T..C ..TA.AC.C.

	3010 3020 3030 3040 3050
Consensus	GDTGGGGNGT WGTCTRRMGGW ATRKCAGAYC CDCGCCTHTA YRAGGTDGTGG
DL1	.G.....C.. T...GGA..T ..GT....T.. .G.....T.. TA....T...
DL2	.G.....C.. T...GGA..T ..GT....T.. .G.....T.. TA....T...
DL13	.G.....C.. T...GGA..T ..GT....T.. .G.....T.. TA....T...
DL16	.G.....C.. T...GGA..T ..GT....T.. .G.....T.. TA....T...
ST4	.T.....A.. T...GGA..T ..GT....T.. .A.....T.. CA....G...
R17	.T.....G.. T...GGA..T ..GT....T.. .A.....T.. CA....G...
J20	.A.....T.. T...GGA..T ..GT....T.. .T.....A.. TA....T...
MS2	.T.....A.. T...GGA..T ..GT....T.. .A.....C.. TA....G...
M12	.T.....G.. T...GGA..T ..GT....T.. .G.....T.. CA....T...
fr	.A.....G.. A...AAC..A ..AG....C.. .A.....C.. CG....A...

	3060 3070 3080 3090 3100
Consensus	GWRMRRCTVT CCWSMYWGGT DCCDWSRWK THTTTYGGTS SSACGGACCT
DL1	.TACGA..G. ..TCCCT... T..ATCGATG ..C..C...G GG.....
DL2	.TACGA..A. ..TCCCT... T..ATCGATG ..C..C...G GG.....
DL13	.TACGA..A. ..TCCCT... T..ATCGATG ..C..C...G GG.....
DL16	.TACGA..A. ..TCCCT... T..ATCGATG ..C..C...G GG.....
ST4	.TACGA..C. ..TCCCA... G..TTCGATG ..T..C...G GG.....
R17	.TACGA..C. ..TCCCA... A..TTCGATG ..C..C...G GG.....
J20	.TACGA..G. ..TCCCT... T..ATCGATG ..C..C...G GG.....
MS2	.TACGG..C. ..TCCCA... G..TTCGATG ..C..C...G GG.....
M12	.TACGA..G. ..TCCCT... T..ATCGATG ..C..C...C CC.....
fr	.AGAAG..C. ..AGATT... A..GAGATAT ..A..T...G GG.....

	3110 3120 3130 3140 3150
Consensus	BSMKGCYDAY TAYTACGTMG TBWSMCCBCC NAHNSYDRWM KSRRTWTAYW
DL1	TGCT..C..C ..T.....A. .CAGC..C.. G.CCGCAGTC TCAG.T..TA
DL2	TGCT..C..C ..C.....A. .TAGC..T.. G.ATGCTGTC TCGG.T..TA
DL13	TGCT..C..C ..C.....A. .TAGC..T.. G.ATGCTGTC TCGG.T..TA
DL16	TGCT..C..C ..C.....A. .TAGC..T.. G.ATGCTGTC TCGG.T..TA
ST4	CGCT..C..C ..C.....A. .CAGC..G.. C.CGGCAGTC TCGG.A..TA
R17	CGCT..C..C ..C.....A. .CAGC..G.. T.CGGCGGTC TCGG.A..TA
J20	TGCT..T..C ..C.....A. .CAGC..C.. G.CAGCGGTC TCAG.T..TA
MS2	CGCT..C..C ..C.....A. .CAGC..G.. T.CGGCAGTC TCGG.A..CA

M12 CGCT..T..C ..C.....A. .CAGC..T.. A.CTGCTGTC TCAG.A..TA
fr GCAG..C..T ..C.....C. .GTCA..T.. T.TTCTGAAA GGAA.T..TT

	31603170318031903200
Consensus	CYAARACYSM VTRBGRRMSG YKRSTAYGCS GAHRCYMGWA CMWCRGGYTT
DL1	.T..A..TGC C.AC.GGAG. TTAC.-C..G ..TA.CC.T. .CT.G..T..
DL2	.C..A..TGC A.AT.GGAG. TTAC.-C..G ..CG.CC.T. .CT.G..T..
DL13	.C..A..TGC A.AT.GGAG. TTAC.-C..G ..CG.CC.T. .CT.G..T..
DL16	.C..A..TGC A.AT.GGAG. TTAC.-C..G ..CG.CC.T. .CT.G..T..
ST4	.C..G..TCC G.AT.GGCG. CTAC.-C..G ..TA.CC.T. .CT.G..T..
R17	.C..G..TCC G.AT.GACG. CTGC.-C..G ..TA.CC.T. .CT.G..T..
J20	.T..G..CGC A.AT.GGAG. TTGC.-C..G ..CG.CC.T. .CT.G..T..
MS2	.C..G..TCC G.AC.GGCG. CTGC.-C..G ..TA.CC.T. .CT.G..T..
M12	.T..G..CGC G.AT.GGAG. CTGC.-C..G ..TA.CC.T. .CT.G..T..
fr	.C..G.-TGA A.GG.AGAC. CGAG..T..C ..AG.TA.A. .AA.A..C..

	32103220323032403250
Consensus	CMRKCTYGCT MGHAYCGCDM RRKRRCGHAA GYDCTTYWSC GADAAGCAYG
DL1	.CGT..T... C.T.T...AA AAGAG..A.. .CG...TAG. ..G.....T.
DL2	.CGT..T... C.T.T...GA AAGAG..A.. .CA...TAG. ..G.....T.
DL13	.CGT..T... C.T.T...GA AAGAG..A.. .CA...TAG. ..G.....T.
DL16	.CGT..T... C.T.C...GA AAGAG..A.. .CA...TAG. ..G.....T.
ST4	.CGT..T... C.T.T...TC GAGAA..C.. .TT...CAG. ..A.....T.
R17	.CGT..T... C.T.T...TC GAGAA..C.. .TT...CAG. ..A.....T.
J20	.CGT..T... C.T.T...GA AAGAG..T.. .CA...TAG. ..G.....T.
MS2	.CGT..T... C.T.T...TC GAGAA..C.. .TT...CAG. ..A.....C.
M12	.CGT..T... A.A.T...AA AAGAG..T.. .CG...TAG. ..G.....T.
fr	.AAG..C... C.C.T...GA GGTGG..A.. .CA...TTC. ..T.....C.

	32603270328032903300
Consensus	RCWSWGGYCG STACATWGCR TG GTTCCATA CTGGAGGTGA RRTCACCGAY
DL1	A.AGT..T.. C.....A..A GA.....T
DL2	A.AGT..T.. C.....A..A AA.....T
DL13	G.AGT..T.. C.....A..A AA.....T
DL16	A.AGT..T.. C.....A..A AA.....T
ST4	A.AGT..C.. C.....A..G AG.....C
R17	A.AGT..T.. C.....A..G AA.....C
J20	A.AGT..T.. C.....A..A AA.....T
MS2	A.AGT..T.. C.....A..G AA.....C
M12	A.AGT..T.. C.....A..G AA.....T
fr	A.TCA..T.. G.....T..G GA.....T

	33103320333033403350
Consensus	AGYATGAAGT CCGCYGGYGT RCGYRTHMTR CGYACTTCGG ARTGGCTRMM
DL1	..T..... ..T..C.. G..CG.AA.G ..C..... .G.....GAC
DL2	..T..... ..T..C.. G..TG.AA.A ..C..... .G.....GAC
DL13	..T..... ..T..C.. G..TG.AA.A ..C..... .G.....GAC
DL16	..T..... ..T..C.. G..TG.AA.A ..C..... .G.....GAC
ST4	..T..... ..C..C.. G..TA.TA.G ..C..... .G.....AAC
R17	..T..... ..C..C.. G..CA.CA.G ..C..... .G.....AAC
J20	..T..... ..C..C.. G..TG.TA.G ..C..... .A.....AAC
MS2	..C..... ..C..C.. G..CG.TA.A ..C..... .G.....AAC

M12	..T.....T..C..	G..CG.AC.G	..C.....	.G..-----
fr	..T.....C..T..	A..CG.TA.G	..T.....	.G.....ACA

STOP 4

	3360	3370	3380	3390	3400
Consensus	GCCGGTTCCC	RYRTTCCCKC	AGGARTGYGG	GCCAGCGAGC	TCTCCTCRGT	
DL1	ACA.....T.	...G..T..G..	
DL2	ATA.....T.	...A..T..G..	
DL13	ATA.....T.	...A..T..G..	
DL16	ATA.....T.	...A..T..G..	
ST4	ACA.....T.	...G..T..G..	
R17	ACA.....T.	...G..T..G..	
J20	ACA.....G.	...G..T..G..	
MS2	ACA.....T.	...G..T..G..	
M12	-----	-----	-----	-----	-----	
fr	GTG.....T.	...A..C..A..	

	3410	3420	3430	3440	3450
Consensus	AGCWRAACYG	AGGGACCCCC	GTAARCGGGG	TGGGTGTGCT	CGAAAGAGCA	
DL1	...TG.-.C.A.....	
DL2	...TG.-.C.A.....	
DL13	...TG.-.C.A.....	
DL16	...TG.-.C.A.....	
ST4	...TG.-.C.A.....	
R17	...TG.-.C.A.....	
J20	...TG.-.C.A.....	
MS2	...TG.-.C.A.....	
M12	-----	-----	-----	-----	-----	
fr	...AA...T.G.....	

	3460	3470	3480	3490	3500
Consensus	CGGGTCCGCG	WAAGCGGTCC	GGCTCMWCMG	AAATGGTGGG	YGAGGSTTCG	
DL1	A.....--	...CA.C.	...-...C.	C...C...	
DL2	A.....--	...CA.C.	...-...C.	C...C...	
DL13	A.....--	...CA.C.	...-...C.	C...C...	
DL16	A.....--	...CA.C.	...-...C.	C...C...	
ST4	A.....--	...CA.C.	...-...C.	C...C...	
R17	A.....--	...CA.C.	...-...C.	C...C...	
J20	T.....--	...CA.C.	...-...C.	C...C...	
MS2--	A.....	...CA.C.	...-...C.	C...C...	
M12	-----	-----	-----	-----	-----	
fr	A.....--	...AT.A.	T...G...	

	3510	3520	3530	3540	3550
Consensus	SCCTCWRGGA	CYKSCCCYKR	AAGRKMGRGC	CCGGGATTCT	CCCATTGTTG	
DL1	G...-AG...	.CTC...CTA	...AGAG.A.-	
DL2	G...-AG...	.CTC...CTA	...AGAG.A.-	
DL13	G...-AG...	.CTC...CTA	...AGAG.A.-	
DL16	G...-AG...	.CTC...CTA	...AGAG.A.-	
ST4	G...-AG...	.CTC...TTG	...AGAG.G.-	
R17	G...-AG...	.CTC...CTG	...AGAG.A.-	
J20	G...-AG...	.CTC...CTA	...AGAG.A.-	
MS2	G...-AG...	.CTC...CTA	...AGAG.A.-	
M12	-----	-----	-----	-----	-----	

fr

C....TA... .TGG...CGA ...GTCA.G.

	3560 3570 3580
Consensus	GTARCTAGYT GCTTGGCTAG YKACCACCCA
DL1	...A...C. TT.....
DL2	...A...C. TT.....
DL13	...A...C. TT.....
DL16	...A...C. TT.....
ST4	...A...C. TT.....
R17	...A...C. TG.....
J20	...A...C. TT.....
MS2	...A...C. TT.....
M12	-----
fr	...G...T. CT.....

Appendix A2
Group II

Alignment: Levivirus Group II.

	10 20 30 40 50
Consensus	GGKTGGBKVS SMHYYYDKKS SGSKTBCYBY YHHMCTTSMW GKCGMKMKRR
T72	-----
DL10	-----
DL20	----..GTGG GACCCCTTTC G.GG.C.TGC TCAA...CCT .T..AGCTAA
GA	-.G...TTAG GAATTTAGGG G.GG.T.CCT CACC...GAA .G..CTAGGA
KU1	..T...CGCC CCTTTCGGGG C.CT.G.TTT CTTC...GAA .G..CTAGGG

	60 70 80 90 100
Consensus	WKS BWTTKYW WWARYGDCTY WWKYSWGMMG TCKTTYAYC ATGCCGTTAG
T72	-----
DL10	-----
DL20	TGCCA..TTT TT.AT.T..T TAGCGA.AC. ----CT.C.
GA	ATGGT..GCA TA.GC.G..T AATTCT.CA. ..G..CC.C.
KU1	ATGTT..GCA AA.GC.A..C ATGCCT.CA. ..T..TC.T.
	ORF1

	110 120 130 140 150
Consensus	GTTTAGRTGA CGGTATRTTC CAYATACCGG AGGADCTATG TTTCCGAARY
T72G... ..A.-. ..T..... ..G..... ..AC
DL10A... ..A.-. ..T..... ..T..... ..GT
DL20A... ..A... ..T..... ..T..... ..GT
GAG... ..G.-. ..C..... ..T..... ..GT
KU1A... ..A.-. ..T..... ..A..... ..AC

	160 170 180 190 200
Consensus	MRAATATMGA YMGA AHYTAY MAKGTTAMAC TTRTHTCTTA CGAYRAKAAR
T72	AG....C.. TC...CC..C A.G....C.. ..A.T.... ...CA.G..G
DL10	CG....A.. CA...AT..C A.G....C.. ..A.A.... ...CG.G..A
DL20	CA....C.. CA...AT..C A.G....A.. ..A.T.... ...CA.G..G
GA	CA....A.. CA...AT..C A.G....A.. ..A.A.... ...CA.G..A
KU1	AG....C.. TC...TT..T C.T....A.. ..G.C.... ...TA.T..A

	210 220 230 240 250
Consensus	GGDAADSTHR YTCYGYGA YTCTTTYGAR YM DRYMGARA AYTAYCTCYT
T72	..T..TG.CA C...T..T.. T....C..A TCGACA..G. .C..T...C.
DL10	..G..GC.TG T...C..C.. C....C..G CAAGTC..A. .C..T...T.
DL20	..G..TC.TG T...C..C.. C....T..G CAAGTC..A. .C..T...T.
GA	..G..GC.TG T...C..C.. C....T..G CAAGTC..A. .C..T...T.
KU1	..A..AG.AA C...T..C.. C....T..G TCTGTC..G. .T..C...C.

	260 270 280 290 300
Consensus	YCAAAAYCGT TCGAMYWCGT ATAAACCYGG WTAYRTYCGW MRNGRHTTYC
T72	T.....T... ..ATT... ..T...T..A AAG.GA..C.
DL10	T.....T... ..ATA... ..C...T..A CGT.AC..C.
DL20	T.....C... ..ATA... ..T...T..T CGC.AC..T.
GA	T.....T... ..CTA... ..T...T..A CGT.AC..T.
KU1	C.....T... ..CCA... ..T...T..A AAA.AT..C.

	310 320 330 340 350
Consensus	GWARACCVAC DAACTTYTGG AAYGGCTWTC GCTRTTTCMA TCAGCCMGTY
T72	.T.A...G.. A....T... ..T...A.. ...A...A.C..C
DL10	.A.G...A.. G....T... ..T...T.. ...A...A.A..T
DL20	.A.G...A.. T....T... ..C...A.. ...G...A.A..T
GA	.A.G...A.. G....C... ..C...A.. ...G...A.A..T
KU1	.T.A...C.. G....T... ..T...A.. ...A...C.C..C

	360 370 380 390 400
Consensus	GGYRYSTTYA CTCGGAAACT HKMYRAYGGT GGGMGWCAAG WBGCYGATTA
T72	..CGTC..T.TGACA.C... ..A.A.... TC..T....
DL10	..TACC..C.CTCTG.T... ..A.A.... TT..T....
DL20	..TACC..C.CTCTG.T... ..A.A.... TT..T....
GA	..TACC..C.CTCTG.T... ..A.A.... TC..T....
KU1	..CACG..T.ATCTG.T... ..C.T.... AG..C....

	410 420 430 440 450
Consensus	YGGYATHGTR AACCCTAATA AGTTYACBGS YAAYAGYCAG CAYYTKGGRG
T72	C..T..C..GC..T.C T..T..T... ..CT.G..A.
DL10	T..C..C..GC..G.G C..C..C... ..CT.G..G.
DL20	T..C..T..GC..T.C C..C..T... ..CT.G..G.
GA	T..T..C..AT..T.C C..T..C... ..CT.G..G.
KU1	T..T..A..GC..C.C C..C..T... ..TC.T..A.

	460 470 480 490 500
Consensus	ABAACATGGT DATTTAYCCW GGTCCYTTYT CDATHAATAT TGAYMABMGW
T72	.G.....G.....T..AT..C. .G..C..... ...TA.TA.A
DL10	.C.....T....C..TC..T. .T..T..... ...TC.CC.A
DL20	.C.....A....C..TC..C. .T..A..... ...TC.GC.T
GA	.T.....A....C..TC..T. .T..A..... ...TC.GC.T
KU1	.G.....T....T..AT..C. .A..A..... ...CA.CA.A

	510 520 530 540 550
Consensus	GCTWSYGTYG AAGTCCTYAA YAARCTYTCD CARTCNAACC TCAATATTGG
T72	...AGT..C.T...T..A T..A..T..T ..A..T.....
DL10	...TCT..C.C...T..G T..A..T..G ..A..A.....
DL20	...TCT..C.C...T..A T..G..T..A ..G..G.....

GA	...TCC..T.C.. C..G..C..G ..A..T....
KU1	...TCT..C.C.. T..A..T..T ..A..C....

	560 570 580 590 600
Consensus	GGTTGCDATT GCDGARGCYA ARATGACTGC TTCRYTRCTY KCWMRDCAAT
T72G... ..A..A..T. .A..... ..AC.G..T T.TCGT....
DL10A... ..T..A..T. .G..... ..AT.A..C G.TAAA....
DL20G... ..T..G..T. .G..... ..AT.A..C G.TAAG....
GAA... ..T..A..C. .G..... ..GT.A..C G.TAAA....
KU1T... ..G..A..C. .G..... ..AT.A..C T.ACGT....

	610 620 630 640 650
Consensus	CDATHKCKCT CATCMGAGCC TACACTGCTG CAAAGCGCGG TAADTGGCGR
T72	.G..CT.T..C..... ..G.....G
DL10	.A..AG.T..A..... ..A.....A
DL20	.T..AG.T..A..... ..T.....A
GA	.T..AG.T..A..... ..T.....A
KU1	.T..TG.G..A..... ..G.....A

	660 670 680 690 700
Consensus	GAGGTGYTWT CWCARCTCCT YATYKCCGAA CACCGTTTCA SRRSWCCYKY
T72T.A. .A..A..... T..CG..... ..GAGCT..CTC
DL10C.T. .T..G..... C..CT..... GAGCT..TGT
DL20T.T. .T..A..... C..CT..... GAGCT..TGC
GAC.T. .T..G..... C..CT..... GAGCT..TGC
KU1C.T. .A..G..... C..TG..... CGAGA..CTC

	710 720 730 740 750
Consensus	WARGGATCTC GGAGGTCGAT GGCTCGAACT GCAGTACGGY TGGYTWCYCYC
T72	A.G..... ..T...T.A..T.
DL10	T.A..... ..T...T.A..T.
DL20	T.A..... ..T...T.A..C.
GA	T.A..... ..C...C.A..C.
KU1	T.A..... ..T...C.T..T.

	760 770 780 790 800
Consensus	TTATGAGYGA TWTVAARGCT GSHTATGAYY TGCTYACGCA RACYMAWYTR
T72T.. .A.C..G... .GC.....TCT..... G..TA.AT.A
DL10C.. .A.G..A... .CA.....TTT..... A..CA.AC.G
DL20T.. .T.G..A... .CA.....TTC..... A..CA.AC.G
GAT.. .T.G..A... .CA.....TTT..... A..CA.AT.G
KU1T.. .A.A..A... .GT.....CTT..... A..TC.TT.A

	810 820 830 840 850
Consensus	CCTGCKTTHA TGCCYYTDMG RGTWASYCGY ACCGTTGGCG SHACRCAYAA
T72T..A.TC.TA. A..T.CC..C ..GC..A..C..
DL10T..C.CT.GA. G..T.CC..T ..GA..G..C..
DL20T..C.CT.GA. G..T.CC..C ..GA..G..C..

GAT..C.CT.AA. G..T.CC..C	GA..G..C..
KU1G..T.TC.TC. A..A.GT..T	CT..G..T..

	860 870 880 890 900
Consensus	TTAYAAAGTG CGYAAAGTCG AATCTGCAGG RGATACTTGG TCMTATAGSS
T72	...T..... ..T..C.... ..A..... ..A.....CG
DL10	...T..... ..C..C.... ..G..... ..C.....CG
DL20	...T..... ..C..C.... ..G..... ..C.....CG
GA	...C..... ..C..C.... ..G..... ..C.....GC
KU1	...C..... ..T..T.... ..G..... ..C.....CG

	910 920 930 940 950
Consensus	AYCGBYTSTC RGTRAATTAC CGAATATGGT ATTTYATYTC YGAYCCVCGV
T72	.T..GC.C.. A..G..... ..C..C.. T..T..C..A
DL10	.T..TT.G.. G..G..... ..C..T.. C..T..A..G
DL20	.T..TT.G.. G..G..... ..C..T.. C..C..G..A
GA	.C..CT.G.. G..G..... ..C..T.. C..C..G..C
KU1	.C..GC.C.. A..A..... ..T..T.. T..T..G..A

	960 970 980 990 1000
Consensus	CTCGCATGGG CHAGTTCYCT HGGKCTYYTW AACCCWYTRG AAATYTAYTG
T72A.....T.. A..T..CT.AAT.A.T..T..
DL10T.....C.. T..G..TC.ATC.A.C..T..
DL20T.....C.. T..G..TC.TTT.G.C..C..
GAT.....C.. C..G..TC.TTT.A.C..T..
KU1C.....C.. A..T..CT.ATC.A.C..T..

	1010 1020 1030 1040 1050
Consensus	GGARAAGACR CCBTGGTCKT TCGTYGTTGA CTGGTTYTR CCYGTHGGWA
T72	...A....A ..G....G.T.....TC.G ..C..A..A.
DL10	...G....G ..T....T.C.....CT.A ..T..T..T.
DL20	...A....G ..C....G.T.....CT.A ..T..C..T.
GA	...G....G ..C....G.T.....CT.A ..T..C..T.
KU1	...A....A ..G....G.T.....TC.G ..T..C..A.

	1060 1070 1080 1090 1100
Consensus	ATCTKATMGA AGCYATGAGY AAYCCKCTYG GCYTMGAYAT HATTTTCYGGS
T72G..C.. ...C.....T ..C..T..T. ..T.A..C.. C.....T..G
DL10T..A.. ...C.....T ..T..T..T. ..C.C..T.. T.....C..C
DL20T..A.. ...C.....T ..T..T..T. ..C.C..C.. T.....C..C
GAT..A.. ...C.....T ..T..T..T. ..C.C..T.. T.....C..C
KU1T..C.. ...T.....C ..T..G..C. ..C.A..T.. A.....C..G

	1110 1120 1130 1140 1150
Consensus	ACDAARACYT GGCAACTYGA ATCWAARHTK AAYGCRWCSM TTMMVGCDBM
T72	..A..G..C.C... ..T..AT.G ..C..AA.CA ..AAC..ATC
DL10	..G..G..C.C... ..A..AC.T ..T..GA.GC ..ACA..TTC
DL20	..G..G..C.T... ..A..AC.T ..C..GA.GC ..CCG..TCC

GA	..G..G..T.C.. ...A..AC.T ..T..GA.GC ..CCG..TTC
KU1	..T..A..C.C.. ...T..GA.G ..T..GT.CC ..AAA..GGA

	1160 1170 1180 1190 1200
Consensus	DGGHTGGKYY GGRACWGCAA AGYTRWCTGC ATAYGCGAAW RMVKRYGACA
T72	A..C...TCT ..G..A.... ..T.AT.... ...T.....A GCGTAT....
DL10	G..T...TCT ..A..T.... ..T.AT.... ...C.....A GCATAT....
DL20	G..T...TCT ..A..T.... ..T.GA.... ...T.....A GCGTAT....
GA	G..T...TCT ..A..T.... ..T.GA.... ...C.....A GCATAT....
KU1	T..A...GTC ..A..T.... ..C.AT.... ...C.....T AACGGC....

	1210 1220 1230 1240 1250
Consensus	GRTCDACTTT CTAYTCCTTY CCHACBCKKH TGCCKTAYGT GAAATCYCCA
T72	.G..T..... ..T.....T ..A..G..TAT..C..C...
DL10	.G..A..... ..C.....T ..A..T..TAT..T..C...
DL20	.A..G..... ..T.....C ..C..C..GTG..C..T...
GA	.A..G..... ..T.....T ..C..T..TTG..C..C...
KU1	.A..T..... ..T.....T ..T..T..TCT..C..T...

	1260 1270 1280 1290 1300
Consensus	CTWAGTGGRC TTCACHTRGC VAAYGCRYTM GCMYTAATYA ACCAACGCCT
T72	..T.....G.A.G.. C..T..GC.C ..AT....T.
DL10	..T.....G.T.A.. G..T..AT.A ..CT....C.
DL20	..T.....G.C.A.. G..T..GT.A ..CT....C.
GA	..T.....G.T.A.. G..T..AT.A ..CT....C.
KU1	..A.....A.A.G.. A..C..AC.C ..CC....T.

STOP 1

ORF2

	1310 1320 1330 1340 1350
Consensus	GAAAAGGTAA WWAYGGAGTT AGCCAYATGG CAACTTTACG YAGTTTCGTA
T72 AT-C..... ..C.... T.....
DL10 TA.T..... ..T.... C.....
DL20 TT.C..... ..T.... C.....
GA TT.C..... ..T.... C.....
KU1 AT-C..... ..C.... T.....

	1360 1370 1380 1390 1400
Consensus	CTCGTCGATA ATGGCGGTAC GGGGAATGTT ACTGTCGTTC CTGTTAGCAA
T72
DL10
DL20
GA
KU1

	1410 1420 1430 1440 1450
Consensus	TGCCAACGGC GTCGCTGAGT GGCTTTCTAA TAACTCDCGC AGTCARGCTT
T72G.... ..A....
DL10G.... ..A....
DL20A.... ..A....
GAG.... ..G....
KU1T.... ..A....

	14601470148014901500
Consensus	ATCGCGTGAC TGCMAGTTAT CGTGCGTCAG GYGCSGAYAA GCGCAARTAY
T72C.....C..G..C..A..C
DL10C.....C..G..C..G..C
DL20C.....C..G..C..A..C
GAC.....C..G..C..A..T
KU1A.....T..C..T..A..C

	15101520153015401550
Consensus	RCYATYAARC TKGARGTRCC GAARATCGTY ACYCARRBHG TWAAYGGHGT
T72	A..C..C..G..G..G..A..A....T..C..GACA..T..T..T..
DL10	A..C..C..A..T..A..A..A....T..T..AGTT..T..C..T..
DL20	A..C..T..A..T..A..A..A....T..T..AGTT..A..T..A..
GA	G..C..T..A..T..A..A..A....T..C..AGTT..A..T..T..
KU1	A..T..T..G..G..A..G..G....C..T..AAGC..T..T..C..

	15601570158015901600
Consensus	HGARYTGCCT GKTTCSGCRT GGAAGGCTTW YGCCTCYATY GACCTGACCA
T72	T..GC.....T...C..A.....A C.....C..T
DL10	C..GC.....T...C..A.....A T.....T..T
DL20	T..GC.....T...C..A.....A C.....T..C
GA	T..GC.....G...C..A.....A T.....T..C
KU1	A..AT.....T...G..G.....T C.....C..T

	16101620163016401650
Consensus	TCCCKATCTT TGCDGCDACY GAYGAYGTGA CYSTTATTTT CAAGTCRCTC
T72	...T.....G..G..C..C..C....TC.....A...
DL10	...G.....T..A..T..C..T....TG.....A...
DL20	...T.....T..A..C..T..T....CG.....A...
GA	...T.....T..A..C..C..C....TG.....G...
KU1	...T.....A..T..C..C..C....TC.....A...

ORF3-T72,KU1

	16601670168016901700
Consensus	GCYGGYCTST TCAARRTTGG GRACCCKRTY GCYGAHGCYA TCTCTTCDCA
T72	..T..T..C.....GA....A....GG.T..T..T..C.....A..
DL10	..T..T..G.....AG....A....TA.C..T..A..T.....G..
DL20	..T..C..G.....AG....G....TA.C..C..C..T.....T..
GA	..C..C..G.....AG....A....TA.C..T..A..T.....A..
KU1	..T..T..C.....GA....A....GG.T..T..T..C.....A..

STOP 2

ORF3-DL10,DL20,GA

	17101720173017401750
Consensus	GAGYGGSTTC TACGCSAAG GCGTAGTTCT TCAGCATTTGG GTCWGAAAGC
T72	...T..G...C.....A.....
DL10	...C..C...G...- - - - -T.....
DL20	...T..C...G...- - - - -T.....
GA	...T..C...G...- - - - -T.....
KU1	...T..G...C.....A.....

ORF4

	1760 1770 1780 1790 1800
Consensus	AWTTAAACAT AAGGAAAACC TATGTTCCGA TTCASAGAGA TCRAAAAGAC
T72	.A..... .C..... .A.....
DL10	.---..... .C..... .G.....
DL20	.---..... .C..... .G.....
GA	.---..... .G..... .G.....
KU1	.T..... .C..... .G.....

	1810 1820 1830 1840 1850
Consensus	TCTATGTATG GATCGCACTC GCGATTGTGC TGTCCGATTT CACGTCTATC
T72
DL10
DL20
GA
KU1

	1860 1870 1880 1890 1900
Consensus	TTCAGTCATT GGATWTGGGG TCTTCTGATC CTCWWTCTCC RGACTTTGAT
T72T..... .AT..... A.....
DL10A..... .AT..... A.....
DL20T..... .AT..... G.....
GAT..... .TA..... A.....
KU1T..... .AT..... G.....

STOP 3

	1910 1920 1930 1940 1950
Consensus	GGACTTGCCT ACCTTCGTA YGARTGTYTA ACTAAACATC CBTCACTHGG
T72 C..A...T.. .T....T..
DL10 T..G...T.. .T....A..
DL20 T..A...T.. .C....C..
GA T..A...C.. .T....T..
KU1 T..A...T.. .G....T..

	1960 1970 1980 1990 2000
Consensus	RRACAGTAAT TCGGAYGCAC KCCGTAAGGA AYTGGCATAT GCCAAACTTA
T72	AG..... .T.... T..... .T.....
DL10	AA..... .T.... G..... .T.....
DL20	AG..... .C.... G..... .T.....
GA	AG..... .C.... G..... .T.....
KU1	GG..... .C.... T..... .C.....

	2010 2020 2030 2040 2050
Consensus	TGGATAGTGA TCAAAGATGC AAAATCCAAA ACAGTAACGG ATACGACYWV
T72CTG
DL10TAC
DL20TAC
GATAC

	2060 2070 2080 2090 2100
Consensus	WSTCAYATCG ANDSWRGC GT ACTKARYGGH ATAYTMMWRA CYGCMAGGC
T72	AG...T.... .CTCTG.... ...T.AC..C ...T.ACTA. .C..TA....
DL10	TC...C.... .AAGTA.... ...G.GT..T ...C.CAAG. .T..TC....
DL20	TC...T.... .GAGTG.... ...G.GT..T ...C.CAAG. .T..TC....
GA	TC...T.... .GAGTG.... ...T.GC..T ...C.CAAG. .C..CC....
KU1	AG...T.... .TGCAG.... ...T.AC..A ...T.ACTA. .C..TA....

	2110 2120 2130 2140 2150
Consensus	YYYWRTRGCR AASTTRCTTA YRGGYTTTGA ATCTCACTTC CTGAACGATT
T72	TTCAA.A..G ..G..A.... TG..C.....
DL10	CCTTG.G..G ..C..G.... CG..T.....
DL20	CCTTG.G..A ..C..G.... CG..T.....
GA	CCTTG.G..A ..C..A.... CG..T.....
KU1	TTTAA.A..G ..G..A.... TA..T.....

	2160 2170 2180 2190 2200
Consensus	GTTTCRTTCTC CAAYGGAGCC TCACAAGGGT TCAAGTTGCR GGATGCAGCG
T72A..... ..T.....
DL10A..... ..T.....
DL20G..... ..C.....
GAA..... ..C.....
KU1A..... ..T.....

	2210 2220 2230 2240 2250
Consensus	CCGTTTAAAGA AGATCGCTGG GCAAGCAACC GTTACAGCTC CTGTCYTATRA
T72
DL10
DL20
GA
KU1

	2260 2270 2280 2290 2300
Consensus	YHTWGCGGTG SHCGCMGTTA AAACKTGCGS RCCRTGGYWT SSTTATATGC
T72	CC.A..... CA...A.... ...T....G A..A...CT. CG.....
DL10	CA.T..... GC...C.... ...G....C G..G...TA. GC.....
DL20	CA.T..... GC...C.... ...G....C A..G...TA. GC.....
GA	CA.T..... GC...C.... ...G....C A..G...TA. GC.....
KU1	TT.A..... CT...A.... ...T....G A..A...CT. CG.....

	2310 2320 2330 2340 2350
Consensus	AGGAAACGTA TGGCGATGAR ACCARATGGT TTCGTCGGGT RTACGGMAAC
T72
DL10
DL20

GAGA.....A.....C...
KU1GG.....G.....C...

	2360 2370 2380 2390 2400
Consensus	GGTTTGTGTTT CTGTTCCGAA GAACAATAAA ATAGATCGGG CTGCCTGYAA
T72T..
DL10T..
DL20T..
GAT..
KU1C..

	2410 2420 2430 2440 2450
Consensus	GGAGCCTGAT ATGAATATGT AYCTWCAGAA RGGKGCKGGM TCTTTTATWA
T72T..
DL10A..
DL20A..
GAA..
KU1A..

	2460 2470 2480 2490 2500
Consensus	GAMRMCGMCT YCGMTCYGTB RRYATWGATC TTAAYGATCA RACRYGCAAY
T72	..CGC..A.. C..A..T..G AAC..T.... ..C..... A..AC....T
DL10	..AAA..C.. T..C..C..C GGT..A.... ..C..... A..GT....T
DL20	..AAA..C.. T..C..C..T GGT..A.... ..T..... A..GC....C
GA	..AAA..C.. T..C..C..C GGT..A.... ..C..... G..GC....T
KU1	..CGC..A.. C..A..T..G AAC..T.... ..C..... G..AC....T

	2510 2520 2530 2540 2550
Consensus	CAGGARTTRG CCCGACTTGG CAGCATTGAT GGKTCRCTYG CTACTATWGA
T72A..G.T..G..T.T..
DL10A..A.T..A..C.T..
DL20A..A.G..G..C.T..
GAA..A.T..G..C.T..
KU1G..A.T..G..T.A..

	2560 2570 2580 2590 2600
Consensus	TYTWAGTAGY GCTAGYGAYT CYRTCTCTGA YCGYCTYGTM TGGGATYTAC
T72	.T.A....TC..T. .CG..... T..C..C..CT...
DL10	.C.T....CT..T. .TG..... C..C..T..CT...
DL20	.C.T....CT..C. .CA..... C..T..C..CT...
GA	.C.T....CC..T. .CA..... C..T..T..CC...
KU1	.T.A....CC..T. .CG..... C..C..C..AT...

	2610 2620 2630 2640 2650
Consensus	TTCCRCCRCA YGTYTATTCA TAYCTCSMYC GYATYCGHWC RTCKTTTCAW
T72G..G.. T..T..... ..T...CAC. .T..C..CT. A..T.....T
DL10G..A.. C..T..... ..C...GCT. .C..C..TT. A..G.....T
DL20A..G.. C..C..... ..C...GCT. .T..T..CT. G..T.....T

GAG..G.. C..T..... ..C...GCT. .T..C..AA. A..G.....T
KU1G..G.. T..T..... ..T...CAC. .C..T..CT. A..T.....A

	2660 2670 2680 2690 2700
Consensus	ATGATYGAYG GDCRBYTRCA YAAGTGRRY CTRTTTTCTA CBATGGGWAA
T72C..T. .G.AGT.A.. T.....AAC ..A..... .G.....T..
DL10C..T. .T.GGT.G.. C.....GGC ..G..... .T.....A..
DL20T..C. .G.GCT.A.. C.....GGT ..A..... .C.....T..
GAC..T. .G.GTT.A.. T.....GGT ..A..... .C.....T..
KU1T..T. .A.GTC.G.. T.....AAC ..A..... .G.....T..

	2710 2720 2730 2740 2750
Consensus	YGGYTTYACR TTCGARCTCG AGTCCATGAT HTTYTGGGCY YTDAGYAASA
T72	C..T..T..AG.... A..T.....T C.G..T..G.
DL10	C..C..T..AA.... T..C.....T T.G..T..G.
DL20	C..C..T..AA.... C..T.....C C.G..T..G.
GA	T..C..C..GA.... C..T.....T T.A..C..G.
KU1	C..T..T..GA.... A..T.....T C.T..C..C.

	2760 2770 2780 2790 2800
Consensus	SYRTWATGYG GTMCMTSGGT GTTACTGGCT YAYTWGGCRT CTAYGGDGAY
T72	GTG.A...TC ..A.C.C... T.C.T...A. ...T..A..C
DL10	GTG.T...CT ..C.A.G... C.T.A...A. ...C..T..T
DL20	GCG.T...CT ..C.A.G... C.T.A...G. ...C..T..C
GA	GCA.T...CT ..C.A.G... C.T.A...A. ...C..T..T
KU1	CTG.T...TC ..A.C.C... T.C.T...A. ...T..G..C

	2810 2820 2830 2840 2850
Consensus	GATATAATCG TYCCYRYKRA RTGYSSHCCV MYYCTYYTWM AGGTWYTATC
T72C..CACGA. A..CGCA..C CTT..TC.ACAT....
DL10C..CGTTG. G..TGCC..A ACT..CC.TAAC....
DL20T..TGTTG. A..TGCC..G ACC..CC.TAAT....
GAC..CGTTG. G..TCGT..A ACT..CC.TAAC....
KU1T..CACGA. A..CGCA..C CTT..CT.ACTT....

	2860 2870 2880 2890 2900
Consensus	SGCYGTRAAY TTTCTTCCKA AYWAGRAGAA RACRTTYACD ACSGGKTAYT
T72	G..T..G..CT. .CC..A.... G..G..T..T ..C..T..T.
DL10	C..T..G..CT. .TA..A.... G..G..C..T ..G..T..C.
DL20	C..T..G..CG. .TC..A.... G..G..T..G ..G..T..C.
GA	C..T..A..CT. .TG..G.... A..A..T..A ..G..T..C.
KU1	G..C..A..TT. .CC..A.... A..A..C..G ..C..G..T.

	2910 2920 2930 2940 2950
Consensus	TYCGTGARAG TTGYGGKGCH CAYTTCTTYA AAGRYGICYM MRTRAAACCT
T72	.C.....G.. ...T..G..A ..T.....T. ...GT..TTC AG.A.....
DL10	.T.....A.. ...T..T..T ..C.....T. ...GT..CGA CA.G.....
DL20	.T.....G.. ...C..T..T ..T.....T. ...GC..CGA CA.G.....

GA .T.....A.. ...T..T..C ..C.....C. ...AT..CGA CA.G.....
KU1 .C.....G.. ...T..G..A ..T.....C. ...GT..TTC AG.A.....

	2960 2970 2980 2990 3000
Consensus	TTTTACTGCA AGCGGCCRAT GGAAACCCTT CCSGAYRTHM TRTTRCTMTG
T72G.. ..C..TA.TA .G..A..C..
DL10A.. ..C..TG.CA .G..G..A..
DL20A.. ..C..TG.CA .G..G..C..
GAA.. ..C..TG.CA .G..G..A..
KU1G.. ..G..CG.AC .A..A..A..

	3010 3020 3030 3040 3050
Consensus	YAAYAGRATH MGDGGTTGGS RKACYRTYGG KGGAATRTCA GAYCCKCGRC
T72	T..C..A..T C.T.....G GT..TA.C.. G.....A... ..C..T..A..
DL10	C..C..G..A A.G.....C AG..CG.T.. T.....G... ..T..G..G..
DL20	C..C..G..A A.G.....C AG..CG.T.. T.....G... ..T..G..G..
GA	C..C..G..A A.A.....C AG..CG.T.. T.....G... ..T..G..A..
KU1	T..T..G..C C.G.....G GG..TA.C.. G.....A... ..C..T..G..

	3060 3070 3080 3090 3100
Consensus	TCTTTCCDAT WTGGAARGAG TTYGCWGATA TGATHCCGCC KAAGTTTAAG
T72A.. A.....G... ..T..A.... ..A..... T.....
DL10T.. T.....G... ..T..T.... ..T..... G.....
DL20G.. T.....G... ..T..T.... ..T..... G.....
GAT.. T.....G... ..C..T.... ..C..... G.....
KU1G.. A.....A... ..T..A.... ..A..... T.....

	3110 3120 3130 3140 3150
Consensus	GGTGGATGYA ACCTKGATCG CGAYACTTAC CTYGTYTAC CKGATAAACC
T72T..T..... ...C..... ..T..T.... .G.....
DL10C..G..... ...T..... ..C..T.... .G.....
DL20T..G..... ...T..... ..C..T.... .T.....
GAC..G..... ...T..... ..C..C.... .T.....
KU1T..G..... ...C..... ..T..C.... .G.....

	3160 3170 3180 3190 3200
Consensus	TGGTRWHWCA YTRGTTTCWR WYGCKAMGRW RCGHWSTGGT TTTAACYAYD
T72AAAA.. T.A.....TG TT..G.A.AA A..AAG....C.TA
DL10GTCA.. T.G.....AG TC..T.A.GT A..TTC....C.TT
DL20GTCA.. C.G.....TA TT..T.A.GT A..CTC....C.CG
GAGTTT.. T.G.....TA TT..T.A.GT A..CTC....C.TG
KU1GTAA.. T.A.....TG AT..G.C.GT G..TAG....T.CA

	3210 3220 3230 3240 3250
Consensus	MRTTYCSKWR BSRCYAWGAA AAYGGYCGYT AYRTYCAITG GYTRCATATG
T72	AG..T.GTAG TGA.T.T... ..T..T..C. .TA.T..T.. .T.A.....
DL10	CG..C.CGTA CGG.C.T... ..T..T..C. .CG.C..C.. .T.G.....
DL20	CG..C.CGTA CGG.T.T... ..C..C..T. .CG.C..T.. .C.A.....

GA	CG..C.CGTA	CGG.C.T...	..T..T..C.	.CG.C..C..	.T.G.....
KU1	AA..C.GTAG	GCG.C.A...	..T..C..C.	.TA.C..T..	.T.A.....

	3260 3270 3280 3290 3300
Consensus	GGATCGGGHG AAGTDYYYGA RACYATWTCY TCCGCCAGGT WTAGGTGCAA
T72C.GCTC.. A..T..A..T T.....
DL10A.TCTC.. G..C..A..C T.....
DL20A.TCTC.. G..C..A..C T.....
GAA.TCTT.. G..C..T..C A.....
KU1T.ATCC.. A..T..A..T T.....
	STOP 4

	3310 3320 3330 3340 3350
Consensus	ACCTAAYTCG GAATGGAGRA YACAGATCCC KCTATTTTCCT CAGGAADTAG
T72T... ..G. C..... G.....A...
DL10C... ..A. C..... T.....T...
DL20C... ..A. C..... T.....T...
GAC... ..A. C..... T.....T...
KU1C... ..A. T..... T.....G...

	3360 3370 3380 3390 3400
Consensus	AGGCCTGCGT TCTCTCCTGA TAGTATCAGG ACCTCCCCGK AYGGGGTGGG
T72T.....
DL10T.....
DL20C.....
GAT.....
KU1T.....

	3410 3420 3430 3440 3450
Consensus	TGKGACCGWA AGGYMCTAT GGRGGTGAWH YCYCCACCG CCBWWAWTGG
T72	..G....A. ...TTC.... ..A....TA T.CT.--... ..GTA.--...
DL10	..T....A. ...CCA.... ..G....AT C.CC.--... ..CAA.A-...
DL20	..T....A. ...CCA.... ..A....AC C.TC.--... ..CAA.A-...
GA	..T....A. ...CCA.... ..A....AC C.TC..... ..CAA.A-...
KU1	..G....T. ...TTC.... ..A....TA C.CT.--... ..TTT.T...

	3460 3470 3480 3490
Consensus	CGGTTYWMGG TGACTAGTTT GCTTGGCTAG TCACCACCCA
T72	..-...TAA.. ..
DL10TTA.. ..
DL20CTC.. ..
GACTC.. ..
KU1TAA.. ..

Appendix A3a
Group III

Alignment: Allolevivivirus Group III.

	10 20 30 40 50
Consensus	GGGKKHCCCC CCKTAGGGGG KYWCYYYAYR YAGYAGTAYT TCAMYVAKTA
TW18	...GGA.... ..T..... TCA.CTC.CA C..C....C. ...CTA.G..
HL4-9	...GGA.... ..T..... TCA.CTC.CA C..C....T. ...CTG.G..
BR12	...GGA.... ..T..... TCA.CTC.CA C..C....T. ...CTG.G..
BZ1	...GGA.... ..T..... TCT.CTC.CA C..C....T. ...CTG.G..
VK	...GGA.... ..T..... TCA.CTC.CA C..C....T. ...CTG.G..
QB	-----
M11	...TTC.... ..G..... GTA.TCT.TG T..T.---- -..ACC.T..
MX1	-.GTT.... ..G..... GTA.TCT.TG T..T.---- -..ACC.T..
	ORF1

	60 70 80 90 100
Consensus	YDRGAGGMMA YATGCCWMRW YTACCDMGKG SWCTKCGWTT CGGASCSRAY
TW18	TGA...AC. T.....TAAA T....AC.T. GT..G..T..G.CG.T
HL4-9	TGA...AC. T.....TAAA T....AC.T. GT..G..T..G.CG.T
BR12	TAA...AC. T.....TAAA T....AC.T. GT..G..T..G.CG.T
BZ1	TAA...AC. T.....TAAA T....GC.T. GT..G..T..G.CG.T
VK	TGA...AC. T.....TAAA T....AC.T. GT..G..T..G.CG.T
QB	TAA...AC. T.....TAAA T....GC.T. GT..G..T..G.CG.T
M11	CTG...CA. C.....ACGT C....TA.G. GA..T..A..G.GA.C
MX1	CTG...CA. C.....ACGT T....TA.G. CA..T..T..C.GA.T

	110 120 130 140 150
Consensus	AWKGARRTYY TWARYGAYTT YCARGARCTC TGGTWTCCAG ABYYCHKYRT
TW18	.AT..AA.TC .T.AT..T.. T..G..G...T..... .CCT.TTTA.
HL4-9	.AT..AA.TC .T.AT..T.. T..A..G...T..... .TCT.TTTA.
BR12	.AT..AA.TC .T.AT..T.. T..G..G...T..... .TCT.TTTA.
BZ1	.AT..AA.TC .T.AT..T.. T..G..G...T..... .TCT.TTTA.
VK	.AT..AA.TC .T.AT..T.. T..G..G...T..... .TCT.TTTA.
QB	.AT..AA.TC .T.AT..T.. T..G..G...T..... .CCT.TTTA.
M11	.TG..GG.CT .A.AT..C.. C..G..A...A..... .GTC.CGCG.
MX1	.TG..GG.CT .A.GC..C.. C..G..A...A..... .GTC.ATTA.

	160 170 180 190 200
Consensus	CGAWTCYKMY RHSAHDYWYC CKTKGTAYAC MYTSARRGGT MRYRTSKKKR
TW18	...A..TTCC GAC.CGCAT. .G.G...C.. AC.G.AA... CGTG.GTTGA
HL4-9	...A..TTCC GAC.CGCAT. .G.G...C.. AC.G.AA... CGTG.GTTGA
BR12	...A..TTCC GAC.CGCAT. .G.G...C.. AC.G.AA... CGTG.GTTGA
BZ1	...A..TTCC GAC.CGCAC. .G.G...C.. AC.G.AA... CGTG.GTTGA
VK	...A..TTCC GAC.CGCAT. .G.G...C.. AC.G.AG... CGTG.GTTGA
QB	...A..TTCC GAC.CGCAT. .G.G...C.. AC.G.AG... CGTG.GTTGA
M11	...T..CGAT ACG.TTTTC. .T.T...T.. CT.C.AA... AACA.GGGTG
MX1	...T..CGAT GTG.AATAC. .T.T...T.. CT.C.GA... AGCA.CGGTG

	210 220 230 240 250
Consensus	RHKCYWYHT NGAYDMMYRY BKMMCNAAYR WWRKYGKYCG YSARRTHMGD
TW18	ACG.TCATC. T..TGATCGC CTAC.T..TG TAGGT.GT.. CC.GG.CA.G
HL4-9	ATG.CCATC. T..TGATCGT CTAC.T..TG TAGGC.GT.. TC.GA.CA.G
BR12	ACG.TCACA. G..TGACCGC TTAC.T..TG TAAGT.GT.. CC.AG.AC.G
BZ1	ACG.TCATT. A..TGATCGT CTAC.T..TG TAGGC.GT.. TC.AA.AA.G
VK	ACG.TCATA. T..TGACCGT TTAC.C..TG TAAGC.GT.. CC.AG.AA.G
QB	ACG.CCACC. T..TGATCGT CTAC.T..TG TAGGC.GT.. CC.GG.AA.G
M11	GCT.TTTTT. C..CACTTAC GGCA.A..CA ATATT.TC.. TC.GG.TC.A
MX1	GAT.TTCT. T..TTCTTAC GGCA.G..TA ATATC.TC.. CG.GA.TC.T

	260 270 280 290 300
Consensus	MGNACDCCDC AYYGYGYIAC HGTHCCKATW GCCWSTWCWG GCYTWMGKCC
TW18	C.T..T..A. .TC.T.TT.. T..T..G..T ...TC.T.A. ..C.TC.T..
HL4-9	C.T..T..A. .TC.T.CT.. T..T..G..T ...TC.T.A. ..C.TC.T..
BR12	C.T..T..A. .TC.C.CT.. T..T..G..T ...TC.A.A. ..C.TC.T..
BZ1	C.T..T..G. .CC.T.CT.. C..C..G..T ...TC.T.A. ..C.TC.T..
VK	C.T..A..A. .TC.C.TT.. T..C..G..T ...TC.T.A. ..C.TC.T..
QB	C.C..T..A. .TC.C.TC.. C..T..G..T ...TC.T.A. ..C.TC.T..
M11	A.G..G..A. .CC.T.CT.. A..T..T..A ...TC.T.T. ..T.AA.G..
MX1	A.A..G..T. .CT.T.CT.. A..A..T..A ...AG.T.T. ..T.AA.G..

	310 320 330 340 350
Consensus	VKKWACMWCY GTWYRGATATG AYCCHDCMRS MYTRYBTTTC AGGATTVYTG
TW18	GGTA..AA.C ..TCA..... .T..CG.AGC AC.ATCG... ----..AT..
HL4-9	GGTA..AA.C ..TCA..... .T..CA.AGC AC.ATCG... ----..AT..
BR12	AGTA..AA.C ..TCA..... .T..CG.AGC AC.ATCG... ----..AT..
BZ1	GGTA..AA.C ..TCA..... .T..TA.AGC AC.GTCG... ----..AT..
VK	AGTA..AA.C ..TCA..... .T..CG.AGC AC.ATCG... ----..GT..
QB	GGTA..AA.C ..TCA..... .T..CG.AGC AC.ATCG... ----..AT..
M11	CTGT..CT.T ..ATG..... .C..AT.CAG CT.GCTT...CC..
MX1	CTGT..CT.C ..TTG..... .C..AA.CAG CT.GCTC...CC..

	360 370 380 390 400
Consensus	ARCRYBMGDG YKGWBTGGGA YWYGGYRWK GGYGAYRSTG SRRAYMTYGT
TW18	.A.GCTC.T. TT.AT..... TTTC..TAAT ..C..TAG.. CGA.CC.T..
HL4-9	.A.GCTC.T. TT.AT..... TTTC..TAAT ..C..TAG.. CGA.CC.T..
BR12	.A.GCTC.T. TT.AT..... TTTC..TAAT ..C..CAG.. CGG.CC.T..
BZ1	.A.GCCC.T. TT.AT..... TTTC..TGAT ..T..TAG.. CGG.CC.T..
VK	.A.GCTC.T. TT.AT..... TTTC..TAAT ..C..CAG.. CGG.CC.T..
QB	.A.GCTC.T. TT.AC..... TTTC..TAAT ..C..TAG.. CGA.CC.T..
M11	.A-ATGA.A. CG.TT..... CAAT..CATG ..T..TAC.. GAG.TA.T..
MX1	.G-ATGA.G. CT.AG..... CAAT..CATG ..T..TGC.. GAG.TA.C..

	410 420 430 440 450
Consensus	CWWTAAWGAC TTYBTBTTYM RYACBYBGC RCCTAARGAK TTYGATTTYT
TW18	.AT...T... ..TT.G..TC GC..TTCC.. A.....G..G ..C.....C.
HL4-9	.AT...T... ..TT.G..TC GT..TTTC.. G.....G..G ..C.....C.
BR12	.AT...A... ..TG.T..CC GT..CTTT.. A.....G..T ..C.....C.
BZ1	.AT...T... ..TT.G..CC GC..TTTC.. A.....G..G ..C.....C.
VK	.AT...A... ..CG.G..TC GC..CTTT.. A.....G..T ..C.....C.
QB	.AT...T... ..TC.G..TC GC..CTTT.. A.....G..G ..T.....T.
M11	.TA...T... ..TC.C..CA AT..GCCG.. A.....G..G ..C.....C.
MX1	.TA...A... ..TC.C..CA GC..CCCG.. A.....A..G ..C.....C.

	460 470 480 490 500
Consensus	CKAAYTCYTT RGYKCCWCGY TAYASYMAKG CCTTCTCYGC BTTTAAYGCV
TW18	.G..T..T.. A.TT..T..T ..T.CTC.G.C... T.....T..C
HL4-9	.G..T..C.. A.TT..T..T ..T.CTC.G.C... T.....T..C
BR12	.G..T..C.. A.CT..T..T ..T.CCC.G.C... C.....T..C
BZ1	.G..C..C.. G.CT..T..C ..T.CCC.G.C... T.....T..C
VK	.G..C..C.. A.TT..T..T ..T.CCC.G.C... T.....T..C
QB	.G..C..C.. A.TT..T..T ..T.CTC.G.C... G.....T..C
M11	.T..T..C.. A.CG..A..C ..C.GTA.T.T... T.....C..T
MX1	.G..T..C.. A.CG..A..C ..C.GTA.T.C... T.....T..C

	510 520 530 540 550
Consensus	AARTATGGCR YTATSATYGG HGAAGGGCDC GARACWMTWA ARTATYTCGS
TW18	..G.....A C...G..C.. C.....T. ..G..TA.A. .A...C...G
HL4-9	..G.....A C...G..C.. C.....T. ..G..TA.A. .A...C...G
BR12	..A.....G T...G..C.. T.....T. ..G..TA.A. .A...C...G
BZ1	..G.....A C...G..C.. C.....T. ..G..TA.A. .A...C...G
VK	..A.....G T...G..C.. T.....T. ..G..TA.A. .A...C...G
QB	..G.....A C...G..C.. C.....T. ..G..TA.A. .A...C...G
M11	..A.....G T...C..T.. A.....G. ..G..AC.T. .G...T...C
MX1	..A.....G T...C..C.. T.....A. ..A..AC.T. .G...T...C

	560 570 580 590 600
Consensus	GCTDTTACTK CGCAGRCTRC RTRARGSWKW CCGYGCGYGT MRRCRYGGMG
TW18	...G.....GA..G. G.G.G.GTTA ...C..T... AAA.GC...C.
HL4-9	...T.....GA..G. G.G.G.GTTA ...C..T... AAG.GT...C.
BR12	...T.....GA..G. G.G.G.GTTA ...C..T... AAG.GC...C.
BZ1	...T.....GA..G. G.G.G.GTTT ...T..T... AAG.AC...C.
VK	...T.....GA..G. G.G.G.GTTA ...T..T... AAG.GC...C.
QB	...T.....GA..G. G.G.G.GTTA ...C..T... AAG.GT...C.
M11	...A.....TG..A. A.A.A.CAGT ...T..C... CGG.AC...A.
MX1	...G.....TG..A. A.A.A.CAGT ...T..C... CGG.AC...A.

	610 620 630 640 650
Consensus	ATTTACGHGS TCTYCGBARR RTYMTYSAST CYTAYMAYAA DGGTMRDTGG
TW18T.C ...T..C.GG G.TA.CC.G. .T..CC.T.. T...AAG...
HL4-9T.C ...T..T.GG G.TA.CC.G. .C..CC.T.. G...AAG...
BR12T.C ...T..C.GG G.TA.CC.G. .T..CC.T.. A...AAA...
BZ1C.C ...T..T.GG G.TA.CC.G. .T..CC.C.. G...AAA...
VKT.C ...T..T.GG G.TA.CC.G. .T..CC.T.. A...AAA...
QBT.C ...T..T.GG G.TA.CC.G. .C..CC.T.. T...AAG...
M11A.G ...C..G.GG A.CC.TG.C. .T..CC.T.. G...CAT...
MX1T.G ...C..G.AA A.CC.CG.C. .T..TA.T.. G...CGT...

	660 670 680 690 700
Consensus	AARCCDRCTA CTGCTGGTAA TCTCTGGCTY GARTTYMGKT AYGGYTHRY
TW18	..G..GA... ..T ..A..TC.T. .T..CC.TAT
HL4-9	..A..AA... ..T ..A..CC.T. .T..TC.TAT
BR12	..A..GA... ..T ..A..TC.T. .T..CC.TAT
BZ1	..A..GA... ..T ..A..TC.T. .T..CC.TAT
VK	..A..GA... ..T ..A..TC.T. .T..CC.TAT
QB	..A..GG... ..T ..A..TC.T. .T..CC.TAT
M11	..G..TG... ..C ..A..TC.T. .T..CC.CGT
MX1	..G..TG... ..C ..G..TA.G. .C..TT.AAC

	710 720 730 740 750
Consensus	BCCBCTCTTY YAYGACATYA RARRYGTAT GNWHGAYTGG MMVMRBMKYM
TW18	G..T....C T.T....C. A.GAC..C.. .TTA..T... CAGAACCGTC
HL4-9	G..T....C T.T....C. A.GAC..C.. .TTA..C... CAGAAGCGCC
BR12	G..T....C T.C....C. G.GAT..T.. .TTA..T... CAGAGCCGCC
BZ1	G..C....C T.T....C. G.GAT..C.. .TTA..T... CAGAATCGTC
VK	G..T....C T.T....C. G.GAT..C.. .CTA..T... CAAAACCGCC
QB	G..T....T T.T....C. G.GAT..C.. .TTA..C... CAGAACCGTC
M11	C..G....C C.T....C. A.GAT..C.. .AAC..C... ACGCGCATTA
MX1	T..G....C C.T....T. A.AGT..T.. .GAT..T... AACCGTATTA

	760 770 780 790 800
Consensus	AYGAYARRAT YCARMRMYWY CKYCGDITYT CDGTBGGTCA YGGYGAGGAY
TW18	.T..T.AA.. T..ACGCCTC .TT..G..T. .T..C.... C..T....C
HL4-9	.T..T.GG.. T..ACGCCTC .TT..G..T. .G..T.... C..T....T
BR12	.T..T.AG.. T..ACGCCTC .TT..A..T. .A..T.... T..T....C
BZ1	.C..T.AG.. T..ACGCCTC .TT..G..T. .T..C.... T..C....T
VK	.T..T.AG.. T..ACGCCTC .TT..G..T. .T..T.... C..C....C
QB	.T..T.AG.. T..ACGCCTC .TT..G..T. .T..T.... C..C....T
M11	.C..T.AG.. C..GAAATAT .GC..A..C. .G..G.... T..T....T
MX1	.C..C.AG.. C..GAAACTT .GC..T..C. .T..T.... C..T....T

	810820830840850
Consensus	TTYRHGYTRT CDARTTHGAY RRYYTRTAYC CHGSYBTWDC YYAYTTYMRR
TW18	..-GC.T.G. .G.A..C..C AACT.A..T. .A.CCG.TG. TT.C..TAAA
HL4-9	..-AT.T.G. .A.A..C..C AGCT.A..T. .A.CCG.TT. TT.C..TAAA
BR12	..-AC.T.G. .A.G..T..C AACT.G..T. .C.CCT.AG. CT.C..TAAA
BZ1	..-AT.T.G. .G.A..T..C AACT.A..T. .C.CCG.AG. TT.C..TAAA
VK	..-AT.T.G. .A.A..C..C AACT.A..C. .C.CTT.AG. CT.C..TAAA
QB	..-AC.T.G. .G.A..C..C AATC.G..C. .T.CCG.TG. TT.C..TAAA
M11	..TAA.C.G. .T.--.C..C GGCT.A..T. .T.GCC.TA. TC.T..CCGA
MX1	..CAA.C.A. .A.--.A..T GGTT.G..T. .T.GCC.AA. CC.C..CAGG

	860870880890900
Consensus	YTRWVHGGBG AGATTACHST CSARCGCCGT CATCGDYRBG GYATADYYTA
TW18	C.GAAA..T.AC. .G.A.....TCAT. .T...TCT..
HL4-9	C.AAGA..G.AC. .G.A.....TCAT. .T...TCT..
BR12	C.GAAA..T.TC. .G.A.....TCAT. .C...TCT..
BZ1	C.GAAA..T.TC. .G.A.....TCAT. .T...TCT..
VK	C.GAAA..T.AC. .G.A.....TCAC. .T...TCC..
QB	C.GAAA..G.AC. .G.A.....TCAT. .C...TCT..
M11	T.ATCT..T.CG. .C.G.....ATGG. .T...GTC..
MX1	T.ATCC..C.CG. .C.G.....GTGG. .T...ACC..

	910920930940950
Consensus	CGCKAAYCGC GRRGGHTAYG CYRYWTTYGA YAACGGTTCC MTTCGGCCYG
TW18	...T..T... .AA..A..T. .TGTT..C.. T..... C.....T.
HL4-9	...T..T... .AG..A..T. .TGTT..C.. T..... C.....T.
BR12	...G..T... .AA..A..T. .TGTT..C.. T..... C.....T.
BZ1	...T..T... .AA..A..T. .TGTT..C.. T..... C.....T.
VK	...G..T... .GG..A..T. .TGTT..C.. C..... C.....T.
QB	...T..C... .AA..A..T. .TGTT..C.. C..... C.....T.
M11	...T..T... .AA..C..C. .CACA..T.. C..... A.....C.
MX1	...T..T... .AA..T..C. .CACA..C.. C..... A.....C.

	9609709809901000
Consensus	TGTCCGAYTG GAAGGARCTY GCYRHYGCDT TYATCAAYCC KSRHGAAGTT
TW18T..G..T ..CACC..A. .C.....T.. GCAT.....
HL4-9T..A..T ..CACT..A. .C.....T.. TCAT.....
BR12T..A..T ..CGTC..G. .C.....T.. TCAC.....
BZ1T..G..T ..CACC..G. .C.....T.. TCAA.....
VKT..G..T ..CGTC..G. .C.....T.. TCAC.....
QBT..G..T ..CACT..A. .C.....T.. GCAT.....
M11C..A..C ..TAAC..T. .T.....T.. TGGC.....
MX1C..A..C ..TAAC..G. .T.....C.. TGGC.....

	10101020103010401050
Consensus	GCNTGGGARY TAACWCCHTA CAGCTTYRTY GYBGATTGGT TYWTSAAAYGT
TW18	..T.....GCT..C..CG.T .TT..... .CT.G..T..
HL4-9	..A.....GTT..C..CG.T .TT..... .TT.G..T..
BR12	..T.....GTT..T..CG.T .CC..... .TT.G..C..
BZ1	..C.....GTT..C..TG.T .TT..... .TT.G..C..
VK	..T.....GTT..T..TG.T .TT..... .CT.G..C..
QB	..T.....GTT..C..CG.T .TT..... .CT.G..T..
M11	..G.....ATT..A..CG.C .TT..... .TA.C..C..
MX1	..A.....ATA..A..CA.C .TG..... .TA.C..C..

	10601070108010901100
Consensus	YGGYGAYATH MTYGMKCARC ARRRKCARYK RTATCAKAAY ATYGABATYG
TW18	C..T..C..A C.T.CT..A. .GGGT..GCT A.....T..T ..C..T..T.
HL4-9	C..T..C..A C.T.CT..G. .GGGT..GCT A.....T..T ..C..T..T.
BR12	T..C..C..A C.T.CT..G. .AGGT..GCT A.....T..T ..C..G..C.
BZ1	C..T..C..A C.C.CT..A. .AGGT..ACT A.....T..T ..C..T..T.
VK	C..C..C..A C.T.CT..G. .AGGT..ACT A.....T..T ..C..T..T.
QB	T..T..C..A C.T.CT..A. .AGGT..GCT A.....T..T ..C..T..T.
M11	T..C..T..C A.C.AG..G. .GAAG..ATT G.....G..C ..C..C..T.
MX1	T..C..T..T A.T.AG..G. .GAAG..ATG G.....G..T ..T..C..T.

	11101120113011401150
Consensus	THGAYGGHTW YSASMGWCGY GAYATHCGNH TSMRHTCNKT YWCYMTTHAAA
TW18	.A..C..T.T TG.CA.A..T ..C..C..GC .CAAA..CT. CA.TA.A...
HL4-9	.A..C..T.T TG.CA.A..T ..C..T..AC .CAAA..TT. CA.TA.A...
BR12	.A..T..T.T CG.CA.A..T ..C..A..GT .CAAA..AT. CA.TA.T...
BZ1	.A..C..C.T CG.CA.A..T ..C..A..GT .CAAA..GT. TA.CA.A...
VK	.A..T..T.T TG.CA.A..T ..C..A..GT .CAAA..GT. TA.CA.T...
QB	.A..C..C.T TG.CA.A..T ..C..C..GC .CAAA..TT. CA.CA.A...
M11	.T..C..A.A CC.GC.A..T ..T..A..TA .GCGC..CG. TA.TC.C...
MX1	.C..C..T.A CC.GC.T..C ..T..A..CA .GCGT..CG. CT.TC.T...

	11601170118011901200
Consensus	GGWGWRCGDA ATGGVMDRCC TGTWMRCGTW WCTGCBRRYY WRTCKRACYT
TW18	..T.AA..A.GCGG.. ...TAA...T T....TGACC TG..TG--C.
HL4-9	..T.AA..A.GCGA.. ...TAA...T T....CGACC TG..TG--C.
BR12	..T.AA..A.ACAG.. ...TAA...A T....GGACT TA..TA--C.
BZ1	..T.AG..A.ACAG.. ...TAA...T T....GGACC TG..TG--C.
VK	..T.AA..A.ACAG.. ...TAA...A T....GGACC TA..TG--C.
QB	..T.AA..A.GCGG.. ...TAA...T T....TAGCC TG..TG--C.
M11	..A.TA..T.CATA.. ...ACG...T A...--GATC AG..GA..T.
MX1	..A.TA..G.CATA.. ...ACG...T A...--GATC AG..GA..T.

	1210 1220 1230 1240 1250
Consensus	RTYGATWYMT TTTAYARYCG MYYYCATRCK ASYMRBMTYC CGYWMGCYAC
TW18	G.C...TTA.C.GC.. ACTC...A.G .GCAGTC.T. ..TTC..T..
HL4-9	G.C...TCA.C.GC.. ACTC...A.G .CCAAGC.T. ..TTC..T..
BR12	A.C...ACA.T.GT.. ACTT...G.T .GCAATA.T. ..TTC..T..
BZ1	G.T...ACA.T.GT.. ACTC...A.G .GCAGTA.T. ..TTC..T..
VK	A.C...ACA.T.GT.. ACTT...G.T .GCAACA.T. ..TTC..T..
QB	G.C...TTA.C.GC.. ACTC...A.G .GCAATC.T. ..TTC..T..
M11	G.T...TCC.C.AT.. CTCT...A.T .CTCGTA.C. ..CAA..C..
MX1	G.T...TCC.C.AT.. CTCT...A.T .CTCGTA.C. ..CAA..C..

	1260 1270 1280 1290 1300
Consensus	ACTHGMWMTY GATACDWCYT TYWSKWSBWW TAARCACGTB HTNGAYAGTR
TW18	...T.ATC.CTA.C. .TAGTTCGTA ...A.....C C.T..T...A
HL4-9	...C.ATC.CTA.C. .TAGTTCGTA ...A.....T C.T..T...A
BR12	...A.ATC.CTA.C. .TAGTTCGTT ...A.....C C.A..T...A
BZ1	...A.ATC.CTA.C. .TAGTTCGTT ...A.....T C.C..T...A
VK	...T.ATC.CTA.C. .TAGTTCGTT ...A.....T C.A..T...G
QB	...A.ATC.TTA.C. .TAGTTCGTT ...A.....T C.T..T...A
M11	...A.AAC.CAT.T. .CTCGAGCAT ...G.....G T.A..T...A
MX1	...A.CAA.CGT.C. .CTCGAGTAT ...A.....G A.G..C...A

STOP 1

	1310 1320 1330 1340 1350
Consensus	TYKYTYTWWT AACYCARGCG RTWAARCGYT G AMMWCWTTG GGTCAATTHG
TW18	.CTT.T.AT. ...C..A... A.T..G..T. ..AA-.T...
HL4-9	.CTT.T.AT. ...C..A... A.T..G..T. ..AA-.T...
BR12	.CTT.T.AT. ...C..A... A.T..G..T. ..AA-.T...
BZ1	.CTT.T.AT. ...C..A... A.T..G..C. ..AAT.T...
VK	.CGC.T.AT. ...C..A... A.T..G..T. ..AA-.T...
QB	.CTT.T.AT. ...C..A... G.A..G..T. ..AA-.T...
M11	.CTC.C.TA. ...T..G..N A.A..A..C. ..CCA.A...
MX1	.TTC.C.TA. ...T..G... A.A..A..C. ..CCA.A...

ORF2/3

	1360 1370 1380 1390 1400
Consensus	ATC A TGGCWA AATTASARRC TRTYACTTTA RGTRRYATYG GGAARRAHGG
TW18A.G.GA. .G.T..... A..AAC..C.AG.T..
HL4-9A.G.GA. .G.T..... A..AAC..C.AG.T..
BR12A.G.GA. .G.T..... A..AAC..C.AG.T..
BZ1A.G.GA. .G.T..... A..AAC..T.AG.T..
VKA.G.GA. .G.T..... A..AAC..C.AG.T..
QBA.G.GA. .G.T..... G..AAC..C.AG.T..
M11T.C.AG. .A.C..... A..GGT..T.GA.A..
MX1T.C.AG. .A.C..... A..GGT..T.GA.C..

	14101420143014401450
Consensus	WVAVVWWACT CTGRWCCTCA AYCCRCGTGG GGTAATATCCC ACYAACGGYG
TW18	AC.ACAA... ..GT..... .C..G..... ..T.....T.
HL4-9	AA.ACAA... ..GT..... .T..G..... ..T.....T.
BR12	AA.GCAA... ..GT..... .C..G..... ..T.....T.
BZ1	AC.AAAA... ..AA..... .C..A..... ..C.....T.
VK	AA.ACAA... ..GT..... .T..G..... ..T.....T.
QB	AA.ACAA... ..GT..... .T..G..... ..T.....C.
M11	TG.CGTT... ..GA..... .C..G..... ..T.....T.
MX1	TG.CGTT... ..AA..... .C..G..... ..C.....T.

	14601470148014901500
Consensus	TTGCCCKCGCT TTCDSAAGCG GGTGCAGTTC CTGCRYTGGA GAAGCGTGTY
TW18G.... ..AC..... ..AC..... ..T
HL4-9G.... ..AC..... ..GC..... ..T
BR12T.... ..AG..... ..AT..... ..T
BZ1T.... ..AG..... ..AT..... ..T
VKT.... ..GG..... ..AT..... ..T
QBT.... ..AC..... ..GC..... ..T
M11G.... ..TG..... ..GT..... ..C
MX1G.... ..AG..... ..AT..... ..T

	15101520153015401550
Consensus	ACMRTTTCKG TRTCHCAGCC TTCYCGYAAT CGYAAGAAAT ACAARGTYCA
TW18	..CG....G. .A..C..... ..T..C... ..C.....C.G..C..
HL4-9	..CG....G. .A..A..... ..T..T... ..T.....C.G..C..
BR12	..CG....G. .A..T..... ..T..T... ..T.....C.G..C..
BZ1	..CA....G. .A..T..... ..T..T... ..T.....C.G..T..
VK	..CG....G. .A..T..... ..C..C... ..T.....C.G..C..
QB	..CG....G. .A..T..... ..T..C... ..T.....C.G..C..
M11	..AA....T. .G..A..... ..T..C... ..T.....T.G..T..
MX1	..AA....T. .A..A..... ..T..C... ..T.....C.A..T..

	15601570158015901600
Consensus	RGTWAARATC CARAACCRA CCKCTTGYHC TGCAARCGGT WCTTGTGACC
TW18	G..T..G... ..G.....G. ..G....CA.A.... T.....
HL4-9	A..T..G... ..G.....G. ..G....TA.A.... T.....
BR12	G..T..A... ..G.....G. ..G....TC.A.... T.....
BZ1	G..T..A... ..A....A. ..G....TT.A.... T.....
VK	G..T..A... ..A....G. ..G....TT.A.... T.....
QB	G..T..G... ..G.....G. ..G....CA.A.... T.....
M11	G..A..G... ..G....A. ..T....CA.G.... A.....
MX1	G..A..G... ..G....A. ..T....CA.G.... A.....

	1610 1620 1630 1640 1650
Consensus	CWTCMGTTAC TCGYYMRGCH TATKCTGAYG TGACBTTYTC GTTCACRCAG
TW18	.A..C..... ...CCAG..A ...G....C.T..C..G...
HL4-9	.A..C..... ...CCAG..A ...G....C.T..T..G...
BR12	.A..C..... ...CCAG..A ...G....C.T..T..G...
BZ1	.A..C..... ...CCAG..A ...G....C.T..T..G...
VK	.A..C..... ...CCAG..A ...G....C.C..T..G...
QB	.A..C..... ...CCAG..A ...G....C.C..T..G...
M11	.T..A..... ...TTCA..C ...T....T.G..C..A...
MX1	.T..A..... ...TTCG..T ...G....C.G..C..G...

	1660 1670 1680 1690 1700
Consensus	TAYAGYACYG WTGAGGAACG HGCWYTYGTW CGHACAGAGC TYVHHGICYCT
TW18	..T..T..C. A..... A..TT.T..T ..T..... .TATC..T..
HL4-9	..T..T..C. A..... A..TT.T..T ..T..... .TGTC..T..
BR12	..T..C..C. A..... A..TT.T..T ..T..... .TGCT..T..
BZ1	..T..T..C. A..... A..TT.T..T ..T..... .TGCT..T..
VK	..T..T..C. A..... A..TT.T..T ..C..... .TGCT..T..
QB	..T..T..C. A..... A..TT.T..T ..T..... .TGCT..T..
M11	..T..C..T. T..... C..AC.C..A ..A..... .CCAA..T..
MX1	..C..C..T. A..... T..AC.C..A ..A..... .TAAA..C..

	1710 1720 1730 1740 1750
Consensus	GYTVGCKRRT CCWMTGYTKR TYRATGCTAT YGAYMRNYTG AAYCCRGCR
TW18	.C.C..TAG. ..TC..C.GA .CG..... T..TCAGC.. ..T..A..G.
HL4-9	.C.C..TAG. ..TC..T.GA .TG..... C..TCAGC.. ..C..A..G.
BR12	.C.C..TGG. ..TC..C.GA .CG..... C..TCGAC.. ..C..A..G.
BZ1	.C.C..TGA. ..TC..C.GA .CG..... C..TCAGC.. ..C..A..G.
VK	.C.C..TGG. ..TC..C.GA .CG..... C..TCGAC.. ..T..A..A.
QB	.C.C..TAG. ..TC..C.GA .CG..... T..TCAGC.. ..C..A..G.
M11	.T.A..GGA. ..TA..C.TG .CA..... C..CAATC.. ..T..G..G.
MX1	.T.G..GGA. ..AA..C.TA .CG..... C..TAACT.. ..T..G..G.

STOP 2

	1760 1770 1780 1790 1800
Consensus	AYTGAACDBY RCTYMTTGCC GGTGDTGGCT CAGGGBMWAR MCCYGDWYCG
TW18	.T.....ACT G..CA.....G.....TCA.A C..C.ATT..
HL4-9	.T.....ACT G..CA.....G.....TCT.A A..C.ATC..
BR12	.T.....ATT G..CA.....G.....GAA.A C..C.ATC..
BZ1	.T.....GTT G..CA.....G.....GAA.A C..C.ATC..
VK	.T.....ATT G..CA.....G.....GAA.A C..C.ATC..
QB	.T.....ACT G..CA.....G.....TCA.A A..C.ATC..
M11	.T.....TGC G..TC--- --...T.....CCT.G C..T.GAC..
MX1	.C.....TGC A..TC--- --...A.....CCT.G C..T.TAC..

	1810 1820 1830 1840 1850
Consensus	GKTVHYRWTC CGGAYCCRCR RMYBGAKCCG CCGCCAGGGW CAGGYAVNTA
TW18	.T.ATC---.T..A.. GATT..T...AC.AG..
HL4-9	.T.ATCAT..T..A.. GATC..T...AT.AA..
BR12	.T.GAT---.T..A.. GATT..T...AT.GT..
BZ1	.T.GAT---.T..A.. GATT..T...TT.GC..
VK	.T.AAT---.T..A.. GATT..T...AT.CC..
QB	.T.ATT---.T..A.. GATT..T...AT.AG..
M11	.G.CCTGA..C..G.. ACCG..G...AT.GT..
MX1	.G.CCTAA..C..G.. ACTG..G...AT.GT..

	1860 1870 1880 1890 1900
Consensus	TACYTGTCY TTCSSWATWT GGKMHYTDKM VRRSRTTTAY GARSCTSCKA
TW18	...C.....C ...GCA..T. ..TCCC.AGA GGAGG....C ..GC..C.T.
HL4-9	...T.....T ...GCA..T. ..TCCC.AGA GGAGG....C ..GC..C.T.
BR12	...T.....C ...GCA..T. ..TCTC.GGA AGAGG....T ..GC..C.T.
BZ1	...C.....T ...GCA..T. ..TCAT.AGA GGAGG....T ..GC..C.T.
VK	...T.....T ...GCA..T. ..TCTC.AGA GGAGG....T ..AC..C.T.
QB	...C.....C ...GCA..T. ..TCCC.AGA GGAGG....C ..GC..C.T.
M11	...C.....T ...CGT..A. ..GATC.GTC CAGCG....T ..AG..G.G.
MX1	...C.....T ...CGT..A. ..GATC.TTC CAGCA....T ..AG..G.G.

	1910 1920 1930 1940 1950
Consensus	MYADKDMYCR MYCGTGGSMT ATYTAYHRYG CTRTYGAACT YHVDYCWCGY
TW18	CT.GGAAC.G AC.....CC. ..C..CAAT. ..G.C..... CCGGC.T..C
HL4-9	CT.AGAAC.G AC.....CC. ..C..TAAC. ..A.C..... CCAGC.T..C
BR12	CC.TTGAT.G AC.....CC. ..C..TCGT. ..G.T..... CTCGT.T..C
BZ1	CC.GTGAC.G AC.....CC. ..C..TTAT. ..G.C..... CCAAC.T..T
VK	CT.GTGAC.G AC.....CC. ..C..TCAT. ..G.T..... CCCGC.T..C
QB	CT.AGAAC.G AC.....CC. ..C..TAAT. ..G.T..... CCAGC.T..C
M11	AC.GTTCT.A CT.....GA. ..T..CAAT. ..G.C..... TAGTC.A..C
MX1	AT.GTTCC.A CT.....GA. ..C..TAAC. ..G.C..... CAGTC.T..C

	1960 1970 1980 1990 2000
Consensus	RADTTTGAYG TYRCCCTYRA WGAYCTYTTG GGYAAYACWR ANTGGCGHGA
TW18	A.G.....T. .TG....CG. A..T..T... ..C..T..AA .T.....C..
HL4-9	A.G.....T. .TG....TA. A..T..T... ..C..T..AA .A.....C..
BR12	A.G.....T. .TG....TG. T..T..C... ..T..C..TA .A.....T..
BZ1	G.T.....T. .TG....TG. T..T..C... ..T..C..TA .A.....A..
VK	G.T.....T. .TG....CG. T..T..C... ..T..C..TG .A.....T..
QB	G.A.....T. .TG....CA. A..T..T... ..C..T..AA .G.....T..
M11	A.T.....C. .CG....TG. T..T..C... ..C..C..AA .C.....T..
MX1	A.A.....C. .TA....TG. T..C..C... ..T..C..AG .T.....C..

	2010 2020 2030 2040 2050
Consensus	YTGGGAYKSH MGRCTBAGDT AYACCACGTT YCGCGGTWGC CGWRGYAAYG
TW18	C.....TTCA C.G..G..T. .T..... C.....T.. ..TG.C..T.
HL4-9	C.....TTCA C.G..T..T. .T..... T.....T.. ..TA.C..C.
BR12	C.....TTCT C.G..T..G. .T..... C.....T.. ..TG.C..T.
BZ1	C.....TTCA C.G..T..T. .T..... C.....T.. ..TG.C..T.
VK	C.....TTCA C.G..C..A. .T..... C.....T.. ..TG.C..C.
QB	T.....TTCT C.G..T..T. .T..... C.....T.. ..TG.C..T.
M11	C.....CGGC A.A..G..A. .C..... T.....T.. ..TG.C..C.
MX1	T.....CGGC A.A..T..G. .T..... T.....A.. ..AG.T..C.

	2060 2070 2080 2090 2100
Consensus	GDTAYATTGA CCTYGAYGCV WCTTMKYTBR YKMDGAYSA RTRYNWTRCD
TW18	.T..T..... ..C..T..G A...ATC.TG CTACT..TC. G-GCTA.G.G
HL4-9	.T..T..... ..T..T..G T...ATC.TG CTACT..TC. G-GCTA.G.G
BR12	.T..T..... ..T..T..G A...ATC.CG CTACT..TC. A-GCTA.G.T
BZ1	.T..T..... ..T..T..G A...ATC.TG CTACT..CC. A-GCGA.G.T
VK	.T..T..... ..T..T..G A...ATC.CG CTACT..CC. A-GCAA.G.T
QB	.T..T..... ..T..T..G A...ATC.TG CTACT..TC. G-GCTA.G.G
M11	.G..C..... ..T..C..A A...CGT.GA TGAAA..TG. A.ATCT.A.A
MX1	.A..C..... ..T..C..C A...CGT.GA TGCAG..TG. A.ATCT.A.A

	2110 2120 2130 2140 2150
Consensus	YSDTCAGAAG TAYBWWRTKC GYRMRGGSAA RMRWCCYGGT GYYTTYGGYW
TW18	TGA..... ..CGATA.T. .CGAG..C.. AAAA..T... .CT..C..TA
HL4-9	TGA..... ..CGATA.T. .CGAA..C.. GAAA..C... .CT..C..TA
BR12	CGA..... ..CGATA.T. .CACG..C.. AAGA..T... .CC..T..CA
BZ1	TGA..... ..CGATA.T. .CACG..C.. GAAA..C... .CC..C..TA
VK	TGA..... ..CGATA.T. .CACA..C.. AAGA..T... .CC..C..TA
QB	TGA..... ..TGATA.T. .CGAG..C.. GAAA..T... .CT..C..TA
M11	TCG..-..... ..TCTTG.G. .TGAA..G.. ACGT..C... .TC..C..TA
MX1	TCT..-..... ..TTTAG.G. .TGAG..G.. ACGT..C... .CT..C..TT

	2160 2170 2180 2190 2200
Consensus	MHRTYGARCR RTTYRTYTAT CTTAARTCGA TAAAYGCHTA YTGYTCKCTY
TW18	ACA.T..G.G A..CA.T... ..G.... ..T..T.. T..C..T..T
HL4-9	ACG.T..G.G A..CA.T... ..G.... ..T..T.. T..C..T..T
BR12	AAA.T..A.A A..TA.T... ..G.... ..T..A.. T..T..T..C
BZ1	ACA.C..A.A A..CA.C... ..G.... ..T..C.. T..T..T..T
VK	ATA.T..A.A A..TA.T... ..G.... ..T..A.. T..T..T..T
QB	ACA.T..G.G A..CA.T... ..G.... ..T..T.. T..C..T..T
M11	ACA.C..G.G G..TG.T... ..G.... ..C..T.. C..C..G..C
MX1	CAA.T..A.G G..TG.T... ..A.... ..C..A.. C..T..T..T

	2210 2220 2230 2240 2250
Consensus	AGCGATATTR CGGCCTAYCR CDCCGAYGGH GTRRTAGTTG GCTTTTGGCG
TW18G.....T.A..G....T..T..GA.....
HL4-9G.....T.A..G....T..T..GA.....
BR12G.....T.A..G....C..C..AA.....
BZ1G.....T.A..G....T..T..GA.....
VKG.....T.A..G....T..C..AA.....
QBG.....T.A..G....T..C..GA.....
M11A.....T.G..A....T..T..GA.....
MX1A.....C.A..T....C..A..GG.....

	2260 2270 2280 2290 2300
Consensus	YGAYCCRTCH AGYGGKGGWG CCATACCR TT YGACTTYAVN VARTTTGAYW
TW18	C..T..A..T..T..T..T..G..T....C.CT A.G....TA
HL4-9	C..T..A..T..T..T..T..G..T....C.CT A.G....TA
BR12	C..T..A..T..T..T..T..G..T....C.CT C.G....TA
BZ1	C..T..A..T..T..T..T..G..T....C.CA G.G....TA
VK	C..T..A..T..T..T..T..G..T....C.CG G.A....TA
QB	C..T..A..C..T..T..T..G..T....C.CT A.G....TA
M11	T..C..G..A..C..G..A..A..C....T.AC G.A....CT
MX1	C..C..G..A..C..G..A..A..C....T.GC G.G....TT

STOP 3

	2310 2320 2330 2340 2350
Consensus	MGAMYAAATG TCCDATHCAA GCYGTKATWG TCGTTCCTCG TSYT AG TAA
TW18	A..CT.....T..T...C..G..A.....GC.....
HL4-9	A..CT.....T..T...C..G..A.....GC.....
BR12	A..CT.....T..T...C..G..A.....GC.....
BZ1	A..CT.....A..T...T..G..A.....GC.....
VK	A..CT.....G..T...C..G..A.....GC.....
QB	A..CT.....T..T...C..G..A.....GC.....
M11	C..AC.....G..A...C..T..A.....CT.....
MX1	C..AC.....G..C...C..T..T.....CT.....

ORF4

	2360 2370 2380 2390 2400
Consensus	CTRARGRWGA WMTGC ATG TC WAAGACABYR YMBTCGCRTA ASTCKCTYAG
TW18	..A.G.AT..AA.....T.....GCA TCT....A..C..T..T..
HL4-9	..A.G.AT..AA.....T.....GCA TCT....A..C..T..T..
BR12	..A.G.AT..AA.....T.....GCA TCT....A..C..T..T..
BZ1	..A.G.AT..AA.....T.....GCA TCC....G..C..T..C..
VK	..A.G.AT..AA.....T.....GCA TCT....A..C..T..T..
QB	..A.G.AT..AA.....T.....GCA TCT....G..C..T..C..
M11	..G.A.GA..TC.....T.....TCG CAG....G..G..G..T..
MX1	..A.A.GA..TC.....A.....CTG CAG....G..G..G..T..

	2410 2420 2430 2440 2450
Consensus	CGSAMAAYTS CGCCGHGCGY CGAACACAAG AATYGWGGTH GAAGRTAACC
TW18	..C.C..T.GA..T.T.A...AG.....
HL4-9	..C.C..T.GA..T.T.A...CG.....
BR12	..C.C..T.GA..T.T.A...TG.....
BZ1	..C.C..T.GA..C.T.A...CG.....
VK	..C.C..T.GA..T.T.A...CG.....
QB	..C.C..T.GA..C.T.A...TG.....
M11	..G.A..C.CT..T.C.T...TA.....
MX1	..G.A..C.CC..T.C.T...TG.....

	2460 2470 2480 2490 2500
Consensus	TCGCACTKTC YATYGCVAAY GAYYTAHTRT YBGCNYWDGR TSWDKMRYCR
TW18T.. C..T..C..C ..TT..C.A. TG..TTAT.G .CAATCGC.A
HL4-9T.. C..C..C..C ..TC..C.G. TG..TTAT.G .CAATCGC.A
BR12T.. C..T..C..C ..CT..A.G. TG..ATAT.G .CAATCGC.A
BZ1T.. C..T..C..C ..TT..A.G. TG..TTAT.G .CAATCGC.A
VKG.. C..T..C..C ..TT..A.G. TG..ATAT.G .CAATCGC.A
QBT.. C..T..C..C ..TT..C.G. TG..CTAT.G .CAGTCGC.A
M11T.. C..C..A..C ..TC..T.G. CT..GTTG.A .GTTGAGT.G
MX1G.. T..C..G..T ..TC..T.A. CC..GCTA.A .GTAGAAC.G

	2510 2520 2530 2540 2550
Consensus	TTTARYTCBG ARKMNGASTG TATHWSHYKY DSHCCGARAT TYGRCVDVWC
TW18AC..T. .AGCT..G.. ...TTCACTC AGT....G.. .C.A.GGGA.
HL4-9AC..C. .AGCT..G.. ...TTCACTC AGT....G.. .T.A.GGGA.
BR12GT..T. .GTCT..G.. ...TTCACTC AGC....A.. .C.A.GGAA.
BZ1AT..C. .GTCC..G.. ...TTCACTC GGT....A.. .C.A.GAAA.
VKGT..C. .GTCT..G.. ...TTCACTC AGC....A.. .C.A.GGAA.
QBAC..T. .GGCT..G.. ...TTCATTC AGC....G.. .C.A.GGGA.
M11GT..G. .GGAG..C.. ...CAGCCGC TCC....A.. .C.A.CTCT.
MX1AT..G. .AGAA..C.. ...AAGTCGT TCA....A.. .C.G.ATCT.

	2560 2570 2580 2590 2600
Consensus	HSCRGAYVAM TTTAGGAWWW MYTATCTTVV WGCYGAGRTH ATGTCGAAGT
TW18	CC.G..TG.CTAA AT.....AA A..C...G.C
HL4-9	CC.G..TC.CTAA AT.....AA A..C...G.C
BR12	CC.G..TA.CTAA AT.....AA A..C...A.C
BZ1	CC.G..CA.CTAA AT.....AA A..C...A.C
VK	CC.G..TA.CTAA AT.....AA A..C...A.C
QB	CC.G..TG.CTAA AT.....AA A..C...A.C
M11	TG.G..TC.AATT CC.....GC T..C...A.T
MX1	AC.A..TC.AATT CC.....CG T..T...A.A

	2610 2620 2630 2640 2650
Consensus	AYGAYKMHTT YAGCCTAGGT ATYRATACCG AAGCYGYWGC MTGGRARAAG
TW18	.T..CGAT.. C..... ..TG.....T.CT.. C...G.A...
HL4-9	.T..CGAT.. C..... ..TG.....T.CT.. C...G.A...
BR12	.T..CGAT.. C..... ..TG.....T.CT.. A...A.G...
BZ1	.T..CGAT.. C..... ..TG.....T.CT.. A...G.G...
VK	.T..CGAT.. C..... ..CG.....T.CT.. A...G.A...
QB	.T..CGAC.. C..... ..TG.....T.TT.. C...G.G...
M11	.T..CTCA.. C..... ..TA.....C.TA.. C...G.G...
MX1	.C..TTCT.. T..... ..TA.....C.TA.. A...G.A...

	2660 2670 2680 2690 2700
Consensus	TTYCTRGCGW CRGAGGCTGA RTGTGCTWWR ACGAAYSHKM GHCTCTATAG
TW18	..C..G..A. .A..... A.....TTACGCTC .T.....
HL4-9	..C..G..A. .A..... A.....TTACGCTC .T.....
BR12	..T..A..A. .A..... A.....TTACGCTC .T.....
BZ1	..C..A..A. .A..... A.....TTACGCTC .T.....
VK	..C..A..A. .A..... A.....TTACGCTC .C.....
QB	..C..G..A. .A..... A.....TTACGCTC .T.....
M11	..C..G..T. .G..... G.....ATATCAGC .T.....
MX1	..C..A..T. .G..... G.....AAGTCTGA .A.....

	2710 2720 2730 2740 2750
Consensus	RCCTRACTAC ARTGAGGATT TCAATTTCTC AYTGGGYGAG DCRTGTMTWC
TW18	G...G..... .G..... ..C....C... T.A...A.T.
HL4-9	G...G..... .G..... ..C....C... T.A...A.T.
BR12	A...A..... .G..... ..C....T... T.G...C.T.
BZ1	G...A..... .G..... ..C....C... T.A...A.T.
VK	A...A..... .G..... ..C....T... T.A...C.T.
QB	G...G..... .G..... ..C....C... T.A...A.A.
M11	A...A..... .A..... ..T....T... G.A...A.T.
MX1	G...A..... .A..... ..T....T... A.A...A.T.

	2760 2770 2780 2790 2800
Consensus	ACATGGCYCG YMGAAAAATA GYYAAGYTRW TAGGAGAYKY NSYKYCSKTT
TW18T.. TA..... .CC...T.AACGC TCCGT.CG..
HL4-9T.. CA..... .TC...C.AATGC TCCGT.CG..
BR12T.. CC..... .TT...T.GATGC TCCGT.CG..
BZ1T.. TA..... .TT...C.AACGC GCCGT.CG..
VKT.. CC..... .TT...T.AACGC GCCGT.CG..
QBT.. TA..... .CC...C.AATGT TCCGT.CG..
M11C.. CC..... .TT...T.ATCTC CGTTC.GT..
MX1T.. CC..... .TT...C.ATTTC AGTTC.GT..

	28102820283028402850
Consensus	GAGGSTRIGY TGCGHCAYTG CCGDTTTCY GGYGGTGCYA CAACRACGAA
TW18G.A..TT..C.. ...T.....C ..T.....C.A.....
HL4-9G.A..TT..C.. ...A.....T ..C.....C.A.....
BR12G.A..TT..T.. ...A.....T ..C.....C.A.....
BZ1G.A..CT..C.. ...T.....C ..T.....T.A.....
VKG.A..TC..T.. ...G.....T ..C.....C.A.....
QBG.A..TT..C.. ...A.....T ..C.....T.A.....
M11C.A..TA..C.. ...T.....T ..T.....C.G.....
MX1C.G..TA..T.. ...G.....C ..C.....C.A.....

	28602870288028902900
Consensus	TARCCGTYYR YAYGGYCATC CGTCCTTCAA GTTTGCKCTT VCRCAAGMGT
TW18	..A....TCG T.C..T....G... C.A....C..
HL4-9	..A....TCG C.C..C....G... C.A....C..
BR12	..A....TCA T.T..T....G... C.A....C..
BZ1	..A....TCA T.C..T....G... C.A....C..
VK	..A....TCA T.C..T....G... C.A....C..
QB	..A....TCG T.C..T....G... C.G....C..
M11	..A....TCA T.C..T....T... A.G....A..
MX1	..G....CTA T.C..C....T... G.G....A..

	29102920293029402950
Consensus	GTACSCCWCG GGCTKTKMMR TAYGTKYW RG CYATHARRGC YTSWACRVRY
TW18G..T..T.GAAA ..T..TTTA. .TC.T.GA.. T.CT..ACAT
HL4-9G..T..T.GAAA ..T..TTTG. .TC.T.GA.. T.CT..ACAT
BR12G..T..T.GAAA ..T..TTTG. .TC.T.GA.. T.CA..GCAT
BZ1G..T..T.GAAG ..T..TCTG. .TC.T.GA.. C.CT..GCAC
VKG..T..T.GAAA ..C..TTTG. .TC.C.GA.. C.CA..ACAT
QBG..T..T.GAAG ..T..TTTA. .TC.C.GA.. T.CT..ACAT
M11G..A..G.TCCA ..C..GCAG. .CC.T.AG.. C.GT..AGGT
MX1C..A..G.TCCA ..T..GCAA. .TT.A.AG.. C.GT..AAAC

	29602970298029903000
Consensus	WTS GAYMTYR GDRTYWCYRA KRTYAGCCCT TTYAATAAAG CAGTTACTGT
TW18	T.C..TA.CA .AA.TT.TG. TA.T..... ..T.....
HL4-9	T.C..TA.CA .GG.TT.TG. TA.T..... ..T.....
BR12	T.C..TA.CA .AG.TT.TG. TA.T..... ..T.....
BZ1	T.C..TA.CA .AG.TT.TG. TA.T..... ..T.....
VK	T.C..TA.CA .AG.TT.TG. TA.T..... ..T.....
QB	T.C..TA.CA .AA.TT.TG. TA.T..... ..T.....
M11	A.G..CC.TG .TA.CA.CA. GG.C..... ..C.....
MX1	A.G..CC.TG .TA.TA.CA. GG.T..... ..C.....

	3010 3020 3030 3040 3050
Consensus	ACCWAAGAAC AGTAARACHG ATCGYTGAT HGCYATCGAR CCHGGYTGGA
TW18	...T..... ..A..C.T..T.. T..T.....G ..T..C....
HL4-9	...T..... ..A..C.T..T.. T..T.....A ..C..C....
BR12	...A..... ..A..C.C..T.. T..T.....A ..C..T....
BZ1	...A..... ..A..C.C..T.. C..T.....A ..C..T....
VK	...A..... ..G..C.C..T.. T..T.....A ..C..T....
QB	...T..... ..G..A.T..T.. T..T.....A ..T..T....
M11	...A..... ..A..T.C..T.. A..C.....A ..T..C....
MX1	...A..... ..A..T.C..C.. C..T.....G ..A..C....

	3060 3070 3080 3090 3100
Consensus	ATATGTTTTT YCARYTRGGY ATYGGTGGYR THHTWCGYGA HMRGYTGCRY
TW18 T..AC.A..C ..C.....CA .TC.A..C.. TCG.T...GT
HL4-9 T..AC.A..C ..C.....CA .CC.A..C.. TCG.T...GT
BR12 C..AT.A..T ..C.....CA .TT.A..T.. TCG.T...GT
BZ1 T..AC.A..T ..C.....CA .CT.A..C.. CCG.C...GT
VK C..AT.A..T ..C.....CA .CC.A..C.. TCG.T...GT
QB C..AC.G..T ..C.....CA .TC.A..C.. TCG.T...GT
M11 C..GC.A..T ..C.....TG .TA.A..C.. AAA.T...GT
MX1 C..GT.G..C ..T.....CG .AA.T..C.. AAA.T...AC

	3110 3120 3130 3140 3150
Consensus	TKBTGGRRYA TCGAYCTGAA YGATCAGACG ATWAAYCARV NBCGCGCWYA
TW18	.GC...GGT.T..... T..... ..A..T..GC GC.....TC.
HL4-9	.GC...GGT.T..... C..... ..A..T..GC AT.....TC.
BR12	.GT...GGT.C..... T..... ..A..C..GC AC.....TC.
BZ1	.GC...GGT.T..... T..... ..A..C..GC GT.....TC.
VK	.GC...GGT.C..... T..... ..A..C..GC GC.....TC.
QB	.GC...GGT.T..... T..... ..A..T..GC GC.....TC.
M11	.TG...GGC.T..... T..... ..T..C..AA CG.....AT.
MX1	.TG...AAT.C..... T..... ..T..C..GG TG.....AT.

	3160 3170 3180 3190 3200
Consensus	YKHRGGCWSY BKTRVYRATR AHYTHGCNAC RGTKGATCTC TCRRSVGCDA
TW18	CGAA...TCC GT.ACTA..A .TT.A..A.. G..T..... ..AGCG..A.
HL4-9	CGAA...TCC GT.ACTA..A .TT.A..A.. G..T..... ..AGCG..A.
BR12	CGAG...TCC GT.ACTA..G .TT.A..A.. G..T..... ..AGCG..A.
BZ1	CGAA...TCT GT.ACTA..A .CT.A..A.. A..T..... ..AGCG..A.
VK	CGAA...TCC GT.ACTA..G .TT.A..G.. G..T..... ..AGCA..A.
QB	CGAA...TCC GT.ACTA..A .CT.A..A.. G..T..... ..AGCG..A.
M11	TTTA...AGC CG.GATG..A .TC.C..C.. G..G..... ..AAGA..T.
MX1	TTCA...AGC TG.AGCA..G .AC.T..T.. A..G..... ..GAGC..G.

	3210 3220 3230 3240 3250
Consensus	GYGATWCTAT WTCKCTTGCB CTYKKYGAGC TCYTDHTRCC YCCWGVBTGG
TW18	.C...T.... A..T.....T ..CTGT.... ..T.AC.G.. C..A.GC...
HL4-9	.C...T.... A..T.....T ..CTGT.... ..T.AT.G.. C..A.GT...
BR12	.C...T.... A..T.....T ..CTGT.... ..T.AT.A.. T..A.GT...
BZ1	.C...T.... A..T.....T ..CTGC.... ..T.GT.G.. T..A.GG...
VK	.C...T.... A..T.....T ..CTGC.... ..T.AT.G.. T..A.GT...
QB	.C...T.... A..T.....C ..CTGT.... ..T.AT.G.. C..A.GC...
M11	.C...A.... T..G.....C ..TGTT.... ..C.TA.G.. T..T.AG...
MX1	.T...A.... T..G.....G ..CGTT.... ..C.GC.A.. C..T.CG...

	3260 3270 3280 3290 3300
Consensus	TTTRARGTYC TKWYGGMYYT HMGRTCACSH ARRGGBMDVT TGCCWRAYGG
TW18	...G.G..C. .TAT..ACC. CA.A....CT .AA..GCGG.TG.C..
HL4-9	...G.G..T. .TAT..ATC. CA.A....CA .AA..GCGG.TG.C..
BR12	...G.G..C. .TAC..ATC. CA.A....CT .AG..GCAA.TG.C..
BZ1	...G.A..C. .TAT..ACC. TC.A....CT .AG..GCGA.TA.C..
VK	...G.G..C. .TAT..ACC. CA.A....CT .AG..GCGA.TG.C..
QB	...G.G..T. .TAT..ACC. CA.A....CT .AG..GCGA.TG.C..
M11	...A.G..C. .GTT..CCT. AA.A....CC .AG..CATC.AG.T..
MX1	...A.A..C. .TAC..ACC. TA.G....GA .GG..TATG.AG.C..

	3310 3320 3330 3340 3350
Consensus	TASHRTYRTT AYYTAYGAGA ARATWTCYTC NATGGGTAAY GGHTWYACMT
TW18	..GTG.TG.. .CC..T.... .G..T..C.. T.....C ..A.AC..A.
HL4-9	..GTG.TG.. .CC..T.... .A..A..C.. G.....C ..A.AC..A.
BR12	..GTG.TA.. .CC..T.... .A..T..T.. C.....T ..T.AC..A.
BZ1	..GTG.CG.. .CT..T.... .G..T..T.. C.....C ..T.AC..A.
VK	..GTG.CA.. .TC..C.... .G..T..T.. C.....T ..T.AC..A.
QB	..GTG.TG.. .CC..C.... .G..T..T.. T.....C ..T.AC..A.
M11	..CCG.CA.. .CT..T.... .A..A..C.. A.....T ..C.AT..C.
MX1	..GAA.CA.. .CC..T.... .A..T..C.. A.....C ..T.TC..C.

	3360 3370 3380 3390 3400
Consensus	TCGARCTYGA GTCGCTTATH TTYGCDKCTC TYGCRYCGKTC YKTWTGYGAR
TW18G..T..C ..T..TT... .T..T..T.. CG.T..T..G
HL4-9G..T..C ..T..TT... .T..T..T.. CG.T..T..G
BR12G..C..T ..T..TT... .C..C..T.. CG.T..T..G
BZ1A..C..T ..T..TT... .T..T..T.. CG.T..T..G
VKG..T..C ..C..TT... .T..T..T.. CG.T..T..G
QBG..C..T ..T..TT... .C..T..T.. CG.T..T..G
M11G..T..A ..T..GG... .T..T..G.. TT.A..C..A
MX1G..C..A ..T..AG... .T..T..G.. TT.A..C..G

	34103420343034403450
Consensus	WTACTGRRCT TASRMYCRTC DRRKGTYACK GTYTAYGGMG AYGAYATWAT
TW18	A.....GA.. ..GACT.A.. TGAG..C..T ..C..C..A. .C..T..T..
HL4-9	A.....GA.. ..GACT.G.. TGAG..C..T ..T..C..A. .C..T..T..
BR12	A.....GA.. ..GACT.A.. TGAG..C..T ..T..C..A. .T..C..T..
BZ1	A.....GA.. ..GACT.A.. TGAG..C..T ..C..C..A. .C..C..T..
VK	A.....AA.. ..GACT.A.. TGAG..C..T ..T..C..A. .T..C..T..
QB	A.....GA.. ..GACT.G.. TGAG..C..T ..T..C..A. .C..T..T..
M11	T.....GG.. ..CGAC.G.. AGAT..T..G ..C..T..C. .T..C..A..
MX1	T.....AA.. ..CAAC.G.. GAGT..C..G ..C..T..C. .T..T..T..

	34603470348034903500
Consensus	HTTRCCRTCM BRYGCRKKYM SYBCBYTNVD KGAAGTYTTY WMSTAYGTWG
TW18	C..A..G..C TGT..AGTCC CTG.CC.TCG G.....C..T AAG..T..T.
HL4-9	T..A..G..C TGT..AGTCC CTG.CC.TCG G.....T..T AAG..T..T.
BR12	T..G..A..C CGT..AGTCC CTG.TC.TCA G.....C..T AAG..T..T.
BZ1	T..G..G..C TGT..AGTCC CTG.TC.TCA G.....C..C AAG..T..T.
VK	T..G..A..C CGT..AGTTC CTG.TC.TCA G.....C..T AAG..T..T.
QB	T..A..G..C TGT..AGTCC CTG.CC.CCG G.....T..T AAG..T..T.
M11	A..G..A..A GAC..GTGCA GTC.TC.AGT T.....T..C TCC..T..T.
MX1	A..G..A..A GAC..GTGCA GCT.GT.GAT T.....T..C TCC..C..A.

	35103520353035403550
Consensus	GTTTTMSDAC CAAYRMKAAR AARACBTTYT YYRRNGGRCC GTTCMGAGAG
TW18ACA.. ...TACT..A ..G..T..T. CCGAG..G..A.....
HL4-9ACG.. ...TACT..A ..G..T..C. CCGAG..G..A.....
BR12ACT.. ...TACT..A ..G..T..C. CTGAG..G..A.....
BZ1ACT.. ...TACT..A ..G..C..T. CCGAA..G..A.....
VKACT.. ...TACT..G ..G..T..T. CTGAG..G..A.....
QBACG.. ...TACT..A ..G..T..T. CCGAG..G..A.....
M11CGT.. ...CAAG..G ..A..G..T. CTAGT..A..C.....
MX1AGA.. ...CGAG..G ..G..C..T. TCGAC..G..C.....

	35603570358035903600
Consensus	TCGTGCGGHA AGCACTACTW TWYKGGCGTW GAYGTYACWC CYTTYTACAT
TW18T.A ..TCT.....A ..T..T..T. .C..C.....
HL4-9T.A ..TCT.....A ..T..T..T. .C..T.....
BR12T.A ..TCT.....A ..T..T..T. .C..T.....
BZ1T.A ..TCT.....A ..T..T..T. .C..T.....
VKT.A ..TCT.....A ..T..T..T. .C..T.....
QBC.A ..TCT.....A ..T..T..T. .C..T.....
M11A.T .TTG.....T ..C..C..A. .T..C.....
MX1A.T .ATG.....T ..C..C..A. .T..C.....

	36103620363036403650
Consensus	ACGYCRCCGY ATAGTGAVYC CYDCYGATYT MATAYTGGTT TTGAAYMASM
TW18	...T.A...TCC. .TG.C...T. A...C.....TA.CC
HL4-9	...T.A...TAT. .TA.C...T. A...T.....TA.CC
BR12	...T.G...CGT. .TG.C...T. A...T.....TA.CC
BZ1	...C.G...CGT. .TG.C...T. A...T.....TA.CC
VK	...T.G...TGT. .TG.C...T. A...T.....TA.CC
QB	...T.A...TGT. .TG.C...T. A...C.....TA.CC
M11	...T.G...TGT. .CT.C...C. C...C.....CC.GA
MX1	...C.A...TGT. .CT.T...C. C...C.....CC.GA

	36603670368036903700
Consensus	TRTATCGKTG GGCCACRATT GAYGGCGTAT GGGATCCTAG RGYMYATYCY
TW18	.A.....G..G... ..T..... G.CCC..C.C
HL4-9	.A.....G..G... ..T..... G.CCC..T.C
BR12	.A.....G..G... ..C..... A.CCC..T.T
BZ1	.A.....G..G... ..C..... G.CCC..T.C
VK	.A.....G..G... ..C..... A.CAC..T.C
QB	.A.....G..A... ..C..... G.CCC..T.T
M11	.G.....T..A... ..C..... G.TAT..C.T
MX1	.G.....T..G... ..T..... G.TAT..C.T

	37103720373037403750
Consensus	GTRTAYMYCA AGTAYMGWMR KYWSCTKCK RAWMWKCTSC RVMGDAATRY
TW18	..G..CCT..TC.TAA GTTG..G..T A.ACAG..G. AAC.T...AC
HL4-9	..A..CCT..TC.TAA GTTG..G..T A.ACAG..G. AAC.T...AC
BR12	..A..TCT..TC.TAA GCTG..G..T A.ACAG..G. AGC.T...AC
BZ1	..G..CCT..TC.TAA GTTG..G..T A.ACAG..G. AGC.T...AC
VK	..A..CCT..TC.TAA GCTG..G..T A.ACAG..G. AGC.T...AC
QB	..G..CCT..TC.TAA GTTG..G..T A.ACAG..G. AAC.T...AC
M11	..A..CAC..TA.ACG TTAC..T..G G.AATT..C. GGA.G...GT
MX1	..A..CAC..CA.ACG TCTG..G..G G.TATT..C. GCA.A...GT

	37603770378037903800
Consensus	YRTRCCTGAT GGWTAYGGTG ATGGNGCYCT CGTCGGATCK GTCYTDAYCA
TW18	TA.A..... ..T..C....G..C..GC.G.T..
HL4-9	TA.A..... ..T..C....A..C..G ...T.G.T..
BR12	TA.A..... ..T..C....T..T..G ...T.G.T..
BZ1	TA.A..... ..T..C....C..C..GC.G.T..
VK	TA.A..... ..T..C....C..T..G ...T.G.T..
QB	TA.A..... ..T..C....T..C..GC.A.T..
M11	CG.G..... ..A..C....T..C..T ...T.A.T..
MX1	CG.A..... ..A..T....T..C..TC.T.C..

	3810 3820 3830 3840 3850
Consensus	RTCCHTTTCGC RRAAAAYCGC GGD TGGRTYC GGYRYGTDCC GRTGATHAYD
TW18	A...T..... GA....C... ..G...A.C. ..TAC..A.. .G....C.CG
HL4-9	A...T..... GA....C... ..G...A.C. ..TAC..A.. .G....C.TA
BR12	A...T..... GA....C... ..G...A.C. ..TAT..T.. .G....C.TA
BZ1	A...A..... GA....C... ..G...A.C. ..TAT..A.. .G....C.TT
VK	A...C..... GA....C... ..A...A.C. ..TAT..T.. .G....C.TA
QB	A...T..... GA....C... ..G...A.C. ..TAC..A.. .G....T.CG
M11	G...T..... AG....T... ..T...G.T. ..CGT..G.. .A....T.TA
MX1	G...T..... AG....T... ..T...G.T. ..CGT..G.. .A....A.TT

	3860 3870 3880 3890 3900
Consensus	GACMABAVRA RRGACCGAGW DCGYRHYGAR YNDGGDTCRT ATCTYTAYGA
TW18	...C.T.CA. GG.....A G..CGCT..G TTA..G..G.T..C..
HL4-9	...C.C.CA. GG.....A G..CGTT..A TCG..G..G.T..C..
BR12	...C.T.CA. GG.....A A..CACT..G TCA..A..G.C..C..
BZ1	...C.C.CA. GG.....A G..CACT..G CCG..G..A.C..T..
VK	...C.T.CA. GG.....A A..CACC..G TCG..A..G.C..C..
QB	...C.T.CA. GG.....A G..CGCT..G TTG..G..G.C..C..
M11	...A.G.GG. AA.....T T..TGAC..A TAT..T..G.C..C..
MX1	...A.G.AG. AA.....T G..TGAC..G CGT..T..A.C..T..

	3910 3920 3930 3940 3950
Consensus	NCTHTKBTCK YKNYRBYWVY YSGAADGTRA CRRTGRGTTS CCBYTHARSG
TW18	C..C.TC..G CGTTGTCTCC CG...A..A. .GA..G...G ..TC.A.GG.
HL4-9	C..C.TT..G CGTTGTTTCT CG...G..A. .GA..G...G ..TT.A.GG.
BR12	T..T.TC..G CGCTGCTTCT CG...A..A. .GA..G...G ..TC.A.GG.
BZ1	T..C.TC..T CGCTGTTTCT CG...A..A. .GA..G...G ..CC.A.GG.
VK	T..T.TC..G CGCTATTTCT CG...A..A. .GA..G...G ..TC.A.GG.
QB	C..C.TC..G CGTTGTCTCT CG...A..A. .GA..G...G ..TC.T.GG.
M11	G..A.GG..G TTGCAGCAAC TC...T..G. .AG..A...C ..CT.T.AC.
MX1	A..A.GG..G TTACAGCAGC TC...T..G. .AG..A...C ..GT.C.AC.

	3960 3970 3980 3990 4000
Consensus	GKYCRYYGRK YKKYGRTHCY RHNDATSBVN YYBBYRYHKA YSVRCWTATH
TW18	.TC.GTC.AG TTGC.A.T.T GTAA..CTGT CTGCTGTCG. TCAA.T...C
HL4-9	.TC.GTC.AG TTGC.A.T.T GTAG..CTAT CTGCCATTG. TCAA.T...C
BR12	.TC.GTC.AG TTGC.A.C.T GTAT..CCAC TTGCCATAG. TCAG.T...A
BZ1	.TC.GTC.AG TTGC.A.T.T ATAT..CGGT TTGCCGTTG. CCAA.T...A
VK	.TC.GTC.AG TTGC.A.T.T ATAT..CCGC TCGCTATCG. TCAG.T...T
QB	.TC.ATC.GG TTGC.A.T.T GCGG..CTAT TTGCCATCG. TCAG.T...C
M11	.GT.GCT.GT CGTT.G.T.C ACTG..GGCA CTCTCGCTT. CGCA.A---C
MX1	.GT.GCT.GT TGTT.G.A.C AACG..GGAG TTTGCACTT. CCGA.A---C

	4010 4020 4030 4040 4050
Consensus	YGWRRRMGBD WWYCTACVRH DRTMAGYRRD KCBRYHRGBR MRTWYGAYRT
TW18	T.TAGGA.CA ATC....AAA GA.A..TAGA T.TACTG.CA AG.TC..TA.
HL4-9	T.TAGGA.TA ATC....AAA GA.A..TAGA T.TACCG.TA AG.TC..TA.
BR12	T.TAAGA.TA ATC....AAA AA.A..CAGA T.TACTG.TA AG.TT..TG.
BZ1	T.TAAGA.CA ATC....AAA AG.A..CAGA T.TACTG.GA AA.TT..TA.
VK	T.TAAGA.TA ATC....AAA AA.A..CAGA T.TACTG.TA AG.TC..TA.
QB	T.TAGGA.TA ATC....GAA GA.A..CAGG T.TACCG.CA AA.TC..TA.
M11	C.AGAAC.GT TAC....CGT TA.C..TGAT G.CGTAA.TG CG.TT..CA.
MX1	C.AGAGC.GG TTT....CGC TA.C..TGAT T.GGTTG.CG CA.AC..CA.

	4060 4070 4080 4090 4100
Consensus	MVWRTRKATM SCGTGCRGTA GYCGTGTYCT GGCWCCCTAC GGGGWYTTCC
TW18	ACAG.AT..C G.....A... .C.....T.. ...A..... ..TC....
HL4-9	ACAG.AT..C G.....G... .C.....T.. ...A..... ..TT....
BR12	ACAG.AT..C G.....A... .C.....T.. ...A..... ..TC....
BZ1	ACAG.AT..C G.....A... .C.....T.. ...A..... ..TT....
VK	ACAG.AT..C G.....A... .C.....T.. ...A..... ..TT....
QB	ACAG.AT..C G.....A... .C.....T.. ...A..... ..TC....
M11	CATG.GG..A C.....A... .T.....C.. ...T..... ..AT....
MX1	CGTA.GG..A C.....A... .T.....C.. ...T..... ..AC....

STOP 4

	4110 4120 4130 4140 4150
Consensus	RGAGGCACGA AGGYTVYRYC YYWAMAAYRR GGYRKMRCTT GGGAGGGSKS
TW18	A.-..... ..T.ACAT. TCT.C.-CGA ..TG TAACGC
HL4-9	A.-..... ..T.ACAT. TCT.C.-CGA ..TA TAACGC
BR12	A.-..... ..T.GCGT. TCT.C.-CGA ..CG TAACGC
BZ1	A.-..... ..T.ATGC. TCT.C.-CGA ..CA TAACGC
VK	A.-..... ..T.GCGT. TCT.C.-CGA ..CG TAACGC
QB	A.-..... ..T.GCGT. TCT.C.-CGA ..CG TAACGC
M11	G..... ..C.CTAT. CTA.A..TGG .. TAG CG... ..GTG
MX1	G..... ..T.CTAT. CTA.A.-TAG .. TAG CA... ..GTG

	4160 4170 4180 4190 4200
Consensus	YWHTAYRSMS CCTRRGTTRK SAATMMWTWA WCWMAMYTWC TCDWWMGAGW
TW18	CAA..TGGCG ...AA-..GT G...AAA.T. T.AC.AT.A. ..TTAC...T
HL4-9	CAA..TGGCG ...AA-..GT G...AAA.T. T.AC.AT.A. ..TTAC...T
BR12	TAT..TAGCG ...AG-..GT G...CAA.T. T.AC.AC.A. ..TTAC...T
BZ1	CAC..TGGCG ...AA-..GT G...AAA.T. T.AC.AT.A. ..TTTA...T
VK	TAT..TGGCG ...GG-..GT G...AAA.T. T.AC.AC.A. ..TTAC...T
QB	CAA..TGGCG ...AA-..GT G...AAA.T. T.AC.AT.A. ..TTAC...T
M11	CAT..TGCAC ...AG...AG C...ACT.A. A.TA.CC.T. ..AAAA...A
MX1	CTA..CGCAC ...AG...AG C...-ACT.A. A.TA.CC.T. ..GAAA...A

	4210 4220 4230
Consensus	GAGWTGRRGG MTCTGCTTWG CCCTCWCTCC TCCCA
TW18	...A-.AG.. A.....T.T....
HL4-9	...A-.AG.. A.....A.T....
BR12	...A-.AG.. A.....T.T....
BZ1	...A-.AG.. A.....T.T....
VK	...A-.AG.. A.....T.T....
QB	...A-.GG.. A.....-- -----
M11	...T-.AA.. C.....T.A....
MX1	...T-.AG.. C.....T.A....

Appendix A3b
Group III QB-like

Alignment: Allolevivivirus Group III QB-like.

	10 20 30 40 50
Consensus	GGGGGACCCC CCTTAGGGGG TCWCCTCACA CAGCAGTAYT TCACTRAGTA
TW18A.....C.....A.....
HL4-9A.....T.....G.....
BR12A.....T.....G.....
BZ1T.....T.....G.....
VKA.....T.....G.....
QB	-----G.....
	ORF1

	60 70 80 90 100
Consensus	TRAGAGGACA TATGCGCTAAA TTACRCRGTG GTCTGCGTTT CGGAGCCGAT
TW18	.G.....A.....
HL4-9	.G.....A.....
BR12	.A.....A.....
BZ1	.A.....G.....
VK	.G.....A.....
QB	.A.....G.....

	110 120 130 140 150
Consensus	AATGAAATTC TTAATGATTT TCARGAGCTC TGGTTTCCAG AYCTCTTTAT
TW18G.....C.....
HL4-9A.....T.....
BR12G.....T.....
BZ1G.....T.....
VKG.....T.....
QBG.....C.....

	160 170 180 190 200
Consensus	CGAATCTTCC GACACGCAYC CGTGGTACAC ACTGAARGGT CGTGTGTTGA
TW18T.....A.....
HL4-9T.....A.....
BR12T.....A.....
BZ1C.....A.....
VKT.....G.....
QBT.....G.....

	210	220	230	240	250
Consensus	AYGCYCA	YHT DGATGAY	CGY YTACCYA	ATG TARGYGG	TCG YCARRTMMGG
TW18	.C..T..TC.	T.....T..C	C....T....	..G.T.....	C..GG.CA..
HL4-9	.T..C..TC.	T.....T..T	C....T....	..G.C.....	T..GA.CA..
BR12	.C..T..CA.	G.....C..C	T....T....	..A.T.....	C..AG.AC..
BZ1	.C..T..TT.	A.....T..T	C....T....	..G.C.....	T..AA.AA..
VK	.C..T..TA.	T.....C..T	T....C....	..A.C.....	C..AG.AA..
QB	.C..C..CC.	T.....T..T	C....T....	..G.C.....	C..GG.AA..

	260	270	280	290	300
Consensus	CGYACWCCRC	AYCGYGY	YAC YGTYCCG	ATT GCCTCTW	CAG GCCTTCG
TW18	..T..T..A.	.T..T.TT..	T..T.....T...
HL4-9	..T..T..A.	.T..T.CT..	T..T.....T...
BR12	..T..T..A.	.T..C.CT..	T..T.....A...
BZ1	..T..T..G.	.C..T.CT..	C..C.....T...
VK	..T..A..A.	.T..C.TT..	T..C.....T...
QB	..C..T..A.	.T..C.TC..	C..T.....T...

	310	320	330	340	350
Consensus	RGTAACAACC	GTTTCAGT	ATG ATCCYRC	AGC ACTRTCG	TTC TTRTTGA
TW18	G.....CG....	...A.....	..A.....
HL4-9	G.....CA....	...A.....	..A.....
BR12	A.....CG....	...A.....	..A.....
BZ1	G.....TA....	...G.....	..A.....
VK	A.....CG....	...A.....	..G.....
QB	G.....CG....	...A.....	..A.....

	360	370	380	390	400
Consensus	CYCGTGTTGA	YTGGGATT	TC GGTRATG	GYG AYAGTG	CGRA CCTTGTC
TW18	.T.....	T.....	...A...C.	.T.....A.
HL4-9	.T.....	T.....	...A...C.	.T.....A.
BR12	.T.....	T.....	...A...C.	.C.....G.
BZ1	.C.....	T.....	...G...T.	.T.....G.
VK	.T.....	T.....	...A...C.	.C.....G.
QB	.T.....	C.....	...A...C.	.T.....A.

	410	420	430	440	450
Consensus	AAWGA	CTTYB TKTTY	CGYAC YTYYG	CRCCT AAGGA	KTTYG ATTTYT
TW18	..T.....	TT .G..T..C.	T.CC..A...G..C.C.....
HL4-9	..T.....	TT .G..T..T.	T.TC..G...G..C.C.....
BR12	..A.....	TG .T..C..T.	C.TT..A...T..C.C.....
BZ1	..T.....	TT .G..C..C.	T.TC..A...G..C.C.....
VK	..A.....	CG .G..T..C.	C.TT..A...T..C.C.....
QB	..T.....	TC .G..T..C.	C.TT..A...G..T.T.....

	460 470 480 490 500
Consensus	YTCYTTRGYT CCTCGYTATA CYCAGGCCTT CTCCGCBTTT AATGCCAART
TW18	T..T..A.T.T.... .T.....T...G.
HL4-9	T..C..A.T.T.... .T.....T...G.
BR12	T..C..A.C.T.... .C.....C...A.
BZ1	C..C..G.C.C.... .C.....T...G.
VK	C..C..A.T.T.... .C.....T...A.
QB	C..C..A.T.T.... .T.....G...G.

	510 520 530 540 550
Consensus	ATGGCRYTAT GATCGGYGAA GGGCTCGAGA CTATAAAATA TCTCGGGCTK
TW18AC...C...G
HL4-9AC...C...T
BR12GT...T...T
BZ1AC...C...T
VKGT...T...T
QBAC...C...T

	560 570 580 590 600
Consensus	TTACTGCGCA GACTGCGTGA GGGTTWCCGY GCTGTTAARC RYGGCGATTT
TW18A...CA. GC.....
HL4-9A...CG. GT.....
BR12A...CG. GC.....
BZ1T...TG. AC.....
VKA...TG. GC.....
QBA...CG. GT.....

	610 620 630 640 650
Consensus	ACGYGCTCTT CGYAGGGTTA TCCAGTCYTA CCAYAADGGT AARTGGAARC
TW18	...T..... ..C.....T.. ...T..T... ..G.....G.
HL4-9	...T..... ..T.....C.. ...T..G... ..G.....A.
BR12	...T..... ..C.....T.. ...T..A... ..A.....A.
BZ1	...C..... ..T.....T.. ...C..G... ..A.....A.
VK	...T..... ..T.....T.. ...T..A... ..A.....A.
QB	...T..... ..T.....C.. ...T..T... ..G.....A.

	660 670 680 690 700
Consensus	CRRCTACTGC TGGTAATCTC TGGCTTGAAT TYCGTTATGG YCTTATGCCY
TW18	.GA.....T..... .T..... C.....T
HL4-9	.AA.....C..... .C..... T.....T
BR12	.GA.....T..... .T..... C.....T
BZ1	.GA.....T..... .T..... C.....C
VK	.GA.....T..... .T..... C.....T
QB	.GG.....T..... .T..... C.....T

	710720730740750
Consensus	CTCTTTYTAYG ACATCARAGA YGTYATGYTA GAYTGGCARA RBCGYCAYGA
TW18C..T.A... C..C...T.. ..T.....G. AC..T..T..
HL4-9C..T.A... C..C...T.. ..C.....G. AG..C..T..
BR12C..C.G... T..T...T.. ..T.....G. GC..C..T..
BZ1C..T.G... T..C...T.. ..T.....G. AT..T..C..
VKC..T.G... T..C...C.. ..T.....A. AC..C..T..
QBT..T.G... T..C...T.. ..C.....G. AC..T..T..

	760770780790800
Consensus	TARRATTCAA CGCCTCCTTC GRTTTTCDGT YGGTCAYGGY GAGGAYTRYG
TW18	..AA..... ..G.....T.. C.....C..TC.GC.
HL4-9	..GG..... ..G.....G.. T.....C..TT.AT.
BR12	..AG..... ..A.....A.. T.....T..TC.AC.
BZ1	..AG..... ..G.....T.. C.....T..CT.AT.
VK	..AG..... ..G.....T.. T.....C..CC.AT.
QB	..AG..... ..G.....T.. T.....C..CT.AC.

	810820830840850
Consensus	TTGTCRARTT YGACARYYTR TAYCCHGCKY TWKCYTACTT TAAACTRARA
TW18G.A.. C....ACT.A ..T..A..CG .TG.T.....G.A.
HL4-9A.A.. C....GCT.A ..T..A..CG .TT.T.....A.G.
BR12A.G.. T....ACT.G ..T..C..CT .AG.C.....G.A.
BZ1G.A.. T....ACT.A ..T..C..CG .AG.T.....G.A.
VKA.A.. C....ACT.A ..C..C..TT .AG.C.....G.A.
QBG.A.. C....ATC.G ..C..T..CG .TG.T.....G.A.

	860870880890900
Consensus	GGKGAGATTA CWCTCGAACG CCGTCATCGT CAYGGYATAT CYTACGCKAA
TW18	..T..... .A..... ..T..T.... .T.....T..
HL4-9	..G..... .A..... ..T..T.... .T.....T..
BR12	..T..... .T..... ..T..C.... .T.....G..
BZ1	..T..... .T..... ..T..T.... .T.....T..
VK	..T..... .A..... ..C..T.... .C.....G..
QB	..G..... .A..... ..T..C.... .T.....T..

	910920930940950
Consensus	YCGCGRRGGA TATGCTGTTT TCGAYAACGG TTCCCTTCGG CCTGTGTCCG
TW18	T....AA... ..T.....
HL4-9	T....AG... ..T.....
BR12	T....AA... ..T.....
BZ1	T....AA... ..T.....
VK	T....GG... ..C.....
QB	C....AA... ..C.....

	960 970 980 990 1000
Consensus	ATTGGAAGGA RCTTGCCRY Y GCRTTCATCA ATCCKCAHGA AGTTGCHTGG
TW18 G.....ACC ..A..... ..G..T..T...
HL4-9 A.....ACT ..A..... ..T..T..A...
BR12 A.....GTC ..G..... ..T..C..T...
BZ1 G.....ACC ..G..... ..T..A..C...
VK G.....GTC ..G..... ..T..C..T...
QB G.....ACT ..A..... ..G..T..T...

	1010 1020 1030 1040 1050
Consensus	GAGYTAATC CYTACAGCTT YGTTGYYGAT TGGTTYTTGA AYGTYGGYGA
TW18	...C..... .C..... C....TT... ..C..... .T..C..T..
HL4-9	...T..... .C..... C....TT... ..T..... .T..C..T..
BR12	...T..... .T..... C....CC... ..T..... .C..T..C..
BZ1	...T..... .C..... T....TT... ..T..... .C..C..T..
VK	...T..... .T..... T....TT... ..C..... .C..C..C..
QB	...T..... .C..... C....TT... ..C..... .T..T..T..

	1060 1070 1080 1090 1100
Consensus	CATACTYGCT CARCARGGTC ARCTATATCA TAATATCGAK ATYGTAGAYG
TW18T... ..A..G.... .G..... ..C.....T ..T.....C.
HL4-9T... ..G..G.... .G..... ..T.....T ..T.....C.
BR12T... ..G..A.... .G..... ..G.....G ..C.....T.
BZ1C... ..A..A.... .A..... ..T.....T ..T.....C.
VKT... ..G..A.... .A..... ..T.....T ..T.....T.
QBT... ..A..A.... .G..... ..T.....T ..T.....C.

	1110 1120 1130 1140 1150
Consensus	GYTTYGACAG ACGTGACATH CGRYTCAAAT CNTTYACYAT WAAAGGTGAR
TW18	.T..T.....C ..GC..... .C..C..T.. A.....A
HL4-9	.T..T.....T ..AC..... .T..C..T.. A.....A
BR12	.T..C.....A ..GT..... .A..C..T.. T.....A
BZ1	.C..C.....A ..GT..... .G..T..C.. A.....G
VK	.T..T.....A ..GT..... .G..T..C.. T.....A
QB	.C..T.....C ..GC..... .T..C..C.. A.....A

	1160 1170 1180 1190 1200
Consensus	CGAAATGGRC RRCCTGTAA CGTWTCTGCB RRCYTRTCTR CTRTYGATWY
TW18G. GG..... ..T.....T GA.C.G...G ..G.C...TT
HL4-9G. GA..... ..T.....C GA.C.G...G ..G.C...TC
BR12A. AG..... ..A.....G GA.T.A...A ..A.C...AC
BZ1A. AG..... ..T.....G GA.C.G...G ..G.T...AC
VKA. AG..... ..A.....G GA.C.A...G ..A.C...AC
QBG. GG..... ..T.....T AG.C.G...G ..G.C...TT

	12101220123012401250
Consensus	ATTTTAYAGY CGACTYCATR CKASCARBMT TCCGTTTCGCT ACACTHGATC
TW18C..CC...A .G.G..GTC.T....
HL4-9C..CC...A .G.C..AGC.C....
BR12T..TT...G .T.G..ATA.A....
BZ1T..TC...A .G.G..GTA.A....
VKT..TT...G .T.G..ACA.T....
QBC..CC...A .G.G..ATC.A....

	12601270128012901300
Consensus	TYGATACTAC CTTTAGTTTCG TWTAAACACG TYCTHGATAG TRTCKYTTTA
TW18	.C..... .A..... .C..T.... .A..TT....
HL4-9	.C..... .A..... .T..T.... .A..TT....
BR12	.C..... .T..... .C..A.... .A..TT....
BZ1	.C..... .T..... .T..C.... .A..TT....
VK	.C..... .T..... .T..A.... .G..GC....
QB	.T..... .T..... .T..T.... .A..TT....

STOP 1 ORF2/3

	13101320133013401350
Consensus	TTAACCCAAC GCRTWAAGCG YTGAAATCTT TGGGTCAATT TGATCATGGC
TW18A.T.... T.....-...
HL4-9A.T.... T.....-...
BR12A.T.... T.....-...
BZ1A.T.... C.....
VKA.T.... T.....-...
QBG.A.... T.....-...

	13601370138013901400
Consensus	AAAATTAGAG ACTGTTACTT TARGTAACAT YGGGAAAGAT GGAMARMAAA
TW18A..... C..... ..C.AC...
HL4-9A..... C..... ..A.AC...
BR12A..... C..... ..A.GC...
BZ1A..... T..... ..C.AA...
VKA..... C..... ..A.AC...
QBG..... C..... ..A.AC...

	14101420143014401450
Consensus	CTCTGRWCCT CAAYCCRCGT GGGGTAAATC CCACYAACGG YGTTGCCKCG
TW18GT... ..C..G... ..T..... T.....G..
HL4-9GT... ..T..G... ..T..... T.....G..
BR12GT... ..C..G... ..T..... T.....T..
BZ1AA... ..C..A... ..C..... T.....T..
VKGT... ..T..G... ..T..... T.....T..
QBGT... ..T..G... ..T..... C.....T..

	14601470148014901500
Consensus	CTTTCRSAAG CGGGTGCAGT TCCTGCRYTG GAGAAGCGTG TTACCRTTTC
TW18AC...
HL4-9AC...
BR12AG...
BZ1AG...
VKGG...
QBAC...

	15101520153015401550
Consensus	GGTATCHCAG CCTTCYCGYA ATCGYAAGAA CTACAAGGTY CARGTTAARA
TW18C...
HL4-9A...
BR12T...
BZ1T...
VKT...
QBT...

	15601570158015901600
Consensus	TCCARAACCC RACCGCTTGY HCTGCAAACG GTTCTTGTGA CCCATCCGTT
TW18G..... G.....C A.....
HL4-9G..... G.....T A.....
BR12G..... G.....T C.....
BZ1A..... A.....T T.....
VKA..... G.....T T.....
QBG..... G.....C A.....

	16101620163016401650
Consensus	ACTCGCCAGG CATATGCTGA CGTGACYTTY TCGTTCACGC AGTATAGYAC
TW18
HL4-9
BR12
BZ1
VK
QB

	16601670168016901700
Consensus	CGATGAGGAA CGAGCTTTTG TTCGYACAGA GCTTRYGCT CTGCTCGCTR
TW18
HL4-9
BR12
BZ1
VK
QB

STOP 2

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      ....|....|....|....|....|....|....|....|....|....|
      1710      1720      1730      1740      1750
Consensus RTCCTCTGYT GATYGATGCT ATYGATCRRRC TGAAYCCAGC RTATGAACR
TW18      G.....C. ...C..... ..T....AG. ....T..... G.....A
HL4-9     G.....T. ...T..... ..C....AG. ....C..... G.....A
BR12      G.....C. ...C..... ..C....GA. ....C..... G.....A
BZ1       A.....C. ...C..... ..C....AG. ....C..... G.....G
VK        G.....C. ...C..... ..C....GA. ....T..... A.....A
QB        G.....C. ...C..... ..T....AG. ....C..... G.....A

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      ....|....|....|....|....|....|....|....|....|....|
      1760      1770      1780      1790      1800
Consensus YTGCTCATTG CCGGTGGTGG CTCAGGGKMW AAMCCCGATY CGGTTRWYAT
TW18      C..... ..TCA ..C.....T ....ATC--
HL4-9     C..... ..TCT ..A.....C ....ATC..
BR12      T..... ..GAA ..C.....C ....GAT--
BZ1       T..... ..GAA ..C.....C ....GAT--
VK        T..... ..GAA ..C.....C ....AAT--
QB        C..... ..TCA ..A.....C ....ATT--

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      ....|....|....|....|....|....|....|....|....|....|
      1810      1820      1830      1840      1850
Consensus TCCGGATCCA CCGATYGATC CGCCGCCAGG GWCAGGYAVN TATACYTGTC
TW18      -..... ..T.... ..A....C.AG ....C....
HL4-9     ..C.... ..A....T.AA ....T....
BR12      -..... ..T.... ..A....T.GT ....T....
BZ1       -..... ..T.... ..T....T.GC ....C....
VK        -..... ..T.... ..A....T.CC ....T....
QB        -..... ..T.... ..A....T.AG ....C....

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      ....|....|....|....|....|....|....|....|....|....|
      1860      1870      1880      1890      1900
Consensus CYTTCGCAAT TTGGTCHYTR GARGAGGTTT AYGARCTCC TACYADKRAY
TW18      .C..... ..CC.A ..G..... .C..G..... ...T.GGA.C
HL4-9     .T..... ..CC.A ..G..... .C..G..... ...T.AGA.C
BR12      .C..... ..TC.G ..A..... .T..G..... ...C.TTG.T
BZ1       .T..... ..AT.A ..G..... .T..G..... ...C.GTG.C
VK        .T..... ..TC.A ..G..... .T..A..... ...T.GTG.C
QB        .C..... ..CC.A ..G..... .C..G..... ...T.AGA.C

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      ....|....|....|....|....|....|....|....|....|....|
      1910      1920      1930      1940      1950
Consensus CGACCGTGGC CTATCTAYHR YGCTRTYGAA CTCYVRYCTC GYRADTTTGA
TW18      .....CAA T...G.C... ...CGGC... .CA.G.....
HL4-9     .....TAA C...A.C... ...CAGC... .CA.G.....
BR12      .....TCG T...G.T... ...TCGT... .CA.G.....
BZ1       .....TTA T...G.C... ...CAAC... .TG.T.....
VK        .....TCA T...G.T... ...CCGC... .CG.T.....
QB        .....TAA T...G.T... ...CAGC... .CG.A.....

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	19601970198019902000
Consensus	TGTTGCCCTY RAWGATCTYT TGGGYAAYAC WRADTGGCGH GAYTGGGATT
TW18C G.A.....T.C..T.. AA.T.....C ..C.....
HL4-9T A.A.....T.C..T.. AA.A.....C ..C.....
BR12T G.T.....C.T..C.. TA.A.....T ..C.....
BZ1T G.T.....C.T..C.. TA.A.....A ..C.....
VKC G.T.....C.T..C.. TG.A.....T ..C.....
QBC A.A.....T.C..T.. AA.G.....T ..T.....

	20102020203020402050
Consensus	CWCGGCTBAG DTATACCACG TTYCGCGGTT GCCGTRGCAA YGTTTATATT
TW18	.A.....G.. T..... ..C..... ..G.... T.....
HL4-9	.A.....T.. T..... ..T..... ..A.... C.....
BR12	.T.....T.. G..... ..C..... ..G.... T.....
BZ1	.A.....T.. T..... ..C..... ..G.... T.....
VK	.A.....C.. A..... ..C..... ..G.... C.....
QB	.T.....T.. T..... ..C..... ..G.... T.....

	20602070208020902100
Consensus	GACCTYGATG CGWCTTATCT YGCTACTGAY CARGCDATGC KYGATCAGAA
TW18C..... ..A..... T.....T ..G..T.... GT.....
HL4-9T.... ..T..... T.....T ..G..T.... GT.....
BR12T.... ..A..... C.....T ..A..T.... TC.....
BZ1T.... ..A..... T.....C ..A..G.... TT.....
VKT.... ..A..... C.....C ..A..A.... TT.....
QBT.... ..A..... T.....T ..G..T.... GT.....

	21102120213021402150
Consensus	GTAYGATATT CGCRMGGCA ARARACCYGG TGCYTTYGGY AAHRTYGARC
TW18	...C..... ...GAG.... .A.A...T.. ...T..C..T ..CA.T..G.
HL4-9	...C..... ...GAA.... .G.A...C.. ...T..C..T ..CG.T..G.
BR12	...C..... ...ACG.... .A.G...T.. ...C..T..C ..AA.T..A.
BZ1	...C..... ...ACG.... .G.A...C.. ...C..C..T ..CA.C..A.
VK	...C..... ...ACA.... .A.G...T.. ...C..C..T ..TA.T..A.
QB	...T..... ...GAG.... .G.A...T.. ...T..C..T ..CA.T..G.

	21602170218021902200
Consensus	RATTYATYTA TCTTAAGTCG ATAAATGCHT ATTGYTCTCT YAGCGATATT
TW18	G...C..T..T.C..... T.....
HL4-9	G...C..T..T.C..... T.....
BR12	A...T..T..A.T..... C.....
BZ1	A...C..C..C.T..... T.....
VK	A...T..T..A.T..... T.....
QB	G...C..T..T.C..... T.....

	2210 2220 2230 2240 2250
Consensus	GCGGCCTATC ACGCCGAYGG YGTRATAGTT GGCTTTTGGC GCGATCCATC
TW18T.. T..G.....
HL4-9T.. T..G.....
BR12C.. C..A.....
BZ1T.. T..G.....
VKT.. C..A.....
QBT.. C..G.....

	2260 2270 2280 2290 2300
Consensus	YAGTGGTGGT GCCATACCGT TTGACTTCAC DVARTTTGAT AAGACTAAAT
TW18	T..... TA.G.....
HL4-9	T..... TA.G.....
BR12	T..... TC.G.....
BZ1	T..... AG.G.....
VK	T..... GG.A.....
QB	C..... TA.G.....

STOP 3

	2310 2320 2330 2340 2350
Consensus	GTCCDATTCA AGCYGTGATA GTCGTTCCCTC GTGCTTAGTA ACTAAGGATG
TW18T.....C.....
HL4-9T.....C.....
BR12T.....C.....
BZ1A.....T.....
VKG.....C.....
QBT.....C.....

ORF4

	2360 2370 2380 2390 2400
Consensus	AAATGCATGT CTAAGACAGC ATCYTCGCRT AACTCTCTYA GCGCACAATT
TW18T...A.....T.....
HL4-9T...A.....T.....
BR12T...A.....T.....
BZ1C...G.....C.....
VKT...A.....T.....
QBT...G.....C.....

	2410 2420 2430 2440 2450
Consensus	GCGCCGAGCY GCGAACACAA GAATTGAGGT HGAAGGTAAC CTCGCACTKT
TW18T.....A.....T.....
HL4-9T.....C.....T.....
BR12T.....T.....T.....
BZ1C.....C.....T.....
VKT.....C.....G.....
QBC.....T.....T.....

	24602470248024902500
Consensus	CCATYGCCAA CGAYYTAMTR TTGGCHTATG GTCARTCGCC ATTTARYTCY
TW18T..... ..TT..C.AT....A.....AC..T
HL4-9C..... ..TC..C.GT....A.....AC..C
BR12T..... ..CT..A.GA.....A.....GT..T
BZ1T..... ..TT..A.GT....A.....AT..C
VKT..... ..TT..A.GA.....A.....GT..C
QBT..... ..TT..C.GC.....G.....AC..T

	25102520253025402550
Consensus	GARKCYGAGT GTATTTTCAYT CRGYCCGARA TTYGACGRRA CCCC GGAYVA
TW18	..AG.T.... ..C. .A.T....G. ..C....GG.TG.
HL4-9	..AG.T.... ..C. .A.T....G. ..T....GG.TC.
BR12	..GT.T.... ..C. .A.C....A. ..C....GA.TA.
BZ1	..GT.C.... ..C. .G.T....A. ..C....AA.CA.
VK	..GT.T.... ..C. .A.C....A. ..C....GA.TA.
QB	..GG.T.... ..T. .A.C....G. ..C....GG.TG.

	25602570258025902600
Consensus	CTTTAGGATA AATTATCTTA AAGCCGAGRT CATGTCGAAG TATGACGAYT
TW18G.T.
HL4-9G.T.
BR12A.T.
BZ1A.T.
VKA.T.
QBA.C.

	26102620263026402650
Consensus	TCAGCCTAGG TATYGATACC GAAGCTGYTG CMTGGRARAA GTTYCTRGCA
TW18T..... ..C.. .C...G.A.. ...C..G...
HL4-9T..... ..C.. .C...G.A.. ...C..G...
BR12T..... ..C.. .A...A.G.. ...T..A...
BZ1T..... ..C.. .A...G.G.. ...C..A...
VKC..... ..C.. .A...G.A.. ...C..A...
QBT..... ..T.. .C...G.G.. ...C..G...

	26602670268026902700
Consensus	GCAGAGGCTG AATGTGCTTT AACGAACGCT CGYCTCTATA GRCCTRACTA
TW18T..... ..G...G....
HL4-9T..... ..G...G....
BR12T..... ..A...A....
BZ1T..... ..G...A....
VKC..... ..A...A....
QBT..... ..G...G....

	2710 2720 2730 2740 2750
Consensus	CAGTGAGGAT TTCAATTTCT CACTGGGYGA GTCRTGTMTW CACATGGCTC
TW18C.. ...A...A.T
HL4-9C.. ...A...A.T
BR12T.. ...G...C.T
BZ1C.. ...A...A.T
VKT.. ...A...C.T
QBC.. ...A...A.A

	2760 2770 2780 2790 2800
Consensus	GYMGAAAAAT AGYYAAGYTR ATAGGAGAYG YKCCGTCCGT TGAGGGTATG
TW18	.TA..... ..CC...T.AC. CT.....
HL4-9	.CA..... ..TC...C.AT. CT.....
BR12	.CC..... ..TT...T.GT. CT.....
BZ1	.TA..... ..TT...C.AC. CG.....
VK	.CC..... ..TT...T.AC. CG.....
QB	.TA..... ..CC...C.AT. TT.....

	2810 2820 2830 2840 2850
Consensus	YTGCGYCAYT GCCGDTTTTC YGGYGGTGCY ACAACAACGA ATAACCGTTC
TW18	T...T..C.T.... C..T....C
HL4-9	T...T..C.A.... T..C....C
BR12	T...T..T.A.... T..C....C
BZ1	C...T..C.T.... C..T....T
VK	T...C..T.G.... T..C....C
QB	T...T..C.A.... T..C....T

	2860 2870 2880 2890 2900
Consensus	RYAYGGYCAT CCGTCCTTCA AGTTTGCCTTCCRCAAGCG TGTACGCCTC
TW18	GT.C..T...
HL4-9	GC.C..C...
BR12	AT.T..T...
BZ1	AT.C..T...
VK	AT.C..T...
QB	GT.C..T...

	2910 2920 2930 2940 2950
Consensus	GGGCTTTGAA RTAYGTTYTR GCTCTYAGAG CYTCWACRCA YTTTCGATATC
TW18A..T...T.AT.... .T..T..A.. T.....
HL4-9A..T...T.GT.... .T..T..A.. T.....
BR12A..T...T.GT.... .T..A..G.. T.....
BZ1G..T...C.GT.... .C..T..G.. C.....
VKA..C...T.GC.... .C..A..A.. T.....
QBG..T...T.AC.... .T..T..A.. T.....

	2960 2970 2980 2990 3000
Consensus	AGRRTTTCTG ATATTAGCCC TTTTAATAAA GCAGTTACTG TACCWAAGAA
TW18	..AA..... ..T.....
HL4-9	..GG..... ..T.....
BR12	..AG..... ..A.....
BZ1	..AG..... ..A.....
VK	..AG..... ..A.....
QB	..AA..... ..T.....

	3010 3020 3030 3040 3050
Consensus	CAGTAARACM GATCGYTGTA TYGCTATCGA RCCYGGYTGG AATATGTTTT
TW18A..CT.... .T..... G..T..C... ..
HL4-9A..CT.... .T..... A..C..C... ..
BR12A..CC.... .T..... A..C..T... ..
BZ1A..CC.... .C..... A..C..T... ..
VKG..CC.... .T..... A..C..T... ..
QBG..AT.... .T..... A..T..T... ..

	3060 3070 3080 3090 3100
Consensus	TYCAAYTRGG YATCGGTGGC ATYYTACGYG AYCGGYTGCG TTGYTGGGGT
TW18	.T...C.A.. C..... ..TC....C. .T...T.... ..C.....
HL4-9	.T...C.A.. C..... ..CC....C. .T...T.... ..C.....
BR12	.C...T.A.. T..... ..TT....T. .T...T.... ..T.....
BZ1	.T...C.A.. T..... ..CT....C. .C...C.... ..C.....
VK	.C...T.A.. T..... ..CC....C. .T...T.... ..C.....
QB	.C...C.G.. T..... ..TC....C. .T...T.... ..C.....

	3110 3120 3130 3140 3150
Consensus	ATCGAYCTGA AYGATCAGAC GATAAAYCAG CRYCGCGCTC ACGARGGCTC
TW18T.... .T..... ..T.... .GC..... ..A.....
HL4-9T.... .C..... ..T.... .AT..... ..A.....
BR12C.... .T..... ..C.... .AC..... ..G.....
BZ1T.... .T..... ..C.... .GT..... ..A.....
VKC.... .T..... ..C.... .GC..... ..A.....
QBT.... .T..... ..T.... .GC..... ..A.....

	3160 3170 3180 3190 3200
Consensus	YGTTACTAAT RAYTTAGCRA CRGTTGATCT CTCAGCRGCA AGCGATTCTA
TW18	C..... A.T....A. .G..... ..G.... ..
HL4-9	C..... A.T....A. .G..... ..G.... ..
BR12	C..... G.T....A. .G..... ..G.... ..
BZ1	T..... A.C....A. .A..... ..G.... ..
VK	C..... G.T....G. .G..... ..A.... ..
QB	C..... A.C....A. .G..... ..G.... ..

	3210 3220 3230 3240 3250
Consensus	TATCTCTTGC YCTCTGYGAG CTCTTRYTRC CYCCAGGBTG GTTTGARGTY
TW18 T.....T... ..AC.G. .C.....C..G..C
HL4-9 T.....T... ..AT.G. .C.....T..G..T
BR12 T.....T... ..AT.A. .T.....T..G..C
BZ1 T.....C... ..GT.G. .T.....G..A..C
VK T.....C... ..AT.G. .T.....T..G..C
QB C.....T... ..AT.G. .C.....C..G..T

	3260 3270 3280 3290 3300
Consensus	CTTAYGGAYC TYMGATCACC WAARGGGCRR TTGCCTRACG GTAGTGTYRT
TW18T...C. .CA..... T..A....GGG...TG.
HL4-9T...T. .CA..... A..A....GGG...TG.
BR12C...T. .CA..... T..G....AAG...TA.
BZ1T...C. .TC..... T..G....GAA...CG.
VKT...C. .CA..... T..G....GAG...CA.
QBT...C. .CA..... T..G....GAG...TG.

	3310 3320 3330 3340 3350
Consensus	TAYYTAYGAG AARATWTCYT CBATGGGTAA YGGWTACACA TTCGARCTYG
TW18	..CC..T... ..G..T..C. .T..... C..A.....G..T.
HL4-9	..CC..T... ..A..A..C. .G..... C..A.....G..T.
BR12	..CC..T... ..A..T..T. .C..... T..T.....G..C.
BZ1	..CT..T... ..G..T..T. .C..... C..T.....A..C.
VK	..TC..C... ..G..T..T. .C..... T..T.....G..T.
QB	..CC..C... ..G..T..T. .T..... C..T.....G..C.

	3360 3370 3380 3390 3400
Consensus	AGTCGCTTAT YTTYGCTTCT CTYGCYCGTT CCGTTTGTGA GATACTGRAC
TW18 C..T..... ..T..T...G..
HL4-9 C..T..... ..T..T...G..
BR12 T..T..... ..C..C...G..
BZ1 T..T..... ..T..T...G..
VK C..C..... ..T..T...A..
QB T..T..... ..C..T...G..

	3410 3420 3430 3440 3450
Consensus	TTAGACTCRT CTGAGGTCAC TGTYTACGGA GAYGAYATTA TYTTRCCRTC
TW18A.C..... ..C..T... ..C..A..G..
HL4-9G.T..... ..C..T... ..T..A..G..
BR12A.T..... ..T..C... ..T..G..A..
BZ1A.C..... ..C..C... ..T..G..G..
VKA.T..... ..T..C... ..T..G..A..
QBG.T..... ..C..T... ..T..A..G..

	34603470348034903500
Consensus	CYGTGCAGTY CCTGCYCTYC RGGAAGTYTT YAAGTATGTT GGTTTTACDA
TW18	.T.....CC..T. G.....C.. T.....A.
HL4-9	.T.....CC..T. G.....T.. T.....G.
BR12	.C.....CT..T. A.....C.. T.....T.
BZ1	.T.....CT..T. A.....C.. C.....T.
VK	.C.....TT..T. A.....C.. T.....T.
QB	.T.....CC..C. G.....T.. T.....G.

	35103520353035403550
Consensus	CCAATACTAA RAAGACYTTY TCYGARGGGC CGTTCAGAGA GTCGTGCGGY
TW18A.....T..T ..C..G....T
HL4-9A.....T..C ..C..G....T
BR12A.....T..C ..T..G....T
BZ1A.....C..T ..C..A....T
VKG.....T..T ..T..G....T
QBA.....T..T ..C..G....C

	35603570358035903600
Consensus	AAGCACTACT ATTCTGGCGT AGATGTTACT CCCTTYTACA TACGYCRCCG
TW18T.....C....T.A...
HL4-9T.....T....T.A...
BR12T.....T....T.G...
BZ1T.....C.G....
VKT.....T.G....
QBT.....T.A...

	36103620363036403650
Consensus	YATAGTGAVY CCTRCCGATT TAATAYTGGT TTTGAATAAC CTATATCGGT
TW18	T.....CC ...G..... ..C....
HL4-9	T.....AT ...A..... ..T....
BR12	C.....GT ...G..... ..T....
BZ1	C.....GT ...G..... ..T....
VK	T.....GT ...G..... ..T....
QB	T.....GT ...G..... ..C....

	36603670368036903700
Consensus	GGGCCACRAT TGAYGGCGTA TGGGATCCTA GRGCMCATYC YGTRTAYCTC
TW18G.. ...T..... .G..C...C. C..G..C...
HL4-9G.. ...T..... .G..C...T. C..A..C...
BR12G.. ...C..... .A..C...T. T..A..T...
BZ1G.. ...C..... .G..C...T. C..G..C...
VKG.. ...C..... .A..A...T. C..A..C...
QBA.. ...C..... .G..C...T. T..G..C...

	37103720373037403750
Consensus	AAGTATCGTA AGYTGCTGCC TAAACAGCTG CARCGTAATA CTATACCTGA
TW18T.....A.....
HL4-9T.....A.....
BR12C.....G.....
BZ1T.....G.....
VKC.....G.....
QBT.....A.....

	37603770378037903800
Consensus	TGGTTACGGT GATGGNGCYC TCGTCGGATC GGTCYTRATC AATCCHTTTCG
TW18G..C.....C.G...T....
HL4-9A..C.....T.G...T....
BR12T..T.....T.G...T....
BZ1C..C.....C.G...A....
VKC..T.....T.G...C....
QBT..C.....C.A...T....

	38103820383038403850
Consensus	CGAAAAACCG CGGRTGGATC CGGTAYGTWC CGGTGATYAY DGACCAYACA
TW18G.....C..A.....C.C G.....T...
HL4-9G.....C..A.....C.T A.....C...
BR12G.....T..T.....C.T A.....T...
BZ1G.....T..A.....C.T T.....C...
VKA.....T..T.....C.T A.....T...
QBG.....C..A.....T.C G.....T...

	38603870388038903900
Consensus	AGGGACCGAG ARCGCRYYGA RYYRGGRTCR TATCTYTAYG AYCTYTTYTC
TW18G...GCT.. GTTA..G..GT..C..C..C..C..
HL4-9G...GTT.. ATCG..G..GT..C..C..C..T..
BR12A...ACT.. GTCA..A..GC..C..T..T..C..
BZ1G...ACT.. GCCG..G..AC..T..T..C..C..
VKA...ACC.. GTCG..A..GC..C..T..T..C..
QBG...GCT.. GTTG..G..GC..C..C..C..C..

	39103920393039403950
Consensus	KCGYTRYTC YCGGAARGTA ACGATGGGTT GCCYYTWAGG GGTC CRTCGR
TW18	G..T.GTC.. C.....A... ..TC.A... ..G...A
HL4-9	G..T.GTT.. T.....G... ..TT.A... ..G...A
BR12	G..C.GCT.. T.....A... ..TC.A... ..G...A
BZ1	T..C.GTT.. T.....A... ..CC.A... ..G...A
VK	G..C.ATT.. T.....A... ..TC.A... ..G...A
QB	G..T.GTC.. T.....A... ..TC.T... ..A...G

	3960 3970 3980 3990 4000
Consensus	GTTGCGATYC TRYRDATCBR YYYGCRYTHG AYCARTTAT HTGTARGAGY
TW18T. .GTAA...TG TCT..TG.C. .T..A..... C....G...C
HL4-9T. .GTAG...TA TCT..CA.T. .T..A..... C....G...T
BR12C. .GTAT...CA CTT..CA.A. .T..G..... A....A...T
BZ1T. .ATAT...GG TTT..CG.T. .C..A..... A....A...C
VKT. .ATAT...CG CTC..TA.C. .T..G..... T....A...T
QBT. .GCGG...TA TTT..CA.C. .T..G..... C....G...T

	4010 4020 4030 4040 4050
Consensus	AATCCTACRA ARRTAAGYAG RTCTACYGGB AARTTYGATR TACAGTATAT
TW18A. .GA....T.. A.....T..C ..G..C...A
HL4-9A. .GA....T.. A.....C..T ..G..C...A
BR12A. .AA....C.. A.....T..T ..G..T...G
BZ1A. .AG....C.. A.....T..G ..A..T...A
VKA. .AA....C.. A.....T..T ..G..C...A
QBG. .GA....C.. G.....C..C ..A..C...A

	4060 4070 4080 4090 4100
Consensus	CGCGTGCRGT AGCCGTGTTC TGGCACCCTA CGGGGTYTTC CAGGGCACGA
TW18A..
HL4-9G..
BR12A..
BZ1A..
VKA..
QBA..

STOP 4

	4110 4120 4130 4140 4150
Consensus	AGGTTRYRYC TCTACACGAG GYR TAA CCTG GGAGGGCGCY AHTATRGCGC
TW18ACAT.TG..... .C .A...G....
HL4-9ACAT.TA..... .C .A...G....
BR12GCGT.CG..... .T .T...A....
BZ1ATGC.CA..... .C .C...G....
VKGCGT.CG..... .T .T...G....
QBGCGT.CG..... .C .A...G....

	4160 4170 4180 4190 4200
Consensus	CTRRTTGTGA ATMAATTATC ACAAYTACTC TTWMGAGTGA GAGRGGGATC
TW18	..AA..... .A..... .T..... .AC..... .A.....
HL4-9	..AA..... .A..... .T..... .AC..... .A.....
BR12	..AG..... .C..... .C..... .AC..... .A.....
BZ1	..AA..... .A..... .T..... .TA..... .A.....
VK	..GG..... .A..... .C..... .AC..... .A.....
QB	..AA..... .A..... .T..... .AC..... .G.....

	4210 4220
Consensus	TGCTTWGCCC TCTCTCCTCC CA
TW18T..... ..
HL4-9A..... ..
BR12T..... ..
BZ1T..... ..
VKT..... ..
QB----- ----- --

Appendix A4
Group IV

Alignment: Allolevivivirus Group IV.

	10 20 30 40 50
Consensus	GGGGGTAGGS SSSRTAAAGG GGGCCTGCCY TCAMCGCACT ACAGAGGAGA
BR1	-.....C CCCG...-- .T...C.....
BR8	-.....C CCCG...-- .T...C.....
HB-P22	-.....C CCCG...-- .T...C.....
HB-P24	-.....C CCCA...-- .T...A.....
SPG GGG.....C...C.....
NL95C CCCA...-- .T...A.....
FI	-----
	ORF1

	60 70 80 90 100
Consensus	ATCTATGCCW RCCCTWCCRA KAGGWCTTCG CTTYGGWTCG AAHGGCGARR
BR1A A...T..G. G...T.... .C...A... ..T....GG
BR8A A...T..G. G...T.... .C...A... ..T....GG
HB-P22A G...A..A. G...T.... .T...T... ..T....AA
HB-P24A A...T..G. G...T.... .C...A... ..C....GG
SPA A...T..G. G...T.... .C...A... ..T....AG
NL95A A...T..G. G...A.... .C...A... ..T....AA
FIT A...T..A. T...T.... .T...A... ..A....AA

	110 120 130 140 150
Consensus	TYBTWAAYGA YTTCRRSGMG CTYTGGTTTC CRRAGCKCSW HDCHKYMGAAH
BR1	.CT.A..T.. C...GAG.C. ..C..... .AG...G.CA TA.CGTC..T
BR8	.CT.A..C.. C...GAG.C. ..C..... .AG...G.CA TA.CGTA..T
HB-P22	.CT.A..C.. T...AAC.A. ..C..... .GG...T.GT CT.ATCC..A
HB-P24	.CG.T..C.. C...AAC.C. ..C..... .AG...G.GA AA.CTTC..C
SP	.TC.T..T.. C...GAG.C. ..C..... .GG...G.CA TA.CGTA..T
NL95	.CG.A..T.. T...AAC.C. ..C..... .AG...G.GA AG.ATTC..T
FI	.CT.A..T.. T...AGC.C. ..T..... .GA...G.GT CT.TTTC..C

	160 170 180 190 200
Consensus	YYNVRSHWHG GVHSCTRYRM VCTCWCKGGY TAYDTNAVTR RYCWGYYBKV
BR1	CTGAGCAAT. .GAC..GCAA G...A.T..T ..TA.C.C.A AC.T.CCTGG
BR8	CTAAGCAAT. .GAC..GCAA G...A.T..T ..TA.C.C.A AC.T.CCGGA
HB-P22	TTGAACCTTA. .AAC..ATAA C...A.G..T ..CA.T.G.A AC.T.CCGGG
HB-P24	TCCGGGCTT. .ATC..ACGA A...A.T..T ..CG.A.G.A AT.A.CCGGG
SP	CTAAGCAAT. .GAC..GCAA G...A.T..T ..TA.C.C.A AC.T.CCTGG
NL95	CTCGAGCTC. .CTC..ACAC A...A.T..T ..CG.A.G.A AT.A.CCTGG
FI	TCTCAGCTT. .ACG..ACGA A...T.T..C ..TT.G.A.G GC.A.TTCTC

	210 220 230 240 250
Consensus	VKAYDVNRNN MNVDWSCSBA ATMMDVSVDY BMMWKBBNYK MRDACRCCGY
BR1	CT.CAGTAAT ATATTC.CG. ..AAGGGAGT CACTGTTGCT CGT..G...T
BR8	CT.CAGCAAC ACATTC.CG. ..AAGGGGGT CACTGTTGCT CGT..G...T
HB-P22	AT.CGAAGTG AAGGAG.GC. ..AAAGGCTC TCATGTGCTT CGA..A...T
HB-P24	AT.CGAGACA CGCATG.GT. ..CCTAGCAT GCATTGTATT CGT..A...C
SP	CT.CAGTGAC ATATTC.CT. ..AAAGGAGT CACTGCTGCT CGT..G...T
NL95	CT.TACGACG CGCATG.GT. ..CCTCGCAT GCATTGCGTT CGT..A...C
FI	GG.CTACGGC AGG-----..CCTGCCTT GCAAGTCTCG AAG..A...C

	260 270 280 290 300
Consensus	AYMGDRSWAC NGTVCCNGTW AACCATYTYG GWTAYMGGCC NGTYACVACD
BR1	.CA.AAGT.. A..G..T..TC.T. .T..CC.... C..T..G..T
BR8	.CA.AAGT.. T..G..T..TC.C. .T..CC.... A..T..A..T
HB-P22	.CC.TAGT.. C..G..A..AC.T. .A..CA.... G..T..G..A
HB-P24	.CA.AAGT.. G..G..A..TC.T. .T..CA.... T..T..G..T
SP	.CA.AAGT.. A..G..C..TC.T. .T..CA.... A..T..G..T
NL95	.CA.AAGT.. T..A..T..TT.T. .T..TA.... A..T..G..T
FI	.TC.GGCA.. A..C..G..TC.T. .T..CC.... T..C..C..G

	310 320 330 340 350
Consensus	GTWGAGTAYR HTCCHVWBGG NACBTWCRTV CGCCTNGAYG GSVMYGTDNR
BR1	..T.....CA T...CGAC.. C..C.A.G.GA..T. .GCAC..AAA
BR8	..T.....CA T...CGAC.. C..T.A.G.GT..T. .GCAC..GAA
HB-P22	..A.....TA A...ACTG.. A..T.T.A.CG..T. .GGAT..AAA
HB-P24	..T.....CA C...TAAT.. G..T.T.A.CT..C. .GGAT..GAA
SP	..T.....CA T...CGAC.. A..T.A.G.AC..T. .GCAC..GAA
NL95	..T.....TG T...AAAC.. A..T.T.A.CA..T. .GGAT..AAA
FI	..A.....CG T...CAAT.. T..G.T.G.GT..T. .CACC..TCG

	360 370 380 390 400
Consensus	VWTYDVMGGD GRVKYGGTYA RYGSSWCRST BRRDCTHRVS AAYTWYRTKR
BR1	GT.CGAA..G .GCTT...T. AT.GGT.AG. TGAT..CACG ..T.ACG.GA
BR8	GT.CGAA..G .GCTT...T. AT.GGT.AG. TGAT..CACG ..T.TCG.GA
HB-P22	GT.TTCC..A .GCTT...T. GC.CGT.GC. TAGA..TGAC ..C.ACG.TG
HB-P24	GT.TTCC..T .GAGC...C. GT.GGT.GC. CAAA..TAGC ..C.ATG.TG
SP	AT.TGAA..G .ACTT...T. AT.GGT.AG. TGAT..CACG ..T.TCG.GA
NL95	GT.TTCC..T .GATC...T. GT.GGT.GC. TAAG..AAAC ..C.TCG.TG
FI	CA.CAGC..G .AGTT...T. AT.GCA.AG. GAGG..TGAC ..C.ATA.TG

	410 420 430 440 450
Consensus	TYDVNYTVGC KKCNCAGGSY GGYTTTCGATT ACCARTCGGT AATCGGWCCW
BR1	.CTCGT.A.. TG.T....GT ..C..... ..A..... ..A..T
BR8	.CTCAT.A.. TG.T....GT ..C..... ..G..... ..A..T
HB-P22	.CGGTC.G.. TT.G....CC ..T..... ..G..... ..A..A
HB-P24	.TAACT.G.. GG.C....GT ..C..... ..G..... ..T..A
SP	.CTCAT.A.. TG.T....GT ..C..... ..A..... ..A..T

NL95	.TAACT.A..	GT.A....GC	..C.....G.....A..A
FI	.TAACC.C..	TG.C....GT	..C.....G.....A..A

	460 470 480 490 500
Consensus	AGGTTCTCDK CGCRMTTYKC YGCGTTYAGY ACCAARTATG GTRYBTTRCT
BR1TG ...AC..CT. C.....C..CA.... ..GTC..A..
BR8TG ...AC..CT. C.....C..CA.... ..GTC..G..
HB-P22AT ...AA..CT. C.....T..TA.... ..ACT..G..
HB-P24TG ...AA..CG. T.....C..TG.... ..ACG..A..
SPTG ...GC..CT. C.....T..CA.... ..GTC..A..
NL95GT ...AA..CT. C.....T..TG.... ..GCT..G..
FITT ...AA..TT. C.....T..TA.... ..ACT..A..

	510 520 530 540 550
Consensus	MGGVGAAGGG AGAGARACDC TTARKTATCT TCTCCTSSTS BTTTCGAGRR
BR1	C..G..... ..G..T. ...AG..... ..GC.G C.....AA
BR8	C..A..... ..A..T. ...AG..... ..GC.G C.....AA
HB-P22	C..G..... ..A..A. ...GT..... ..GC.G T.....AG
HB-P24	A..C..... ..A..A. ...GT..... ..GC.G T.....GG
SP	C..A..... ..A..T. ...AG..... ..CG.C G.....AA
NL95	C..G..... ..A..A. ...GT..... ..GC.G T.....AA
FI	C..A..... ..A..G. ...GT..... ..GC.G T.....AA

	560 570 580 590 600
Consensus	TRCGTGAAGG GWWCKYGCY GTWARGCRHG GHGATYTCAA RCGYCTCAGG
BR1	.G..... .TA..GC..C ..A.G..GT. .C...C.... G..C.....
BR8	.G..... .TA..GC..C ..A.G..GT. .C...T.... G..T.....
HB-P22	.A..... .AT..GC..C ..A.G..GT. .A...C.... G..T.....
HB-P24	.G..... .AT..GC..T ..A.A..AC. .T...C.... G..C.....
SP	.G..... .TA..GC..C ..A.G..GT. .C...C.... G..T.....
NL95	.G..... .AT..GT..C ..A.G..GC. .T...C.... G..T.....
FI	.G..... .TT..TC..T ..T.A..GA. .A...C.... A..C.....

	610 620 630 640 650
Consensus	AATSTKRTHH SDASVTWCGA RCCDMBNWCC NHDNVHGGNM RMDRSMVAVS
BR1	...G.GA.CT CT.CG.T... G..GACAT.. TCTAAA..GA GGAAGGC.AG
BR8	...G.GA.CT CT.CG.T... G..GATCT.. CCGAGA..GA AGATGGC.AG
HB-P22	...C.GG.TA GG.GC.A... A..GACGT.. GTGAAT..AC GTCGACA.GC
HB-P24	...C.GA.AC GT.CG.T... G..GCGCT.. GTTCCC..AA AGCGGCA.AG
SP	...G.GA.AT CG.CG.T... G..GAGTA.. ATAAAA..TA AACGAGC.AG
NL95	...C.GA.AC GT.CA.T... A..ACGGT.. ATTTCT..CC GACGGCA.CG
FI	...G.TA.CC GA.CG.T... A..TCGTT.. CAAGCT..GA AGAGGCA.CG

	660 670 680 690 700
Consensus	RDCNDMSTTY WMMVRRDCCT ATYGCGAYVR RMYRYRCRGW AACAAAGGTyr
BR1	AT.TGAG..C TCACAAA... ..C....CAA GCTTAC.G.TTG
BR8	AT.AGAG..C TCACAGA... ..C....CAA ACTTAC.G.TTA
HB-P22	AG.CTCC..T TCAGGGA... ..C....CGA ACTTGC.A-- -----
HB-P24	AG.ATCG..C TCAGAAG... ..C....CAG AACTGC.A-- -----
SP	GG.CGAG..T TCACAGA... ..C....CAA GCTTAC.G.ACG

NL95	AA.GACG..C TCAGAGA... ..C....TCG ACTCGT.A-- -----
FI	AT.GTCG..T AACAAGT... ..T....CAA GCTCGC.A-- -----

	710 720 730 740 750
Consensus	AAGTTARACC RAGTGRNGRY RRKTGGAADR RCAGTAGTGC GAGYRACCTG
BR1A... A....AG.GT AAG.....TA G..... ..TG.....
BR8A... A....AA.GT AAG.....TA G..... ..TG.....
HB-P22	----- --AT.GC AAG.....AG A..... ..CG.....
HB-P24	----- --AT.GC AGT.....AG A..... ..TG.....
SPG... G....AA.GT AAG.....TA G..... ..TG.....
NL95	----- --AC.AC GGG.....AG A..... ..TG.....
FI	----- --GT.GC GAG.....GA A..... ..TA.....

	760 770 780 790 800
Consensus	TGGYTMGART TCCGHTAYGG RYTKATGCCCK TTRTTYTAYG ACATHMARTC
BR1	...T.A..G.T..T.. GC.T....G ..G..C..C.TC.G..
BR8	...C.A..G.T..T.. AC.G....T ..G..T..T.TC.G..
HB-P22	...C.C..A.T..T.. GT.G....G ..G..C..C.AC.A..
HB-P24	...T.A..A.T..C.. GT.G....G ..A..T..C.CC.G..
SP	...T.A..G.T..T.. GC.G....G ..A..C..C.AC.G..
NL95	...T.A..G.A..C.. AC.G....G ..G..T..C.CA.G..
FI	...C.C..A.C..C.. AT.G....G ..A..T..C.TC.G..

	810 820 830 840 850
Consensus	YKTVATGGAR GAYTTCATGC GYRTBCAYAA GARRATCGCR AARHTWCARC
BR1	CG.G....A ..C..... .TG.C..T.. ..AG....G ..AA.A..G.
BR8	CG.C....A ..C..... .TG.C..T.. ..AG....A ..GA.A..G.
HB-P22	TT.A....A ..C..... .TG.T..T.. ..AA....A ..AA.T..G.
HB-P24	CG.A....A ..C..... .TG.C..T.. ..AG....G ..AC.T..G.
SP	CG.C....A ..C..... .TG.T..T.. ..AG....A ..AA.T..G.
NL95	TG.A....G ..C..... .TA.C..T.. ..AG....G ..AT.A..G.
FI	TG.A....A ..T..... .CG.G..C.. ..GA....G ..AA.T..A.

	860 870 880 890 900
Consensus	GRTTYTCDGC HGGRCAYGGB AAGCTNGWDR MGGTNWVBKV KMNDTTYTWY
BR1	.A..T..T.. A..G..C..TG.AGA C...TAGTTC GCGG..T.AC
BR8	.A..C..T.. C..A..C..TG.AGA C...TAGTTC GCGG..T.AC
HB-P22	.A..C..G.. T..G..C..TC.TTG A...GTCCGG GACT..C.AC
HB-P24	.A..C..T.. A..A..C..CA.TTG A...AAAGGG TAAG..C.TC
SP	.G..T..A.. T..A..T..TC.AGA C...TAGTTC GCGG..T.AC
NL95	.A..T..A.. A..A..C..TT.TGA C...TAAGGG TAGG..C.TC
FI	.A..C..T.. C..G..T..GA.AAA A...CTCGGA TATA..T.AT

	910 920 930 940 950
Consensus	CCKRRYVYBY AYTTTCVVBMT YGAGGTYACY GCRGTGTTRC AGCGGCGTCA
BR1	..GGACGTCC .C...GCTC. T....C..C ..A....A.
BR8	..GGACGTCC .C...GCTC. T....C..C ..A....A.
HB-P22	..GGATGTCC .C...GGCC. T....C..C ..A....G.
HB-P24	..TGATCCTC .C...GCTC. T....C..C ..A....G.
SP	..GGACGTCC .T...AGCC. T....C..T ..A....A.

NL95	..TGACCCGC	.C...GCTA.	T.....C..C	..G.....A.
FI	..GAGCACTT	.T...CAGC.	C.....T..C	..A.....A.

	960 970 980 990 1000
Consensus	TCGTTGGGGK GTNRTMTAYC AGGATACTGR TWCBTWBSCM MCYTTYRAYA
BR1T ..CA.A..C.G .T.T.ATG.C A.T..TA.C.
BR8T ..CG.A..C.G .T.T.ATG.C A.T..CA.T.
HB-P22T ..CA.A..C.G .T.T.ATG.C A.T..CG.C.
HB-P24G ..GA.A..C.G .T.G.TCG.C A.C..CA.T.
SPT ..CA.A..C.G .T.T.TTG.C A.T..CA.C.
NL95G ..AA.A..C.G .T.G.TGC.A C.T..CA.T.
FIT ..TA.C..T.A .A.C.TTG.C A.C..TG.C.

	1010 1020 1030 1040 1050
Consensus	ATGGTCRKCT DRTCCCGGTR ARGGAYTGGM AGACRGCGGC KTTWGCAYTC
BR1GT.. AG.....G .A...C...AA..... G..T...C..
BR8GT.. AG.....G .A...C...AA..... G..T...C..
HB-P22GG.. GG.....G .A...C...CA..... G..T...C..
HB-P24GG.. AA.....G .G...T...CA..... T..A...C..
SPGT.. AG.....A .A...C...AA..... G..T...C..
NL95AG.. TA.....G .G...T...CA..... T..A...C..
FIGG.. AA.....G .G...T...CG..... G..A...T..

	1060 1070 1080 1090 1100
Consensus	CTTAAYCCBG CYGARRYTGC NTGGGARNTH ACWCCYTWSA GYTTCGTBGY
BR1T..T. .C..AGT... A....GG.A ..T..T.AC. .C.....T.T
BR8T..C. .T..AGT... G....GA.C ..T..C.AC. .C.....T.T
HB-P22C..G. .T..AAC... A....AT.A ..A..C.AC. .T.....T.C
HB-P24T..C. .C..GAC... T....GT.A ..T..T.AC. .T.....C.C
SPT..C. .C..AGT... G....AG.T ..T..C.AC. .C.....G.T
NL95T..T. .C..AAC... T....GT.A ..T..C.AC. .C.....T.C
FIT..G. .T..AAC... C....GC.T ..T..C.TG. .T.....C.C

	1110 1120 1130 1140 1150
Consensus	RGATTGGTTT GTDAAYGTWG GKGATATGCT HGAGCAGACT SGGMCARCTY
BR1	G..... .A..T..T. .G..... T.....- G..C..G..T
BR8	G..... .A..T..T. .T..... T.....- G..C..G..T
HB-P22	G..... .G..C..T. .T..... C.....- C..A..A..C
HB-P24	G..... .T..T..A. .T..... C.....- C..C..G..C
SP	G..... .A..T..T. .T..... T.....- G..C..G..T
NL95	G..... .T..T..T. .T..... C..... C..C..G..C
FI	A..... .T..C..A. .T..... A.....- C..C..G..C

	1160 1170 1180 1190 1200
Consensus	TATMGKTCHA YGTYGAYGTH GTYGACGGNT TYGAYMGRRA RGACRTRARR
BR1	...A.G--C. C..C..T..C ..T.....T. .C..CC.G.. G...A.A.AA
BR8	...C.G--C. C..C..T..C ..T.....T. .T..CC.G.. G...A.A.GG
HB-P22	...C.G--C. C..C..C..T ..C.....C. .T..TA.A.. G...G.G.AA
HB-P24	...C.--C. T..C..C..T ..C.....G. .T..TA.A.. G...G.G.AG
SP	...C.G--C. C..C..T..C ..T.....T. .C..CC.G.. A...A.A.AA

NL95
FI

...C.G..A. C..T..C..A ..C.....G. .C..TA.G.. G...G.G.AG
...C.T--T. C..C..T..A ..T.....A. .C..TA.G.. G...G.G.AG

	12101220123012401250
Consensus	CTYMRDWSCR TRTCVGTRCG YGTBHTDRSD RVYGDYRBDR CRVMYDYWGC
BR1	..CAAGTC.G .A..C..A.. C..CT.GACT GGC.ACAGTG .GCACGTA..
BR8	..TAAGTC.A .A..C..A.. C..CT.GACT AGT.ACAGTG .ACACGTA..
HB-P22	..CCGTAG.G .A..C..G.. C..GA.TGCG AAC.GTGCAA .GAACA---
HB-P24	..CAAGAG.G .G..G..A.. T..CA.AAGA GAC.ACGCGA .GCACA---
SP	..CAAATC.G .A..A..A.. C..GC.AACG AAC.ACGTTG .GCATGTT..
NL95	..TAGAAG.G .A..A..G.. C..CA.AAGA GAC.ACGCAA .GACTA---
FI	..CAAGTC.G .G..C..A.. C..TA.AGC- GCC.TC--TG .GGACTCA..

	12601270128012901300
Consensus	DHVBTTYVNN YTVNSWVRRG CDVDDYTGTY GCATRGTTWY TAYTCGCGCG
BR1	TAAG..TACT C.ACGTCAA. .ACGTT..T.A...AT ..T.....
BR8	TAGT..TACT C.ACGTCAA. .ACGTT..T.A...AT ..T.....
HB-P22	ACAG..TGTC C.AAGACGA. .TAGTT..T.A...TC ..T.....
HB-P24	GTCT..CAGC T.AGCTGGA. .GGTGT..T.A...TC ..T.....
SP	TAGC..TCAG C.GCGACAA. .AAAAC..T.A...AT ..C.....
NL95	TAGC..CAAT T.ATCTGGA. .AAAGT..T.A...TC ..T.....
FI	TACT..CGAA C.CCGAAGG. .TGAGC..C.G...TC ..T.....

	13101320133013401350
Consensus	TRCATACCGT YGCGTTYCCG CAAATYTCAC CACAAMTCGA YRCTGARRTM
BR1	.G..... T.....T...T....C.... TA....GA.A
BR8	.G..... T.....T...T....C.... TA....GA.A
HB-P22	.A..... T.....T...T....C.... TA....GG.A
HB-P24	.G..... T.....T...C....C.... TA....GA.A
SP	.G..... T.....T...T....C.... TA....GA.C
NL95	.G..... T.....C....C....C.... CA....AG.A
FI	.G..... C.....T...C....A.... TG....GG.A

	13601370138013901400
Consensus	CGTAGYKTTA ARCACGTDAT CGAYAGYVTY GCMYTWYTAA CYCARCGYKT
BR1CG... .G.....G.. ...T..TA.C ..AT.AT... .C..G..CG.
BR8CG... .G.....G.. ...T..TA.C ..AT.AT... .C..G..CG.
HB-P22CG... .G.....T.. ...C..TA.C ..CC.TC... .T..G..TT.
HB-P24TG... .A.....A.. ...C..TA.T ..CC.TT... .T..G..TT.
SPCG... .G.....A.. ...T..TA.C ..CC.AT... .C..A..CG.
NL95CT... .G.....T.. ...C..TG.T ..CC.TT... .T..G..TT.
FICG... .G.....A.. ...C..CC.C ..CC.TT... .T..A..CT.

STOP 1

ORF2/3

	14101420143014401450
Consensus	TAARARRCGY TRAATYTTTG GGTCAATTYG ATCATGGCWA AATTRAATMM
BR1	...G---.C .G..-C....T.A.A...CA
BR8	...G---.C .G..-C....T.A.A...CA
HB-P22	...A---.C .A...C....T.A.A...CA
HB-P24	...A.GA..C .A--T....C.A.G...AA
SP	...G---.T .G..-C....T.A.A...CA
NL95	...A.AG..C .A...C....C.A.G...AA
FI	...G---.T .A...C....C.T.G...AC

	1460 1470 1480 1490 1500
Consensus	GGTWACDCTT WCCRRDHTHG GWAAGRMNGS NRATMAGACT WTRACWCTWA
BR1	...A..T... T..AAAA.C. .A...AAT.G GG..C..... T.A..T..T.
BR8	...A..T... T..AAAC.T. .A...AAC.G GG..C..... T.A..T..T.
HB-P22	...A..A... A..AAAT.A. .T...GCG.G CG..C..... T.A..T..T.
HB-P24	...T..G... A..GGTA.T. .T...GCT.G AG..C..... T.A..T..T.
SP	...A..T... T..AAAA.C. .A...AAT.G GG..C..... T.A..T..T.
NL95	...T..G... A..GGTA.T. .T...GCT.G AA..C..... T.A..T..T.
FI	...A..T... A..AAGC.T. .T...GAA.C TA..A..... A.G..A..A.

	1510 1520 1530 1540 1550
Consensus	CDCCDCGYGG SGTWAAAYCCB ACKAACGGVG TGGCRWCRCT VTCYGAAGCT
BR1	.A..G..C.. G..A..C..G ..G.....C.GT.G.. G..T.....
BR8	.A..G..C.. G..A..C..G ..G.....C.GT.G.. G..T.....
HB-P22	.T..A..T.. G..A..T..C ..T.....A.GT.G.. G..C.....
HB-P24	.A..G..T.. G..A..T..C ..T.....C.GT.G.. A..T.....
SP	.A..G..C.. G..A..C..G ..G.....C.GT.G.. A..T.....
NL95	.A..G..T.. G..A..C..T ..T.....C.GT.G.. A..T.....
FI	.G..T..C.. C..T..T..T ..G.....G.AA.A.. C..C.....

	1560 1570 1580 1590 1600
Consensus	GGHGCTGTDC CGGCNYTVGA RAARCGMGTR ACTGTGTCAG TYGCKCAGCC
BR1	..A.....T.TT.G.. A..G..C..GT..G.....
BR8	..A.....T.TT.G.. A..G..C..GC..G.....
HB-P22	..T.....A.TC.C.. G..G..A..AT..T.....
HB-P24	..A.....T.GT.G.. G..G..A..AT..T.....
SP	..A.....T.AT.A.. G..G..C..AT..G.....
NL95	..A.....T.TT.G.. G..G..A..AT..T.....
FI	..C.....G.CT.G.. G..A..C..GC..G.....

	1610 1620 1630 1640 1650
Consensus	HTCYCGKAAY CGTAAGAAYT WYAARRTYCA RATTAARCTY CARAACCCGA
BR1	T..C..T..CC. TT..AG.T.. G.....A..C ..A.....
BR8	C..C..T..CC. TT..AG.T.. G.....A..C ..A.....
HB-P22	A..C..T..CC. AT..AG.T.. G.....G..C ..A.....
HB-P24	A..T..T..CC. AT..GA.C.. G.....A..C ..G.....
SP	A..T..G..CC. TT..AG.T.. G.....A..C ..A.....
NL95	A..T..T..CC. AT..AG.T.. G.....A..C ..G.....
FI	A..T..T..TT. TC..GG.C.. A.....G..T ..G.....

	1660 1670 1680 1690 1700
Consensus	CTGCATGCAC GARGGAYGCR TGYGACCCWT CTGTGACGCG WTCYGCKTTC
BR1A..C..A ..T.....A. A..T..T...
BR8A..C..A ..T.....A. A..T..T...
HB-P22A..C..A ..T.....A. T..C..G...
HB-P24A..C..A ..C.....T. T..C..G...
SPG..C..A ..T.....A. A..T..T...

NL95A...C..A	..C.....T.	T..C.-G...
FIA...T..G	..T.....A.	A..T..T...

	1710 1720 1730 1740 1750
Consensus	KCRCGAYSTA ACGCTKTCGT TCACGTCRTA TTCWACYGAS SNDGARCGYG
BR1	G.A-..CG..G....G.. ...T..C..C GAA..A..T.
BR8	G.A-..TG..G....A.. ...T..T..C GCA..A..T.
HB-P22	G.G-..CC..T....G.. ...T..T..C GCT..A..T.
HB-P24	G.G-..CC..G....G.. ...T..T..C GTT..A..T.
SP	G.A-..CG..G....G.. ...T..C..C GAG..A..T.
NL95	T.G...CG..G....G.. ...A..C..G CGT..G..C.
FI	G.A-..CC..G....G.. ...T..T..C GAA..A..T.

	1760 1770 1780 1790 1800
Consensus	CGCTRRTTTCG YACTGARYTR GCRGCTCTVY TSVMRGAYVM HYTGATYRYC
BR1AA.... C.....GT.G ..A.....GT .GCAG..CAA TC....CAT.
BR8AA.... C.....GT.A ..G.....GT .GCAG..TCC CT....TGT.
HB-P22GA.... C.....AT.G ..G.....GT .GCAA..TCC TC....TGT.
HB-P24AG.... C.....AC.G ..A.....GT .GAAG..TGA TT....TGT.
SPGA.... C.....AT.G ..A.....AC .GGCG..TCC AC....TGT.
NL95AA.... C.....AT.A ..G.....GT .GAAG..TGA TC....TGT.
FIGA.... T.....GT.A ..G.....CC .CGCG..TCC TT....TAC.

STOP 2

	1810 1820 1830 1840 1850
Consensus	GATGCDATYG AYAAAYCTKAA YCCWGCNTAC TGAG CGRCGT TAYTGGYWRS
BR1A..C. .T..T..G.. T..T..T...A.... ..C...TAGC
BR8G..T. .C..C..G.. T..T..C...G.... ..C...TAGC
HB-P22A..C. .T..T..G.. C..T..A...G.... ..T...CAGC
HB-P24T..T. .C..T..T.. T..T..A...A.... ..C...CAGC
SPT..T. .C..T..G.. C..A..C...G.... ..C...TAGC
NL95A..C. .T..T..G.. T..T..A...A.... ..C...CAGC
FIT..C. .T..T..G.. T..T..G...G.... ..T...TTAG

	1860 1870 1880 1890 1900
Consensus	CTCGYCNRGY GGYGRGRWTA AHCCMYCMDW NCCWGRCGTY CCGGWTVKTC
BR1T.CG.C ..T.G.GA.. .C..CT.AAT G..A.A...TA.CT..
BR8T.CG.C ..T.G.GA.. .C..CT.AAA G..A.A...TA.CT..
HB-P22T.TA.C ..T.G.GA.. .C..A--AA G..A.A...TA.CG..
HB-P24C.GG.T ..C.G.AA.. .T..C--AA C..T.G...CA.AG..
SPT.CG.C ..T.G.GA.. .T..CT.CGA T..A.A...CT.GT..
NL95C.GG.T ..C.G.AA.. .T..C--TA C..T.G...CA.AG..
FIT.AG.C ..T.A.GT.. .A..CC.AAT A..T.A...CA.GT..

	1910 1920 1930 1940 1950
Consensus	CRRRCGTIYAA RCCGCCHGRC GGWACDGGSH SSTWTMVSTG YCCSTTYDSY
BR1	.AGA...C.. A.....T.A. ..A..G..GC GC.A.ACG.. C..C..CTCC
BR8	.AGA...C.. A.....C.A. ..A..G..GC GC.A.ACG.. C..C..CGCT
HB-P22	.AGA...C.. A.....A.G. ..A..G..CA GC.A.CGC.. T..G..TACC
HB-P24	.GAA...C.. G.....T.G. ..T..A..CA CC.A.CGG.. C..G..CGCC
SP	.AGA...C.. A.....A.A. ..T..G..GC GC.A.AAG.. C..C..CGCC
NL95	.GAA...C.. G.....T.G. ..T..T..CA CC.A.CGG.. C..G..CGCC
FI	.GAG...T.. A.....T.G. ..A..G..CT CG.T.ACG.. C..G..CAGC

	1960	1970	1980	1990	2000
Consensus	TGYTAYCGCC	KYGRTRVWHT	HWWCRMGGHV	GSBMARRASG	GWKYYYSYGM
BR1	..T..C....	TC.G.AGTA.	CTA.GA..AG	.GTA.GG.C.	.TTCTCCT.A
BR8	..T..C....	TC.G.AGTA.	TTA.GA..TG	.GTA.GG.C.	.TTCTCCT.A
HB-P22	..T..C....	TC.G.AATA.	CAT.GA..TA	.GGC.AA.C.	.TTCTCCT.A
HB-P24	..C..C....	GT.G.GAAC.	CAT.AC..AG	.CTA.GG.C.	.AGCTTGT.C
SP	..T..C....	TC.G.AGTA.	TTA.GA..TC	.GTA.GG.G.	.TTCTCCT.A
NL95	..C..C....	GT.G.GAAT.	AAT.AC..AG	.CTA.GG.C.	.AGCTTGC.C
FI	..T..T....	TC.A.ACTA.	TAT.GA..CG	.GCA.GG.C.	.TGTCCCT.A

	2010	2020	2030	2040	2050
Consensus	BMTHTAYGMV	HDKGGMVVBG	ARGYCHHVG	YRHKTTYGAK	TAYGCKSTYG
BR1	CA.C..C.AA	AGG..AGAC.	.A.T.TCG..	TACT..C..T	..T..TC.C.
BR8	CA.T..C.AA	AGG..AGAC.	.A.T.TCG..	TACT..C..T	..C..TC.C.
HB-P22	CA.C..T.CC	AGG..AGAC.	.G.T.CAG..	TATG..C..T	..T..TC.C.
HB-P24	GC.T..T.CG	CTT..CAGT.	.G.C.ATA..	TGAG..C..T	..C..TC.T.
SP	CA.T..T.AA	AGG..AGAC.	.A.T.TCA..	CACT..C..T	..C..TC.C.
NL95	GC.A..C.CG	TGT..CAGT.	.G.C.CTA..	TGAG..C..G	..C..GC.C.
FI	TC.C..T.AA	CAG..ACCG.	.G.T.ACC..	TACG..T..T	..T..TG.C.

	2060	2070	2080	2090	2100
Consensus	AGGATTTYCT	YGGKAACRHN	WYWTGGCGHA	ACTGGGAYVR	BCGMYTRTCR
BR1T..	T..G...ACA	AAT.....T.TCA	G..AT.G..A
BR8C..	T..G...ACG	AAT.....T.TCA	G..AT.G..A
HB-P22C..	T..G...ACC	AAT.....T.TCA	G..AC.A..A
HB-P24C..	C..G...GTG	TTC.....C.CGG	T..CT.A..G
SPC..	T..G...ACG	AAT.....T.TCA	G..AT.A..A
NL95C..	C..G...GAG	TTC.....A.TGG	T..AT.A..A
FIC..	C..T...ACT	AAT.....T.TAG	C..AC.A..A

	2110	2120	2130	2140	2150
Consensus	DMWTAYGAYW	TMGVBAMYCD	YCGTCGTTGY	MHGGAAYG	GKTAYRTHGA
BR1	TCA..T..CT	.A.CT.AT.G	T.....C	C.T..C..C.	.G..CA.T..
BR8	TCA..T..CT	.A.CT.AT.G	T.....C	C.T..C..C.	.G..CA.T..
HB-P22	AAT..T..TA	.A.CT.AT.G	T.....C	C.A..C..T.	.T..CG.T..
HB-P24	ACA..C..TA	.C.AC.CT.A	C.....T	C.C..T..C.	.G..CG.T..
SP	GAT..T..TA	.A.CT.AT.G	T.....C	C.T..C..T.	.G..CA.C..
NL95	AAA..T..TA	.C.AG.CT.A	T.....T	C.C..T..C.	.G..CG.T..
FI	AAT..T..TA	.C.GC.AC.T	C.....C	A.A..T..C.	.G..TG.A..

	2160	2170	2180	2190	2200
Consensus	YYTDRAYGCH	WCHRYVATGC	ARWCHGAYRV	NTDYGTDYTR	TCNGGYVVB
BR1	CT.AG.T..C	A.TGTG....	.GT.A..CGA	A.TC..TC.G	..T..CCGT.
BR8	CT.AA.C..T	A.CGCG....	.GT.A..CGA	A.GT..TC.G	..T..CCGC.
HB-P22	CC.GG.C..A	A.TGCG....	.GA.C..CAG	C.TT..GC.A	..C..TAAG.
HB-P24	CC.AG.C..A	A.CATG....	.GT.A..CGC	T.AT..AT.G	..T..CGCT.
SP	CC.AG.T..A	A.CGCC....	.GT.T..TGA	T.TC..AT.G	..A..CCGC.

NL95
FI

CC.TG.C..C T.CGTA.... .AT.A..CGA G.AT..AT.G ..T..TGCT.
TC.TG.C..T A.AGCG.... .GT.C..CAG C.AT..GC.G ..G..CAAG.

	2210 2220 2230 2240 2250
Consensus	AYSVNGTDSK NAAGRKVMWR YYNCCMRGYR YHTTYGVMKC MMYIARRTAY
BR1	.TCCA..GCG C...GTCAAG TTT..CG.CG CT..C.GCT. CATC.AG..T
BR8	.CCCG..ACG T...GTCAAG TTT..CG.CG CA..T.GCT. CATC.AG..T
HB-P22	.TCCA..GCG G...GTCAAG TTT..AG.TG CC..T.GCG. CCTC.AG..T
HB-P24	.TGAT..TGT C...ATGCAA CCA..CA.TA TC..C.ACT. CCCT.GG..C
SP	.CGGC..GCG A...GTCAAG TTT..CG.CG CC..C.GCT. AATC.AG..T
NL95	.CGAT..TGT T...ATGCAA CCG..CG.TA CC..C.ACT. CCCT.GG..C
FI	.TCGA..ACG C...GGACTA CCC..CG.TA TC..T.CAT. ACCC.GA..C

	2260 2270 2280 2290 2300
Consensus	YWYYTBVAHM TMMWRGRYGR YRYCTRKKTD GAYYTDKCCG MGGTRACRGC
BR1	CTCT.GA.CA .TCAA.GC.A TGC..GGT.G ..CT.AT... A...G..A..
BR8	CTCT.GA.CA .TCAA.GT.A TGC..GGT.G ..CT.AT... A...G..A..
HB-P22	CTCC.TA.CA .AAAG.AT.A TGC..GGG.G ..CT.GT... A...A..G..
HB-P24	TACC.CC.TC .AATG.AT.G TAT..ATG.G ..CT.AG... A...A..G..
SP	CTCT.GA.CA .TCAA.GT.A TGC..GGT.A ..CT.AT... A...A..A..
NL95	TACC.CC.TC .TATG.AT.G TAT..ATG.G ..CT.AG... A...A..G..
FI	TATC.TG.AC .ACAA.AT.G CGC..GGG.T ..TC.TG... C...A..G..

	2310 2320 2330 2340 2350
Consensus	VTACCRYTCY TAYGGAATGG TYATYGGYTT CTGGACRGAC TCTAARAGYC
BR1	C....GT..C ..C..... .T..T..T..A...G..C.
BR8	A....GC..C ..C..... .T..T..T..A...A..T.
HB-P22	C....GT..C ..T..... .C..C..T..G...G..C.
HB-P24	A....AT..T ..C..... .C..T..C..A...G..C.
SP	G....GT..C ..C..... .T..T..T..A...G..C.
NL95	A....GT..C ..C..... .C..T..C..A...G..C.
FI	G....GT..C ..T..... .C..C..C..A...G..C.

	2360 2370 2380 2390 2400
Consensus	CGCAGMTRCC RAVYGAYTTY ACSCRDTTTR ACVGKVVKAA HTGYCCBGTW
BR1C.A.. A.CC..T..C ..G.AG...A ..A.TGCT.. T..C..T..A
BR8C.A.. A.CC..T..C ..G.AG...A ..A.TGCT.. C..C..T..A
HB-P22C.G.. G.GT..C..C ..C.AA...G ..A.TACG.. T..T..T..T
HB-P24C.G.. A.CC..C..T ..G.GT...A ..G.GCGT.. C..C..C..A
SPC.A.. A.CC..T..C ..G.AG...A ..A.TGCG.. T..C..T..A
NL95C.G.. A.CC..C..T ..G.GT...A ..C.GCAT.. C..C..G..A
FIA.A.. G.AT..C..T ..C.GT...G ..A.TACG.. A..T..T..T

STOP 3 ORF4

	2410 2420 2430 2440 2450
Consensus	CAGACSGTDA TMRTCATMCC CTCACYT YRA KYAAMTYAAR RGAGDWWGCA
BR1G..G. .AA....A..T. TA . GC..C.T..A G...ATA...
BR8G..G. .AA....A..T. TA . GC..C.T..A G...ATA...
HB-P22G..G. .CG....A..T. TA . GC..C.T..A G...ATA...
HB-P24C..A. .CG....A..T. TA . GC..C.T..A G...ATA...
SPG..G. .AA....A..T. TA . GC..C.T..A G...ATA...
NL95C..A. .CG....A..T. TA . GC..C.T..G A...GAA...

FI

.....G..T. .CG.....C... -..C.CG. TT..A.C..A G...TTT...

	2460 2470 2480 2490 2500
Consensus	TCYCWAAGAC AGCWWGTCRC ARRARRRAAA TTACYCAVVY ATTGRGTAAG
BR1	..T.A..... ..TA...G. .GA.GAG...T..GCTG.....
BR8	..T.A..... ..TA...G. .GA.GAG...T..GCTG.....
HB-P22	..T.T..... ..TA...A. .GA.AGA...T..CCTA.....
HB-P24	..T.T..... ..TA...A. .AG.AGA...T..AGCA.....
SP	..C.A..... ..TA...G. .GA.GAG...T..GCTG.....
NL95	..T.A..... ..TT...A. .AG.AGA...T..AACA.....
FI	..T.T..... ..AA...G. .GA.GAG...C..CCTG.....

	2510 2520 2530 2540 2550
Consensus	GTCGACATMD MYTTYGAAGA CGACATCCAY WTGTCKATYG CYAAYGAYCT
BR1CA AC..C.....C A....T..T. .C..C..C..
BR8CA AC..C.....T A....T..T. .C..T..C..
HB-P22CG AC..C.....T A....T..C. .C..T..T..
HB-P24CA AC..T.....T A....T..T. .C..T..C..
SPCA AC..C.....T A....T..T. .T..T..C..
NL95CA AC..C.....C A....T..C. .C..C..C..
FIAT CT..C.....T T....G..C. .C..C..C..

	2560 2570 2580 2590 2600
Consensus	YTKKVRGGCS TDYGGCRTHS SNVHDCTTVV BDVNGCKGAR SARTGYATYA
BR1	C.TTGA...C .AC...A.CC CGAAA...CA GAAT..T..G G.G..C..T.
BR8	C.GTGA...C .AC...A.CC CGATG...CG TGAC..T..G G.G..C..T.
HB-P22	T.TTAA...C .TT...G.TG CACCA...AC GTCA..G..G C.G..T..C.
HB-P24	C.TTGA...C .AC...A.TG CACCT...GC GTCG..G..G C.G..T..C.
SP	C.TTGA...C .AC...A.CC CTAAA...GA TTCG..G..G G.G..C..T.
NL95	C.TGCA...C .GC...G.TG CGCCA...GC GTCG..G..G C.G..T..C.
FI	T.TTAG...G .AT...G.AG GCGAA...AG CAGC..T..A G.A..C..T.

	2610 2620 2630 2640 2650
Consensus	RYACYSCRTT CCCDVRYMYK KMDMWRRVYS YDGAYMRMTT YCGBRYYSMD
BR1	AC..CG.A.. ...AAGCCTT GATCAAGGCG CA..CACA.. T..TGTCGAG
BR8	AC..CG.A.. ...AAGCCTT GATCAGAGCG CA..CACG.. T..TGTCGAG
HB-P22	AC..TC.G.. ...TGACACT TCGATGACTG CT..TGCA.. T..CATCGCG
HB-P24	GC..TC.G.. ...TGACACT TCAATGGACG CT..TGCA.. T..CATCCAT
SP	AC..CG.A.. ...GAGCCTG GATCAAGGCG TT..CACG.. C..TGTCGAA
NL95	AC..TC.G.. ...TGACACT TCAATGAACC CA..TGAA.. T..GATTCAA
FI	AT..CG.A.. ...GCGTCTG GATCAGAGTC CG..TACG.. T..TACCGAG

	2660 2670 2680 2690 2700
Consensus	TAYYTVMGNK CHGARATMYT NWSNAAGTWY RRYGSSCAYC CTCTHGGTAY
BR1	..TT.GC.CG .T..G..CT. ATCG....TC GAC.GG..C.A....C
BR8	..CT.AC.CG .C..G..CC. ATCG....TT GAC.GG..T.T....T
HB-P22	..TC.GC.GT .A..G..CC. TAGT....AT AGT.CG..C.C....T
HB-P24	..CC.GC.TT .A..G..CC. TAGC....TC AGT.CG..C.C....T
SP	..CT.AC.CG .C..A..CT. ATCA....TT GAT.GG..C.C....T

NL95 ..TC.GC.TT .A..G..AC. CAGC....TT AGC.CG..C.C....T
FI ..CC.CA.AT .C..G..CC. GTCT....TC AAT.CC..C.C....T

	27102720273027402750
Consensus	WRATACCGAA RMRRYHGCHT GGGARAARTT CCTRGCKGCT GARGAGGGHT
BR1	TG..... AAGACA..A.G..A.. ...G..T..T ..A.....T.
BR8	TA..... GAAGCA..A.G..A.. ...G..T..C ..G.....T.
HB-P22	TG..... GCAGTA..T.G..G.. ...A..T..T ..A.....C.
HB-P24	TG..... GCGGCC..C.A..G.. ...G..T..C ..G.....T.
SP	TG..... GCGGCT..A.A..G.. ...A..G..C ..G.....T.
NL95	TG..... GCAGCT..C.A..G.. ...A..G..T ..A.....A.
FI	AG..... GCAGTC..C.A..G.. ...A..G..T ..A.....T.

	27602770278027902800
Consensus	GTMGACDHAC VAAYGMRGCG CTSWCDMDVK BYAAGTACCA YGAYAATTCT
BR1	..A...TT.. G..C.AA..A ..GT.GCAGG TT..... C..T.....C
BR8	..A...GT.. G..C.AA..A ..GT.ACAAG CT..... T..T.....T
HB-P22	..A...AA.. G..C.AA..A ..GT.GCAGG CT..... T..C.....C
HB-P24	..A...AA.. C..C.AA..T ..CA.TAAGG TT..... T..T.....C
SP	..A...AA.. G..C.AA..A ..GT.GCTAG TT..... C..T.....C
NL95	..A...TT.. C..T.AG..T ..CA.TAAAG TT..... C..T.....C
FI	..C...TC.. A..C.CA..A ..GT.GAGCT GC..... C..C.....C

	28102820283028402850
Consensus	ATTTTTRTCRT GGGYGAGCG YGTWATYCA ACBGCYCGYC GAAAAATACT
BR1G..G.C..... T..A..T..T ..T..T..C.
BR8A..G.T..... C..A..T..T ..C..T..C.
HB-P22G..A.C..... T..T..C..C ..T..T..T.
HB-P24G..A.C..... T..T..T..C ..T..T..T.
SPG..G.C..... T..T..T..C ..G..C..T.
NL95G..G.C..... T..T..T..C ..T..C..T.
FIG..A.C..... T..T..T..C ..T..C..T.

	28602870288028902900
Consensus	TAARCTDATH GYGAGDCYG YNCCRYTBGG GGATGTGGCG YTRCGYKSCC
BR1	...A..A..T ..C...T.T. TG..GT.C.. T.G..CTG..
BR8	...G..A..T ..C...T.T. TG..GT.C.. T.G..TTG..
HB-P22	...G..T..A ..C...T.T. TT..GC.C.. C.G..CTG..
HB-P24	...A..G..C ..T...A.T. TC..GC.G.. T.G..CGC..
SP	...A..A..T ..C...T.T. TA..GT.C.. T.G..CTG..
NL95	...A..G..A ..T...G.T. CG..AT.G.. T.G..CGC..
FI	...G..T..C ..C...T.T. TT..GT.T.. T.A..CTG..

	29102920293029402950
Consensus	GDTTYTCBGG HGGYGCDACG ACCTCGGTDA ACCGTTTACA CGGYCAYCCG
BR1	.A..T..T.. C..T..A...A.T..T...
BR8	.T..T..C.. C..T..A...G.C..T...
HB-P22	.A..T..T.. C..C..G...T.C..C...
HB-P24	.A..T..G.. A..C..T...T.T..C...
SP	.T..T..T.. C..C..G...T.T..T...

NL95	.A..T..G..	C..C..G...T.C..C...
FI	.G..C..T..	T..C..A...T.T..C...

	29602970298029903000
Consensus	TCGTGGAAGC ATGCCTGTCC GCAGGATGTW ACCAAACGYG CRYTVAARTA
BR1T.....C..AT.C..G..
BR8T.....C..AC.C..G..
HB-P22T.....C..AT.C..A..
HB-P24A.....T..AC.G..A..
SPT.....C..AT.C..G..
NL95A.....C..AC.A..G..
FIT.....C..GC.G..G..

	30103020303030403050
Consensus	YCTVMWVGC B TWTAARMDRG CNTGYGGYGA YRYBGWHGAH YTMCGYRTCR
BR1	C..GCAA..C .T...ACGG. .G..C..T.. CGTT.TA..T T.A..CG..A
BR8	T..GCAA..C .T...ACGG. .A..C..T.. CGTT.TA..T T.A..CG..A
HB-P22	C..ACAA..G .A...AATG. .T..T..T.. CATC.TA..C T.A..TG..A
HB-P24	C..AATG..C .A...AAAG. .C..T..T.. CGTC.TT..T T.A..CG..A
SP	C..GCAA..C .T...GCGG. .C..T..T.. CGTT.TA..T C.A..CG..A
NL95	C..CATA..T .A...GAAA. .T..T..C.. CGTC.TA..T T.A..CG..A
FI	C..GCTC..C .A...AAAG. .G..C..C.. TACG.AC..A C.C..TA..G

	30603070308030903100
Consensus	RYGARGTGCG CACTTCRAAT AAAGCAGTCA CTGTTCCRAA GAAYAGYAAR
BR1	AC..G.....A.....G.. ...T..T..G
BR8	AC..G.....A.....G.. ...T..T..G
HB-P22	AT..G.....G.....A.. ...C..T..G
HB-P24	AT..G.....A.....A.. ...C..C..A
SP	AC..G.....A.....A.. ...C..T..A
NL95	AT..G.....G.....A.. ...C..T..A
FI	GC..A.....A.....G.. ...C..T..A

	31103120313031403150
Consensus	ACYGATCGYT GYATYGCYAT CGARCCYGGN TGGAAYATGT TTTTYCARYT
BR1	..T.....C. .T..C..C.. ...A..C..T ...T.... ...C..GC.
BR8	..T.....C. .T..C..T.. ...A..C..G ...T.... ...T..AC.
HB-P22	..C.....T. .C..T..C.. ...A..C..A ...T.... ...C..AC.
HB-P24	..T.....T. .C..T..C.. ...G..C..C ...T.... ...C..GT.
SP	..T.....C. .T..T..T.. ...G..C..C ...T.... ...C..GT.
NL95	..T.....C. .C..T..T.. ...G..C..C ...T.... ...C..GT.
FI	..C.....C. .T..T..C.. ...G..T..C ...C.... ...C..GC.

	31603170318031903200
Consensus	VGGYGTMGGT GCDGTRCTMC GYGATMGGTT GCGYYTDTGG NVKATYGAYC
BR1	C..T..A... ..G..G..A. .C...A.... ...TT.A... AAG..T..C.
BR8	C..T..A... ..G..G..A. .C...A.... ...CT.A... AAG..T..T.
HB-P22	G..T..C... ..G..G..A. .C...A.... ...CC.T... GGG..T..C.
HB-P24	A..C..A... ..T..G..C. .C...A.... ...CC.T... CAG..T..T.
SP	A..C..C... ..A..G..A. .C...A.... ...TT.A... AAG..T..T.

NL95
FI

A..C..C... ..T..G..C. .T...C.... ...CC.T... CAT..T..T.
C..T..C... ..A..A..A. .T...A.... ...CT.G... TCG..C..T.

	3210 3220 3230 3240 3250
Consensus	TYAATGAYCA RTCBVYYAAY CARCGYCTYG CRCGBGATGS RTCDCWGYTD
BR1	.T.....T.. G..GACC..C ..A..C..C. .G..G....G G..T.T.C.A
BR8	.T.....C.. A..GACC..T ..G..T..C. .G..G....G G..T.T.C.A
HB-P22	.T.....T.. A..CATC..T ..G..C..C. A..C....C G..A.A.C.T
HB-P24	.T.....C.. A..TGTT..T ..G..C..C. A..C....C G..G.A.T.G
SP	.T.....C.. A..GACC..T ..A..C..C. .G..T....G G..T.T.C.A
NL95	.C.....T.. A..TGTT..T ..G..C..C. A..T....C A..G.A.T.G
FI	.C.....T.. A..GCTC..C ..G..C..T. A..C....C G..A.A.T.A

	3260 3270 3280 3290 3300
Consensus	RAYCATTTTRG CYACYRTHGA YTTTRTCNGCA GCMAGYGAYT CDATMAGYHT
BR1	A.T.....A. .T..CA.A.. C..A..C... ..C..T..T. .G..C..CT.
BR8	A.T.....A. .T..CA.A.. C..A..C... ..C..T..T. .G..A..CT.
HB-P22	G.C.....G. .C..TG.T.. C..A..C... ..A..T..T. A..C..CC.
HB-P24	G.C.....G. .C..TG.C.. C..A..A... ..A..T..T. .G..A..CT.
SP	A.T.....A. .T..CA.A.. C..A..T... ..C..C..T. A..C..CC.
NL95	G.C.....G. .C..TG.C.. T..A..A... ..A..C..T. .G..A..CT.
FI	G.C.....G. .C..TG.T.. C..G..G... ..C..C..C. .T..A..TA.

	3310 3320 3330 3340 3350
Consensus	WMRRCTDGTB GARYTRCTVM TNCCBCCTGM NTGGTWYGRH STYYTRACVG
BR1	AAAG..A..T ..AT.G..GC .C..T....A A....AT.AC C.TC.G..A.
BR8	AAAG..A..T ..AT.G..GC .C..T....A A....AC.AC C.TT.A..G.
HB-P22	TAAG..G..G ..GT.A..AC .A..G....A C....TT.GC G.CC.G..C.
HB-P24	ACGG..T..T ..AT.G..AA .G..G....C T....TT.AT C.CC.G..C.
SP	TAAG..T..T ..GT.G..CA .G..C....A A....AT.AC C.TC.A..G.
NL95	ACGG..T..T ..AC.G..AA .G..G....C T....TT.AT C.CC.G..C.
FI	AAAA..T..C ..GT.A..GC .T..T....C G....TT.AA C.TC.A..C.

	3360 3370 3380 3390 3400
Consensus	ATCTCCGATC BGAYSARGGH RTHYTRCCHR AYGGDSRWGY YGTBACYTAY
BR1 T..TG.A..C G.TT.A..TG .T..ACGA.T T..G..C..T
BR8 C..TC.A..C G.TT.A..TG .T..ACGA.T T..G..C..T
HB-P22 G..TC.G..T A.CC.A..TG .T..GCGT.C C..T..C..C
HB-P24 G..CC.G..T G.CC.G..AG .T..GCGT.T C..T..T..C
SP C..TG.A..A A.AC.G..TG .C..GCGA.T T..G..C..T
NL95 G..CC.G..A A.CC.G..TG .C..GCGT.T C..T..T..C
FI G..TC.A..A G.TC.A..CA .T..TGAA.T T..C..C..C

	3410 3420 3430 3440 3450
Consensus	GAGAAAATAT CCTCCATGGG WAATGGCTAC ACTTTYGARY TNGAGTCGYT
BR1 A..... ..C..GC .T.....C.
BR8 A..... ..C..GC .T.....C.
HB-P22 T..... ..C..GC .T.....C.
HB-P24 T..... ..C..GC .G.....C.
SP T..... ..C..AC .C.....C.

NL95	T.....T..GC	.A.....T.
FI	T.....C..GT	.A.....C.

	3460 3470 3480 3490 3500
Consensus	HATHTTYGCR GCBMTMGCHM GRAGTGTGTG YGAGTTRYTR GAHMTYGACC
BR1	T..T..C..G ..GC.C..AA .A..... T.....AC.A ..AA.C....
BR8	T..T..C..G ..GC.C..AA .A..... C.....AC.A ..AA.C....
HB-P22	C..A..T..A ..GA.C..TC .G..... C.....AC.G ..TC.T....
HB-P24	A..C..T..G ..CC.A..CA .G..... C.....GT.G ..CC.T....
SP	T..T..T..G ..TA.C..TC .A..... C.....AC.G ..AA.T....
NL95	A..T..C..G ..TC.C..CA .A..... C.....AT.G ..CC.T....
FI	T..C..T..G ..TA.C..TC .G..... T.....AC.G ..CC.C....

	3510 3520 3530 3540 3550
Consensus	ARTCHRCYGT YAGCGTSTAY GGDGATGATA TWATCATCSM YWCNSRTGCV
BR1	.A..TA.T.. T.....G..C ..G..... .A.....GA CA.TCG...C
BR8	.A..TA.T.. T.....G..C ..G..... .A.....GA CA.TCG...C
HB-P22	.A..AA.T.. C.....G..C ..A..... .A.....GA TT.CCG...T
HB-P24	.G..AA.C.. C.....G..C ..T..... .A.....GA CT.ACG...C
SP	.A..TA.T.. T.....G..C ..G..... .A.....GA TA.CCG...C
NL95	.G..AA.T.. C.....G..C ..T..... .A.....GA TT.ACG...C
FI	.G..CG.C.. C.....C..T ..A..... .T.....CC CT.GGA...T

	3560 3570 3580 3590 3600
Consensus	GCDSMDVYMY TDATGGMKGT BTTYGARTAY GTYGGSTTYA CBSCKAAYMR
BR1	..AGCTCCAT .A....AT.. C..T..A..C ..T..G..C .GC.T..CAA
BR8	..AGCTCCAT .A....AT.. C..C..A..C ..T..G..T .GC.T..CAA
HB-P22	..TGCAACCC .T....AT.. G..C..G..C ..T..G..C .CC.T..CAG
HB-P24	..TGATGTCC .G....CG.. T..T..G..T ..T..G..C .GC.T..CAG
SP	..AGCTCCAT .A....AT.. C..T..G..C ..C..G..C .TC.T..CAG
NL95	..TGATGTCC .T....CG.. T..C..G..T ..T..G..T .GC.T..TCG
FI	..GCAGACCC .G....AT.. C..C..G..T ..T..C..T .GG.G..CAG

	3610 3620 3630 3640 3650
Consensus	RAAGAARACB TTYDKHRVBG GHCCSTTYMG VGARTCSTGY GGWAARCACT
BR1	A.....G..G ..CTGCGAT. .A..C..CC. C..A..G..C ..T..G....
BR8	G.....A..G ..CTGCGAT. .A..C..CC. C..A..G..C ..T..G....
HB-P22	A.....G..C ..TGTAAGT. .T..C..TA. A..G..G..C ..A..G....
HB-P24	A.....G..T ..CATTAAG. .C..C..TA. A..G..G..C ..T..G....
SP	A.....A..G ..CTGCGAT. .A..C..CC. C..A..G..C ..T..G....
NL95	A.....A..T ..CATTAAG. .C..C..TA. A..G..G..C ..A..G....
FI	A.....A..T ..CATCACC. .C..G..CC. G..A..C..T ..A..A....

	3660 3670 3680 3690 3700
Consensus	GGYWYYHHGG KGTDGAYGTR ACGCCYTTTT ACATACGHCG MCCRATHCGY
BR1	..TTCCAA.. G..A..T..AC....C.. A..A..T..T
BR8	..TTCCAA.. G..A..T..AC....C.. A..A..T..T
HB-P22	..CACTCT.. G..T..T..AT....C.. C..A..A..T
HB-P24	..CACTCC.. G..T..C..AC....C.. C..A..C..T
SP	..TTCCAA.. G..A..T..AC....A.. A..A..A..T

NL95 ..CACTCC.. G..T..C..AC.....C.. C..A..C..C
FI ..TTTCTC.. T..G..C..GT.....T.. C..G..A..T

	37103720373037403750
Consensus	WGYCTWGYYG AYATGATAYT BGTWYTRAAY ARYMTMTAYM GRTGGGGYAC
BR1	T.C..T.CT. .T.....C. G..TT.A..C .GTA.C..CA .G.....C..
BR8	T.C..T.CT. .T.....C. C..TT.G..T .GTA.C..CA .A.....C..
HB-P22	T.C..A.TC. .C.....C. T..TC.G..T .GTA.C..TC .G.....C..
HB-P24	T.C..A.CC. .C.....C. T..TT.A..C .GTA.A..TC .G.....C..
SP	T.C..A.CC. .T.....C. T..AT.A..T .GTA.C..TA .G.....C..
NL95	T.C..A.CC. .C.....C. T..AT.G..C .GTA.C..CC .G.....T..
FI	A.T..T.CC. .T.....T. G..TT.G..T .ACC.C..TC .G.....C..

	37603770378037903800
Consensus	KRTYGAYGGB RTRTGGGATC CTAGRGYVCT GMCYGTDTAY SARAAGTAYS
BR1	TG.T..T..T G.A..... .A.CA.. .A.C..G..C G.A.....TC
BR8	TG.T..T..C G.G..... .A.CA.. .A.C..A..C G.A.....TC
HB-P22	GA.T..C..C G.A..... .G.TG.. .C.C..A..C C.G.....CG
HB-P24	GA.T..C..G G.G..... .A.TA.. .C.T..G..C C.G.....CG
SP	TG.T..T..C A.A..... .A.CA.. .A.C..T..C G.A.....TC
NL95	GA.T..C..T G.G..... .G.TA.. .C.C..A..T C.G.....TG
FI	TG.C..C..T G.A..... .A.CC.. .A.C..A..C C.G.....CG

	38103820383038403850
Consensus	TBAADHTSCT RCCRAGRRAT TGGMGDMGBA AYMSDATAACC DGAYGGYTAC
BR1	.C..GT.G.. G..A..AA.. ...C.AC.C. .CCGG..... A..C..C...
BR8	.C..AT.G.. G..A..AA.. ...C.AC.C. .CCGG..... G..C..C...
HB-P22	.G..TA.G.. G..G..AA.. ...C.GC.T. .CACG..... T..T..T...
HB-P24	.G..GC.G.. A..A..AG.. ...C.GC.C. .CACT..... T..T..T...
SP	.T..AC.G.. G..A..AA.. ...C.TC.C. .TCGG..... A..C..C...
NL95	.G..AT.G.. A..A..AG.. ...C.GC.T. .CACA..... T..T..T...
FI	.G..GC.C.. A..A..GA.. ...A.GA.G. .CACG..... T..T..T...

	38603870388038903900
Consensus	GGWGACGGNG CYCTCGTCGG ATYKGCYWYR ACKAAYCCRT TYGTWMTMGT
BR1	..A.....A. .T..... .TG..TACG ..G..C..G. .C..TA.A..
BR8	..A.....A. .T..... .TG..TACG ..G..C..G. .C..TA.A..
HB-P22	..A.....C. .T..... .TG..TACA ..G..T..G. .T..TA.A..
HB-P24	..T.....T. .T..... .TG..CACG ..T..C..A. .C..TA.C..
SP	..A.....A. .T..... .TG..TACG ..G..C..G. .T..AA.A..
NL95	..A.....T. .C..... .TG..CACA ..T..C..G. .T..TA.C..
FI	..A.....G. .T..... .CT..CTTA ..G..C..G. .C..AC.C..

	39103920393039403950
Consensus	BMRRAYTWY YMVMGMBDRT AYCKGTRTT RGTTGARGTM CAGARGGAYR
BR1	TAAG..T.AT TCAA.ACTA. .C..G..A.. A.....A..CG...CG
BR8	TAAG..T.AT TCAA.ACTA. .C..G..A.. A.....A..CG...CG
HB-P22	CAAG..T.TC TCGA.ACTA. .C..G..A.. A.....A..CA...CG
HB-P24	CCGG..T.AT TCCA.ATGG. .T..T..G.. G.....G..CG...TG
SP	TAAA..T.AT TCAA.ACTA. .C..G..A.. A.....A..CG...CG

NL95
FI

CCGG..T.AT TCCA.ATGG. .T..T..G.. G.....G..CG...TG
GCGA..C.TC CAGC.CGAG. .C..G..G.. A.....G..AA...TA

	3960 3970 3980 3990 4000
Consensus	YHMMRCGHMR CGAGDWNGGY RSKTAYYTRT AYBMCHTVCT NCGYRAYCGY
BR1	TCAAG..TAGGTA..C AGT..TC.A. .TGC.T.A.. C..CG.T..C
BR8	TCAAG..CAGGTA..T AGT..TC.G. .CTC.C.A.. C..CA.T..C
HB-P22	TCAAG..TCATAC..C AGT..TT.A. .TGC.A.G.. A..CG.C..C
HB-P24	CTAAG..TCATTT..C AGT..CT.A. .TGC.T.G.. G..TG.C..C
SP	TCAAG..CAGGAG..T AGT..TC.A. .TGC.C.C.. A..TG.T..C
NL95	CAAAA..ACATTC..C AGT..CT.A. .TGC.T.A.. A..TG.T..T
FI	CCCCA..TAGAAA..T GCG..TC.A. .TCA.C.A.. T..CG.T..T

	4010 4020 4030 4040 4050
Consensus	GADRCACGTY AYARYCCBTT CCTGYRTRHY RCAGATDGGR BTRGKTTTGA
BR1	..GA.....T .T.GT..T..CG.GAC G.....A..A C.G.G.....
BR8	..GA.....T .C.GT..T..CG.GAC G.....A..A C.G.T.....
HB-P22	..GA.....T .C.AT..C..CG.GTC G.....G..A C.G.G.....
HB-P24	..AG.....T .C.AT..C..CG.ACT G.....G..A G.G.G.....
SP	..GA.....T .C.GT..T..CG.GAC G.....A..A C.G.T.....
NL95	..TG.....T .C.GT..T..CG.GTC G.....G..A C.G.G.....
FI	..GG.....C .C.AC..G..TA.GAT A.....T..G T.A.G.....

	4060 4070 4080 4090 4100
Consensus	TGAARCRCCR CTAGCTACTA GMCTTCGTCG YAAGACWGGT CGGTACAAAG
BR1G.G..GC..... C.....A...
BR8G.G..GC..... C.....A...
HB-P22G.A..GC..... C.....A...
HB-P24A.A..GC..... C.....A...
SPG.G..GC..... C.....A...
NL95G.G..AC..... C.....A...
FIG.A..GA..... T.....T...

	4110 4120 4130 4140 4150
Consensus	TDGCGTGGAT TCAGGACAGT GCCTTCATCC GGCCCCCKTA TTYNMTTAMG
BR1	.G.....
BR8	.A.....
HB-P22	.G.....
HB-P24	.G.....
SP	.G.....
NL95	.T.....
FI	.G.....

STOP 4

	4160 4170 4180 4190 4200
Consensus	GGMMTTCCCG AGGTGAAGYT YGCDMGCWAG RCAC T AGYKH GTRATGGGAA
BR1	..CA..... C. T..AA.. T .. A.....CTT ..G.....
BR8	..CA..... C. T..AA.. T .. A.....CTT ..G.....
HB-P22	..AC..... C. T..TA.. T .. A.....CTA ..A.....
HB-P24	..AA..... C. T..GA.. T .. A.....CTC ..A.....
SP	..AA..... C. C..AA.. T .. G.....CTT ..G.....
NL95	..AA..... C. T..AA.. T .. A.....CTT ..A.....

FI

..AC.....T. T..GC..A.. A.....TGC ..A.....

	4210 4220 4230 4240 4250
Consensus	RGGBSBYMKY TGASSBCCVR RRRRGAGAAA GAAAGRAAAC TCCCYYYHGH
BR1	G..CGGTCTT ...CCG..CG AGAG.....A....CTCA.T
BR8	G..CGGTCTT ...CCG..CG AGAG.....A....CTCA.T
HB-P22	G..GCTCATT ...GGC..CG GAAG.....A....TCCT.A
HB-P24	G..GCCTCTT ...GGT..CG GAAG.....A....TCCT.C
SP	G..TGGTCTC ...CCG..CG AGAG.....G....CTCC.C
NL95	G..GCCTCGT ...GGC..AG GAAG.....A....TCCT.A
FI	A..GGCTCGT ...GCC..GA GAGA.....A....TCTT-T

	4260 4270 4280
Consensus	GAGRGKKGGC TCTGCTTTGC CCWCTCTCCT CCCA
BR1	...G.TG...A.....
BR8	...G.TG...A.....
HB-P22	...G.TG...T.....
HB-P24	...G.TG...A.....
SP	...G.TG...A.....
NL95	...G.TG...A.....
FI	...A.GT... ----- ----

Alignment of Amino Acid Sequences for each Gene Appendix B1 Group I

Amino acid sequences of *Leviviridae* Group I. Note the YGDD motif in all *Leviviridae* replicase proteins.

Group I isolates DL1, DL2, DL13, DL16, J20, ST4, R17, M12, MS2 and fr. GenBank strain M12 sequences were not complete; therefore, the replicase gene was omitted. The most conserved stretch of amino acids are highlighted in red. Gaps indicate degenerative sites.

Group I maturation protein:

Alignment: Align Group I maturation protein.

	10	20	30	40	50		
Consensus	MR F	V	R	YA G	EDNS	L YRSNW	PG	STG
DL1 matura	MRAFSVLDKE	SETFVPLVRT	YADGEVEDNS	FSLKYRSNWT	PGRFNSTGAR			
DL2 matura	MRAFSVLDQE	SETFVPSVRV	YADGQVEDNS	FSLKYRSNWT	PGRFNSTGSR			
DL13 matur	MRAFSVLDQE	SETFVPSVRV	YADGQVEDNS	FSLKYRSNWT	PGRFNSTGSR			
DL16 matur	MRAFSVLDQE	SETFVPSVRV	YADGQVEDNS	FSLKYRSNWT	PGRFNSTGSR			
J20 matura	MRAFSVLDRE	SETFVPSVRV	YADGEVEDNS	FSLKYRSNWT	PGRFNSTGTR			
ST4 matura	MRAFSTLDRE	NETFVPSVRV	YADGETEDNS	FSLKYRSNWT	PGRFNSTGAK			
R17 matura	MRAFSALDKE	SKTFVPSIRV	YANGETEDNS	FSLKYRSNWT	PGRFNSTGAR			
M12 matura	MRAFSVLDQE	NETFVPSVRV	YADGETEDNS	FSLKYRSNWT	PGRFNSTGAR			
MS2 matura	MRAFSTLDRE	NETFVPSVRV	YADGETEDNS	FSLKYRSNWT	PGRFNSTGAK			
fr maturat	MRKFIPTERM	SKSHVVSURE	YADGELEDNS	LPLIYRSNWS	PGQYTSTGPR			

	60	70	80	90	100
Consensus	T WHYPS Y SRGA	DQG Y R G	SWGR	EE	G G S	DARS
DL1 matura	TKQWHYPSSY	SRGALSVTSV	DQGAYKRSGS	SWGRPYEEKA	GYGFSLDARS	
DL2 matura	TKQWHYPSPY	SRGALSVTSV	DQGAYKRSGS	SWGRPYEEKA	GFGFSLDARS	
DL13 matur	TEQWHYPSPY	SRGALSVTSV	DQGAYKRSGS	SWGRPYEEKA	GFGFSLDARS	
DL16 matur	TEQWHYPSPY	SRGALSVTSV	DQGAYKRSGS	SWGRPYEEKA	GFGFSLDARS	
J20 matura	TNQWHYPSPY	SRGALSVTSV	DQGSYKRSGS	SWGRPYEEKA	GFGFSLDARS	
ST4 matura	TKQWHYPSPY	SRGALSVTSI	DQGAYKRSGS	SWGRPYEEKA	GFGFSLDARS	
R17 matura	TKQWHYPSPY	SRGALSVTSI	DQGAYKRSGS	SWGRPYEEKA	GFGFSLDARS	
M12 matura	TKQWHYPSPY	SRGALSVTAI	DQGAYKRSGS	SWGRPYEEKT	GFGFSLDARS	
MS2 matura	TKQWHYPSPY	SRGALSVTSI	DQGAYKRSGS	SWGRPYEEKA	GFGFSLDARS	
fr maturat	TKEWHYPSSY	SRGAIGIKAL	DQGYARLGT	SWGREFEERA	GYGMSIDARS	

	110 120 130 140 150
Consensus	CYSLFPVSQN T I VP NV ANRA TEVL KVTQGNFNLG VALAEARSTA
DL1 matura	CYSLFPVSQN LTYIEVPQNV ANRASTEVLQ KVTQGNFNLG VALAEARSTA
DL2 matura	CYSLFPVSQN LTYIEVPQNV ANRASTEVLQ KVTQGNFNLG VALAEARSTA
DL13 matur	CYSLFPVSQN LTYIEVPQNV ANRASTEVLQ KVTQGNFNLG VALAEARSTA
DL16 matur	CYSLFPVSQN LTYIEVPQNV ANRASTEVLQ KVTQGNFNLG VALAEARSTA
J20 matura	CYSLFPVSQN MTYIEVPQNV ANRASTEVLQ KVTQGNFNLG VALAEARSTA
ST4 matura	CYSLFPVSQN LTYIEVPQNV ANRASTEVLQ KVTQGNFNLG VALAEARSTA
R17 matura	CYSLFPVSQN LTYIEVPQNV ANRASTEVLQ KVTQGNFNLG VALAEARSTA
M12 matura	CYSLFPVSQN LTYIEVPQNV ANRASTEVLQ KVTQGNFNLG VALAEARSTA
MS2 matura	CYSLFPVSQN LTYIEVPQNV ANRASTEVLQ KVTQGNFNLG VALAEARSTA
fr maturat	CYSLFPVSQN LTWIDVPTNV ANRATTEVLG KVTQGNFNLG VALAEARSTA

	160 170 180 190 200
Consensus	SQL TQTIAL KAYTAARRG NWRQ RYLA LNE RKF SK VA RWLELQ
DL1 matura	SQLATQTIAL VKAYTAARRG NWRQTLRYLA LNEDRKFRSK HVAGRWLELQ
DL2 matura	SQLATQTIAL VKAYTAARRG NWRQTLRYLA LNEDRKFRSK HVAGRWLELQ
DL13 matur	SQLATQTIAL VKAYTAARRG NWRQTLRYLA LNEDRKFRSK HVAGRWLELQ
DL16 matur	SQLATQTIAL VKAYTAARRG NWRQTLRYLA LNEDRKFRSK HVAGRWLELQ
J20 matura	SQLATQTIAL VKAYTAARRG NWRQALRYLA LNEDRKFRSK HVAGRWLELQ
ST4 matura	SQLATQTIAL VKAYTAARRG NWRQALRYLA LNEDRKFRSK HVAGRWLELQ
R17 matura	SQLATQTIAL VKAYTAARRG NWRQALRYLA LNEDRKFRSK HVAGRWLELQ
M12 matura	SQLATQTIAL VKAYTAARRG NWRQPVRYLA LNEDRKFRSK HVAGRWLELQ
MS2 matura	SQLATQTIAL VKAYTAARRG NWRQALRYLA LNEDRKFRSK HVAGRWLELQ
fr maturat	SQLSTQTIAL IKAYTAARRG NWRQALRYLA LNENRKFNFSK SVASRWLELQ

	210 220 230 240 250
Consensus	FGW PL SDI QGAYEMLTKV HL F PMRA VRQVG N L GRL PAA
DL1 matura	FGWLPLMSDI QGAYEMLTKV HLQEFLPMRA VRQVGTVNKL DGRLSYPAAN
DL2 matura	FGWLPLMSDI QGAYEMLTKV HLQEFLPMRA VRQVGTVNKL DGRLSYPAAN
DL13 matur	FGWLPLMSDI QGAYEMLTKV HLQEFLPMRA VRQVGTVNKL DGRLSYPAAN
DL16 matur	FGWLPLMSDI QGAYEMLTKV HLQEFLPMRA VRQVGTVNKL DGRLSYPAAN
J20 matura	FGWLPLMSDI QGAYEMLTKV HLQEFLPMRA VRQVGTVNKL DGRLSYPAAN
ST4 matura	FGWLPLMSDI QGAYEMLTKV HLQEFLPMRA VRQVGTVNKL NGRLSYPAAN
R17 matura	FGWLPLMSDI QGAYEMLTKV HLQEFLPMRA VRQVGTVNKL DGRLAYPAAN
M12 matura	FGWLPLMSDI QGAYEMLTKV HLQEFLPMRA VRQVGTVNKL DGRLSYPAAN
MS2 matura	FGWLPLMSDI QGAYEMLTKV HLQEFLPMRA VRQVGTVNKL DGRLSYPAAN
fr maturat	FGWMPLLSDI QGAYEMLTKV HLKAFMPMRA VRQVGQNVSL SGRLTSPAAS

	260 270 280 290 300
Consensus	TCNISRR IIVIWFYINDA RLAWLSSLGI LNPLGIVWEK VPFSF VDWL
DL1 matura	YQTTTCNISRR IIVIWFYINDA RLAWLSSLGI LNPLGIVWEK VPFSFVVDWL
DL2 matura	YQTTTCNISRR IIVIWFYINDA RLAWLSSLGI LNPLGIVWEK VPFSFVVDWL
DL13 matur	YQTTTCNISRR IIVIWFYINDA RLAWLSSLGI LNPLGIVWEK VPFSFVVDWL
DL16 matur	YQTTTCNISRR IIVIWFYINDA RLAWLSSLGI LNPLGIVWEK VPFSFVVDWL
J20 matura	YQTTTCNISRR IIVIWFYINDA RLAWLSSLGI LNPLGIVWEK VPFSFVVDWL
ST4 matura	FQTTTCNISRR IIVIWFYINDA RLAWLSSLGI LNPLGIVWEK VPFSFVVDWL
R17 matura	FQTTTCNISRR IIVIWFYINDA RLAWLSSLGI LNPLGIVWEK VPFSFVVDWL
M12 matura	YQTTTCNISRR IIVIWFYINDA RLAWLSSLGI LNPLGIVWEK VPFSFVVDWL

MS2 matura	FQTTCNISRR	IVIWFYINDA	RLAWLSSLGI	LNPLGIVWEK	VPFSFVVDWL
fr maturat	YKSTCNISRR	IVIWFYINDA	RLAWLSSLGI	LNPLGIVWEK	VPFSFLVDWL

	310 320 330 340 350
Consensus	LPVGNMLEGL TAP GCSY S GTVTDVI GE S I D YG W R TAK
DL1 matura	LPVGNMLEGL TAPVGCSYMS GTVTDVITGE SIISVDAPYG WTVRQGTAK
DL2 matura	LPVGNMLEGL TAPVGCSYMS GTVTDVITGE SIISVDAPYG WTVDRQGTAK
DL13 matur	LPVGNMLEGL TAPVGCSYMS GTVTDVITGE SIISVDAPYG WTVDRQGTAK
DL16 matur	LPVGNMLEGL TAPVGCSYMS GTVTDVITGE SIISVDAPYG WTVDRQGTAK
J20 matura	LPVGNMLEGL TAPVGCSYMS GTVTDVITGE SIISVDAPYG WTVRQGTAK
ST4 matura	LPVGNMLEGL TAPVGCSYMS GTVTDVITGE SIISVDAPYG WTVRQGTAK
R17 matura	LPVGNMLEGL TAPVGCSYMS GTVTDVITGE SIISVDAPYG WTVRQGTAK
M12 matura	LPVGNMLEGL TAPVGCSYMS GTVTDVITGE SIISVDAPYG WTVRQGTAK
MS2 matura	LPVGNMLEGL TAPVGCSYMS GTVTDVITGE SIISVDAPYG WTVRQGTAK
fr maturat	LPVGNMLEGL TAPIGCSYQS GTVTDVISGE STITADDIYG WDTVRPATAK

	360 370 380 390
Consensus	SA HRGV QSV PTTG Y VKSPFS VHT LDALAL RQR L
DL1 matura	AQISAMHRGV QSVWPTTG VY VKSPFSMVHT LDALALIRQR LSR
DL2 matura	AQVSAMHRGV QSVWPTTG VY VKSPFSMVHT LDALALIRQR LLR
DL13 matur	AQVSAMHRGV QSVWPTTG VY VKSPFSMVHT LDALALIRQR LLR
DL16 matur	AQVSAMHRGV QSVWPTTG VY VKSPFSMVHT LDALALIRQR LLR
J20 matura	AQISAMHRGV QSVWPTTG VY VKSPFSIVHT LDALALIRQR LSR
ST4 matura	AQISAMHRGV QSVWPTTGAY VKSPFSMVHT LDALALIRQR LSR
R17 matura	AHVSAMHRGV QSVWPTTGAY VKSPFSMVHT LDALALIRQR LSK
M12 matura	AQVSAMHRGV QSVCPPTG VY VKSPFSMVHT LDALALIRQR LSK
MS2 matura	AQISAMHRGV QSVWPTTGAY VKSPFSMVHT LDALALIRQR LSR
fr maturat	VQISAVHRGV QSVWPTTG VY VKSPFSMVHT LDALALFRQR LWK

Group I capsid protein:

Alignment: Align Group I capsid.

	
	1020304050	
Consensus	MASNF FVL VDNGGTGDV V PSNFANGV AEWISSNSRS QAYKVTCSVR	
DL1 coat	MASNFTQFVL VDNGGTGDVT VAPSNFANGV AEWISSNSRS QAYKVTCSVR	
DL2 coat	MASNFTQFVL VDNGGTGDVT VAPSNFANGV AEWISSNSRS QAYKVTCSVR	
DL13 coat	MASNFTQFVL VDNGGTGDVT VAPSNFANGV AEWISSNSRS QAYKVTCSVR	
DL16 coat	MASNFTQFVL VDNGGTGDVT VAPSNFANGV AEWISSNSRS QAYKVTCSVR	
J20 coat	MASNFTQFVL VDNGGTGDVT VAPSNFANGV AEWISSNSRS QAYKVTCSVR	
ST4 coat	MASNFTQFVL VDNGGTGDVT VAPSNFANGV AEWISSNSRS QAYKVTCSVR	
R17 coat	MASNFTQFVL VDNGGTGDVT VAPSNFANGV AEWISSNSRS QAYKVTCSVR	
M12 coat	MASNFTQFVL VDNGGTGDVT VXPSNFANGV AEWISSNSRS QAYKVTCSVR	
MS2 coat	MASNFTQFVL VDNGGTGDVT VAPSNFANGV AEWISSNSRS QAYKVTCSVR	
fr coat	MASNFEEFVL VDNGGTGDVK VAPSNFANGV AEWISSNSRS QAYKVTCSVR	

	
	60708090100	
Consensus	QSSA NRKYT KVEVPKVAT Q GGV LPV AAWRSY NME LTIP FATN	
DL1 coat	QSSAQNRYT IKVEVPKVAT QTVGGVQLPV AAWRSYLNME LTIPIFATND	
DL2 coat	QSSAQNRYT IKVEVPKVAT QTVGGVQLPV AAWRSYLNME LTIPIFATND	
DL13 coat	QSSAQNRYT IKVEVPKVAT QTVGGVQLPV AAWRSYLNME LTIPIFATND	
DL16 coat	QSSAQNRYT IKVEVPKVAT QTVGGVQLPV AAWRSYLNME LTIPIFATND	
J20 coat	QSSAQNRYT IKVEVPKVAT QTVGGVQLPV AAWRSYLNME LTIPIFATND	
ST4 coat	QSSAQNRYT IKVEVPKVAT QTVGGVELPV AAWRSYLNME LTIPIFATNS	
R17 coat	QSSAQNRYT IKVEVPKVAT QTVGGVELPV AAWRSYLNME LTIPIFATNS	
M12 coat	QSSAQNRYT IKVEVPKVAT QTVGGVELPV AAWRSYLNME LTIPIFATNS	
MS2 coat	QSSAQNRYT IKVEVPKVAT QTVGGVELPV AAWRSYLNME LTIPIFATNS	
fr coat	QSSANNRYT VKVEVPKVAT QVQGGVELPV AAWRSYMNME LTIPVFATND	

	
	110120130	
Consensus	DC LIVKA Q G K GNPI AIAANSIY	
DL1 coat	DCALIVKAMQ GLLKDGNIPI SAIAANSIY	
DL2 coat	DCALIVKAMQ GLLKDGNIPI SAIAANSIY	
DL13 coat	DCALIVKAMQ GLLKDGNIPI SAIAANSIY	
DL16 coat	DCALIVKAMQ GLLKDGNIPI SAIAANSIY	
J20 coat	DCALIVKAMQ GLLKDGNIPI SAIAANSIY	
ST4 coat	DCELIVKAMQ GLLKDGNIPI SAIAANSIY	
R17 coat	DCELIVKAMQ GLLKDGNIPI SAIAANSIY	
M12 coat	DCALIVKAMQ GLLKDGNIPI SAIAANSIY	
MS2 coat	DCELIVKAMQ GLLKDGNIPI SAIAANSIY	
fr coat	DCALIVKALQ GTFKTGNPIA TAIAANSIY	

Group I replicase protein:

Alignment: Align Group I replicase.

	
	10 20 30 40 50	
Consensus	MSK TKKFNS LCIDL DLS LE YQSIASV ATGS PHS DFTAIAYLRD	
DL1 replic	MSKTTKKFNS LCIDLPRDLS LEIYQSIASV ATGSGDPHSR DFTAIAYLRD	
DL2 replic	MSKTTKKFNS LCIDLPRDLS LEIYQSIASV ATGSGDPHSR DFTAIAYLRD	
DL13 repli	MSKTTKKFNS LCIDLPRDLS LEIYQSIASV ATGSGDPHSR DFTAIAYLRD	
DL16 repli	MSKTTKKFNS LCIDLPCDLS LEIYQSIASV ATGSGDPHSR DFTAIAYLRD	
ST4 replic	MSKTTKKFNS LCIDLPRDLS LEIYQSIASV ATGSGDPHSD DFTAIAYLRD	
R17 replic	MSKTTKKFNS LCIDLPRDLS LEIYQSIASV ATGSGNPHSD DFTAIAYLRD	
J20 replic	MSKTTKKFNS LCIDLPRDLS LEIYQSIASV ATGSGDPHSR DFTAIAYLRD	
MS2 replic	MSKTTKKFNS LCIDLPRDLS LEIYQSIASV ATGSGDPHSD DFTAIAYLRD	
fr replica	MSKSTKKFNS LCIDLSRDLS LEVYQSIASV ATGSSDPHSD DFTAIAYLRD	
	
	60 70 80 90 100	
Consensus	ELLTKHP LG GNDEATRR LAIAKL EAN GQINR G FLHD SWD	
DL1 replic	ELLTKHPSLG NGNDEATRRA LAIAKLREAN ERCGQINREG FLHDKSLSWD	
DL2 replic	ELLTKHPSLG NGNDEATRRA LAIAKLREAN ERCGQINREG FLHDKSLSWD	
DL13 repli	ELLTKHPSLG NGNDEATRRA LAIAKLREAN ERCGQINREG FLHDKSLSWD	
DL16 repli	ELLTKHPSLG NGNDEATRRA LAIAKLREAN ERCGQINREG FLHDKSLSWD	
ST4 replic	ELLTKHPTLG SGNDEATRRT LAIAKLREAN DRCGQINREG FLHDKSLSWD	
R17 replic	ELLTKHPTLG SGNDEATRRT LAIAKLREAN DRCGQINREG FLHDKSLSWD	
J20 replic	ELLTKHPSLG NGNDEATRRA LAIAKLREAN ERCGQINREG FLHDKSLSWD	
MS2 replic	ELLTKHPTLG SGNDEATRRT LAIAKLREAN GDRGQINREG FLHDKSLSWD	
fr replica	ELLTKHPNLG DGNDEATRRS LAIAKLLEAN DRCGQINRDG FLHDATASWD	
	
	110 120 130 140 150	
Consensus	PDVLQTSIRS LIGNLLSGY S LF CTFS NGA MGHKLQ DAAPYKKFAE	
DL1 replic	PDVLQTSIRS LIGNLLSGYR SSLFGQCTFS NGASMGHKLQ DAAPYKKFAE	
DL2 replic	PDVLQTSIRS LIGNLLSGYR SSLFGQCTFS NGASMGHKLQ DAAPYKKFAE	
DL13 repli	PDVLQTSIRS LIGNLLSGYR SSLFGQCTFS NGASMGHKLQ DAAPYKKFAE	
DL16 repli	PDVLQTSIRS LIGNLLSGYR SSLFGQCTFS NGASMGHKLQ DAAPYKKFAE	
ST4 replic	PDVLQTSIRS LIGNLLSGYR SSLFGQCTFS NGASMGHKLQ DAAPYKKFAE	
R17 replic	PDVLQTSIRS LIGNLLSGYQ SSLFGQCTFS NGASMGHKLQ DAAPYKKFAE	
J20 replic	PDVLQTSIRS LIGNLLSGYR SSLFGQCTFS NGASMGHKLQ DAAPYKKFAE	
MS2 replic	PDVLQTSIRS LIGNLLSGYR SSLFGQCTFS NGAPMGHKLQ DAAPYKKFAE	
fr replica	PDVLQTSIRS LIGNLLSGYS SQLFRHCTFS NGASMGHKLQ DAAPYKKFAE	

	160	170	180	190	200	
Consensus	QATVTPRAL	AA LV DQC	PWIRH	ESY FRLV	G NGVFTVPKNN	
DL1 replic	QATVTPRALR	AALLVRDQCV	PWIRHAVRYN	ESYEFRLVVG	NGVFTVPKNN	
DL2 replic	QATVTPRALR	AALLVRDQCV	PWIRHAVRYN	ESYEFRLVVG	NGVFTVPKNN	
DL13 repli	QATVTPRALR	AALLVRDQCV	PWIRHAVRYN	ESYEFRLVVG	NGVFTVPKNN	
DL16 repli	QATVTPRALR	AALLVRDQCV	PWIRHAVRYN	ESYEFRLVVG	NGVFTVPKNN	
ST4 replic	QATVTPRALR	AALLVRDQCA	PWIRHAVRYN	ESYEFRLVVG	NGVFTVPKNN	
R17 replic	QATVTPRALR	AALLVRDQCA	PWIRHAVHYN	ESYEFRLVVG	NGVFTVPKNN	
J20 replic	QATVTPRALR	AALLVRDQCA	PWIRHAVRYN	ESYKFRVLVG	NGVFTVPKNN	
MS2 replic	QATVTPRALR	AALLVRDQCA	PWIRHAVRYN	ESYEFRLVVG	NGVFTVPKNN	
fr replica	QATVTPRALK	AAVLVKDQCS	PWIRHSHVFP	ESYTFRLVGG	NGVFTVPKNN	

	210	220	230	240	250	
Consensus	KIDRAACEK	DMNMYLQKGV	G FIRRRL	VGIDLNDQ	I NQ LAQQGS	
DL1 replic	KIDRAACEK	DMNMYLQKGV	GAFIRRRLKS	VGIDLNDQTI	NQRLAQQGSI	
DL2 replic	KIDRAACEK	DMNMYLQKGV	GAFIRRRLKS	VGIDLNDQTI	NQRLAQQGSI	
DL13 repli	KIDRAACEK	DMNMYLQKGV	GAFIRRRLKS	VGIDLNDQTI	NQRLAQQGSI	
DL16 repli	KIDRAACEK	DMNMYLQKGV	GAFIRRRLKS	VGIDLNDQTI	NQRLAQQGSI	
ST4 replic	KIDRAACEK	DMNMYLQKGV	GAFIRRRLKS	VGIDLNDQSI	NQLLAQQGSV	
R17 replic	KIDRAACEK	DMNMYLQKGV	GAFIRRRLKS	VGIDLNDQSI	NQRLAQQGSA	
J20 replic	KIDRAACEK	DMNMYLQKGV	GAFIRRRLRS	VGIDLNDQTI	NQRLAQQGSV	
MS2 replic	KIDRAACEK	DMNMYLQKGV	GAFIRRRLKS	VGIDLNDQSI	NQRLAQQGSV	
fr replica	KIDRAACEK	DMNMYLQKGV	GGFIRRRLKT	VGIDLNDQTI	NQRLAQQGSR	

	260	270	280	290	300	
Consensus	DGSLATIDLS	SASDSISDRL	VW FLPPELY	SYLD IRSHY	G G IRW	
DL1 replic	DGSLATIDLS	SASDSISDRL	VWSFLPPELY	SYLDRIRSHY	GIVDGETIRW	
DL2 replic	DGSLATIDLS	SASDSISDRL	VWSFLPPELY	SYLDRIRSHY	GIVDGETIRW	
DL13 repli	DGSLATIDLS	SASDSISDRL	VWSFLPPELY	SYLDRIRSHY	GIVDGETIRW	
DL16 repli	DGSLATIDLS	SASDSISDRL	VWSFLPPELY	SYLDRIRSHY	GIVDGETIRW	
ST4 replic	DGSLATIDLS	SASDSISDRL	VWSFLPPELY	SYLDRIRSHY	GIVDGETIRW	
R17 replic	DGSLATIDLS	SASDSISDRL	VWNFLPPELY	SYLDRIRSHY	GIVDGETIRW	
J20 replic	DGSLATIDLS	SASDSISDRL	VWSFLPPELY	SYLDRIRSHY	GIIDGETIRW	
MS2 replic	DGSLATIDLS	SASDSISDRL	VWSFLPPELY	SYLDRIRSHY	GIVDGETIRW	
fr replica	DGSLATIDLS	SASDSISDRL	VWSFLPPELY	SYLDMIRSHY	GYVNGKMIRW	

	310	320	330	340	350	
Consensus	ELFSTMGNGF	TFELES MIFW	AIV ATQIHF	N GTIGIYG	DDIICP EIA	
DL1 replic	ELFSTMGNGF	TFELES MIFW	AIVKATQIHF	GNAGTIGIYG	DDIICPSEIA	
DL2 replic	ELFSTMGNGF	TFELES MIFW	AIVKATQIHF	GNAGTIGIYG	DDIICPSEIA	
DL13 repli	ELFSTMGNGF	TFELES MIFW	AIVKATQIHF	GNAGTIGIYG	DDIICPSEIA	
DL16 repli	ELFSTMGNGF	TFELES MIFW	AIVKATQIHF	GNAGTIGIYG	DDIICPSEIA	
ST4 replic	ELFSTMGNGF	TFELES MIFW	AIVKATQIHF	GNAGTIGIYG	DDIICPSEIA	
R17 replic	ELFSTMGNGF	TFELES MIFW	AIVKATQIHF	GNAGTIGIYG	DDIICPSEIA	
J20 replic	ELFSTMGNGF	TFELES MIFW	AIVKATQIHF	GNAGTIGIYG	DDIICPSEIA	
MS2 replic	ELFSTMGNGF	TFELES MIFW	AIVKATQIHF	GNAGTIGIYG	DDIICPSEIA	
fr replica	ELFSTMGNGF	TFELES MIFW	AIVRATQIHF	RNTGTIGIYG	DDIICPTEIA	

	360	370	380	390	400
Consensus	PRVLEAL Y GFKPNLRKTF SG FRES AH RGVDVK PFYI KP					
DL1 replic	PRVLEALAYY GFKPNLRKTF VSGLFRES AHFYRGVDVK PFYIKKPVDN					
DL2 replic	PRVLEALAYY GFKPNLRKTF VSGLFRES AHFYRGVDVK PFYIKKPVDN					
DL13 repli	PRVLEALAYY GFKPNLRKTF VSGLFRES AHFYRGVDVK PFYIKKPVDN					
DL16 repli	PRVLEALAYY GFKPNLRKTF VSGLFRES AHFYRGVDVK PFYIKKPVDN					
ST4 replic	PRVLEALAYY GFKPNLRKTF VSGLFRES AHFYRGVDVK PFYIKKPVDN					
R17 replic	PRVLEALAYY GFKPNLRKTF VSGLFRES AHFYRGVDVK PFYIKKPVDN					
J20 replic	PRVLEALAYY GFKPNLRKTF VSGLFRES AHFYRGVDVK PFYIRKPVDN					
MS2 replic	PRVLEALAYY GFKPNLRKTF VSGLFRES AHFYRGVDVK PFYIKKPVDN					
fr replica	PRVLEALSFY GFKPNLRKTF TSGSFRES AHYFRGVDVK PFYIKKPITD					

	410	420	430	440	450
Consensus	LF LMLI NR RGWGVV G DPRLY VW LS VP FG GTDL ADY					
DL1 replic	LFSLMLIMNR LRGWGVVGGM SDPRLYKVWV RLSSLVPSMF FG GTDLAADY					
DL2 replic	LFSLMLIMNR LRGWGVVGGM SDPRLYKVWV RLSSLVPSMF FG GTDLAADY					
DL13 repli	LFSLMLIMNR LRGWGVVGGM SDPRLYKVWV RLSSLVPSMF FG GTDLAADY					
DL16 repli	LFSLMLIMNR LRGWGVVGGM SDPRLYKVWV RLSSLVPSMF FG GTDLAADY					
ST4 replic	LFALMLILNR LRGWGVVGGM SDPRLYKVWV RLSSQVPSMF FG GTDLAADY					
R17 replic	LFALMLILNR LRGWGVVGGM SDPRLYKVWV RLSSQVPSMF FG GTDLAADY					
J20 replic	LFSLMLIMNR LRGWGVVGGM SDPRLYKVWV RLSSLVPSMF FG GTDLAADY					
MS2 replic	LFALMLILNR LRGWGVVGGM SDPRLYKVWV RLSSQVPSMF FG GTDLAADY					
fr replica	LFSLMLILNR IRGWGVVNGI ADPRLYEVWE KLSRLVPRYL FG GTDLQADY					

	460	470	480	490	500
Consensus	YVVSPP Y K R A RT GF LAR A RK FS KH SG RY					
DL1 replic	YVVSPPPTAVS VYTKTAYGRL LADTRTSGFR LARIAKERKR FSEKHDSGRY					
DL2 replic	YVVSPPNAVS VYTKTAYGRL LADARTSGFR LARIAKERKH FSEKHDSGRY					
DL13 repli	YVVSPPNAVS VYTKTAYGRL LADARTSGFR LARIAKERKH FSEKHDSGRY					
DL16 repli	YVVSPPNAVS VYTKTAYGRL LADARTSGFR LARTAKERKH FSEKHDSGRY					
ST4 replic	YVVSPPPTAVS VYTKTPYGRL LADTRTSGFR LARIARERKF FSEKHDSGRY					
R17 replic	YVVSPPPTAVS VYTKTPYGRL LADTRTSGFR LARIARERKF FSEKHDSGRY					
J20 replic	YVVSPPPTAVS VYTKTAYGRL LADARTSGFR LARIAKERKH FSEKHDSGRY					
MS2 replic	YVVSPPPTAVS VYTKTPYGRL LADTRTSGFR LARIARERKF FSEKHDSGRY					
fr replica	YVVSPPILKG IYSKMNGRRE YAEARTTGFK LARIARWRKH FSDKHDSGRY					

	510	520	530	540
Consensus	IAWFHTGGE TDSMKSAGVR RTSEWL P VP FPQECGP ASSP				
DL1 replic	IAWFHTGGEI TDSMKSAGVR VMRTSEWLTP VPTFPQECGP ASSPR				
DL2 replic	IAWFHTGGEI TDSMKSAGVR VIRTSEWLTP VPIFPQECGP ASSPR				
DL13 repli	IAWFHTGGEI TDSMKSAGVR VIRTSEWLTP VPIFPQECGP ASSPR				
DL16 repli	IAWFHTGGEI TDSMKSAGVR VIRTSEWLTP VPIFPQECGP ASSPR				
ST4 replic	IAWFHTGGEV TDSMKSAGVR IMRTSEWLTP VPTFPQECGP ASSPR				
R17 replic	IAWFHTGGEI TDSMKSAGVR IMRTSEWLTP VPTFPQECGP ASSPR				
J20 replic	IAWFHTGGEI TDSMKSAGVR VMRTSEWLTP VPTFPQECGP ASSPR				
MS2 replic	IAWFHTGGEI TDSMKSAGVR VIRTSEWLTP VPTFPQECGP ASSPR				
fr replica	IAWFHTGGEI TDSMKSAGVR VMRTSEWLQP VPVFPQECGP ASSPQ				

Appendix B2 Group II

Amino acid sequences of *Leviviridae* Group II. Note the YGDD motif in all *Leviviridae* replicase proteins.

Group II isolates DL10, DL20, T72, GA, KUI and TL2. Partial genome sequences of strain TL2 were available in GenBank. The most conserved stretch of amino acids are highlighted in red. Gaps indicate degenerative sites.

Group II maturation protein:

Alignment: Align Group II maturation.

	10	20	30	40	50
Consensus	MFPK NIDR Y V L SYD KG SDDSF E ENYL QN RS YKPGY					
DL10 matur	MFPKSNIDRN YKVTLSISYDE KGKLVSDDSF EQVENYLFQN RSNTYKPGYI					
DL20 matur	MFPKSNIDRN YKVKLISYDK KGNLVSDDSF EQVENYLFQN RSNTYKPGYI					
GA maturat	MFPKSNIDRN YKVKLISYDK KGKLVSDDSF EQVENYLFQN RSTTYKPGYI					
T72 matura	MFPKQNIDRT YKVTLSISYDK KGNVTSDDSF ESTENYLLQN RSNSYKPGYV					
KUI matura	MFPKQNIDRI YHVKLVSYDN KGKVTSDDSF ESVENYLLQN RSTTYKPGYI					
	60	70	80	90	100
Consensus	R FR PTNF WNG R F QP VG FTRKL GGRQ ADYGI VNPKNFT NS					
DL10 matur	RRDFRRPTNF WNGFRYFNQP VGTFTRKLSG GGRQVADYGI VNPKNFTGNS					
DL20 matur	RRDFRRPTNF WNGYRCFNQP VGTFTRKLSG GGRQVADYGI VNPKNFTANS					
GA maturat	RRDFRRPTNF WNGYRCFNQP VGTFTRKLSG GGRQVADYGI VNPKNFTANS					
T72 matura	RKGFRKPTNF WNGYRYFNQP VGVFTRKLDN GGRQVADYGI VNPKNFTANS					
KUI matura	RKDFRKPTNF WNGYRYFHQP VGTFTRKLSG GGRQEADYGI VNPKNFTANS					
	110	120	130	140	150
Consensus	QHLG NMVIY PGPFSINID RASVEVLNKL SQSNLNIGVA IAEAKMTASL					
DL10 matur	QHLGDNMVIY PGPFSINIDH RASVEVLNKL SQSNLNIGVA IAEAKMTASL					
DL20 matur	QHLGDNMVIY PGPFSINIDQ RASVEVLNKL SQSNLNIGVA IAEAKMTASL					
GA maturat	QHLGDNMVIY PGPFSINIDQ RASVEVLNKL SQSNLNIGVA IAEAKMTASL					
T72 matura	QHLGENMVIY PGPFSINIDN RASVEVLNKL SQSNLNIGVA IAEAKMTASL					
KUI matura	QHLGENMVIY PGPFSINIDN RASVEVLNKL SQSNLNIGVA IAEAKMTASL					

	160 170 180 190 200
Consensus	L QSI LIR AYTAAKRG W REV SQLLI EHRF P D LGGRWLELQY
DL10 matur	LAKQSIALIR AYTAAKRGKW REVLSQLLIS EHRFRAPVKD LGGRWLELQY
DL20 matur	LAKQSIALIR AYTAAKRGNW REVFSQLLIS EHRFRAPAKD LGGRWLELQY
GA maturat	LAKQSIALIR AYTAAKRGNW REVLSQLLIS EHRFRAPAKD LGGRWLELQY
T72 matura	LSRQISLIR AYTAAKRGKW REVLSQLLIA EHRFTRPSRD LGGRWLELQY
KU1 matura	LSRQSIALIR AYTAAKRGKW REVLSQLLIA EHRFTRPSKD LGGRWLELQY

	210 220 230 240 250
Consensus	GWLPLMSD K A YDLLTQT LPA MPLRV RTVG THNYK VRNVESAGDT
DL10 matur	GWLPLMSDMK AAYDLLTQTK LPAFMPLRVT RTVGGTHNYK VRNVESAGDT
DL20 matur	GWLPLMSDLK AAYDLLTQTK LPAFMPLRVT RTVGGTHNYK VRNVESAGDT
GA maturat	GWLPLMSDLK AAYDLLTQTK LPAFMPLRVT RTVGGTHNYK VRNVESAGDT
T72 matura	GWLPLMSDIK AGYDLLTQTK LPALMPLRVT RTVGGTHNYK VRNVESAGDT
KU1 matura	GWLPLMSDIK AGYDLLTQTH LPAFMPLRVS RTVGATHNYK VRNVESAGDT

	260 270 280 290 300
Consensus	WSY RLSVN YRIWYFISDP RLAWASSLGL LNPLEIYWEK TPWSFVVDWF
DL10 matur	WSYSDRLSVN YRIWYFISDP RLAWASSLGL LNPLEIYWEK TPWSFVVDWF
DL20 matur	WSYSDRLSVN YRIWYFISDP RLAWASSLGL LNPLEIYWEK TPWSFVVDWF
GA maturat	WSYRHRLSVN YRIWYFISDP RLAWASSLGL LNPLEIYWEK TPWSFVVDWF
T72 matura	WSYSDRLSVN YRIWYFISDP RLAWASSLGL LNPLEIYWEK TPWSFVVDWF
KU1 matura	WSYSDRLSVN YRIWYFISDP RLAWASSLGL LNPLEIYWEK TPWSFVVDWF

	310 320 330 340 350
Consensus	LPVGNLIEAM SNPLGLDIIS GTKTWQLESK NA A GW GTAKL AYA
DL10 matur	LPVGNLIEAM SNPLGLDIIS GTKTWQLESK LNATLTASGW SGTAKLSAYA
DL20 matur	LPVGNLIEAM SNPLGLDIIS GTKTWQLESK LNATLPAPGW SGTAKLTAYA
GA maturat	LPVGNLIEAM SNPLGLDIIS GTKTWQLESK LNATLPASGW SGTAKLTAYA
T72 matura	LPVGNLIEAM SNPLGLDIIS GTKTWQLESK LNATINASGW SGTAKLSAYA
KU1 matura	LPVGNLIEAM SNPLGLDIIS GTKTWQLESK MNASLKADGW VGTAKLSAYA

	360 370 380 390
Consensus	DRSTFYs FPTP PYVKS PLSGLH ANA LALINQRLKR
DL10 matur	KAYDRSTFYs FPTPMPYVKS PLSGLHLANA LALINQRLKR
DL20 matur	KAYDRSTFYs FPTPLPYVKS PLSGLHLANA LALINQRLKR
GA maturat	KAYDRSTFYs FPTPLPYVKS PLSGLHLANA LALINQRLKR
T72 matura	KAYDRSTFYs FPTPMPYVKS PLSGLHMANA LALINQRLKR
KU1 matura	NNGDRSTFYs FPTPLPYVKS PLSGLHMANA LALINQRLKR

Group II capsid protein:

Alignment: Align Group II capsid.

	10 20 30 40 50
Consensus	MATLRSFVLV DNGGTGNVTVPVPSNANGVA EWLSNNSRSQ AYRVTASYRA
DL10 coat	MATLRSFVLV DNGGTGNVTVPVPSNANGVA EWLSNNSRSQ AYRVTASYRA
DL20 coat	MATLRSFVLV DNGGTGNVTVPVPSNANGVA EWLSNNSRSQ AYRVTASYRA
GA coat	MATLRSFVLV DNGGTGNVTVPVPSNANGVA EWLSNNSRSQ AYRVTASYRA
T72 coat	MATLRSFVLV DNGGTGNVTVPVPSNANGVA EWLSNNSRSQ AYRVTASYRA
KU1 coat	MATLRSFVLV DNGGTGNVTVPVPSNANGVA EWLSNNSRSQ AYRVTASYRA
TL2 coat	MATLRSFVLV DNGGTGNVTVPVPSNANGVA EWLSNNSRSQ AYRVTASYRA

	60 70 80 90 100
Consensus	SGADKRKY I KLEVPKIVTQ VNGVELP S AWKA ASIDL TIPIFAATDD
DL10 coat	SGADKRKYTI KLEVPKIVTQ VVNGVELPVS AWKAYASIDL TIPIFAATDD
DL20 coat	SGADKRKYTI KLEVPKIVTQ VVNGVELPVS AWKAYASIDL TIPIFAATDD
GA coat	SGADKRKYAI KLEVPKIVTQ VVNGVELPGS AWKAYASIDL TIPIFAATDD
T72 coat	SGADKRKYTI KLEVPKIVTQ TVNGVELPVS AWKAYASIDL TIPIFAATDD
KU1 coat	SGADKRKYTI KLEVPKIVTQ SVNGVELPVS AWKAFASIDL TIPIFAATDD
TL2 coat	SGADKRKYTI KLEVPKIVTQ VVNGVELPVS AWKAYASIDL TIPIFAATDD

	110 120 130
Consensus	VT ISKSLAG LFK G P A AISSQSGFYA
DL10 coat	VTVISKSLAG LFKVGNPIAE AISSQSGFYA
DL20 coat	VTVISKSLAG LFKVGDPIAD AISSQSGFYA
GA coat	VTVISKSLAG LFKVGNPIAE AISSQSGFYA
T72 coat	VTLISKSLAG LFKIGNPVAD AISSQSGFYA
KU1 coat	VTLISKSLAG LFKIGNPVAD AISSQSGFYA
TL2 coat	VTVISKSLAG LFKVGNPIAD AISSQSGFYA

Group II lysis protein:

Alignment: Align Group II lysis.

	10	20	30	40	50
Consensus	G K S S K D F					
T72 lysis	MPSLHRVGST PKAFFSIGSE SNTGKPMFRF TEIKKTLCMD RTRDCAVRFH					
KU1 lysis	MPSLHRVGST PKAFFSIGSE SITGKPMFRF TEIEKTLCMD RTRDCAVRFH					
DL10 lysis	-----MGLK AKHKENLCSD SQRSKRLYVW IALA--IVLS ---DFTSIFS					
DL20 lysis	-----MGLK AKHKENLCSD SQRSKRLYVW IALA--IVLS ---DFTSIFS					
GA lysis	-----MGLK AKHKENLCSD SERSKRLYVW IALA--IVLS ---DFTSIFS					
TL2 lysis	-----MGLK AKHKENLCSD SQRSKRLYVW IALA--IVLS ---DFTSIFS					
	60	70			
Consensus	L D					
T72 lysis	VYLQSLDLGS SDPHSPDFDG LAYLR----					
KU1 lysis	VYLQSLDLGS SDPHSPDFDG LAYLRDECL					
DL10 lysis	HWIWGLLLILI LQ-TLMDLPT FVMSV----					
DL20 lysis	HWIWGLLLILI LR-TLMDLPT FVMNV----					
GA lysis	HWIWGLLLILI LQ-TLMDLPT FVMNV----					
TL2 lysis	HWIWGLLLILI LR-TLMDLPT FVMNV----					

Group II replicase protein:

Alignment: Align Group II replicase.

	10 20 30 40 50
Consensus	MFRF EI KT LCMDRTRDCA VRFHVYLQSL D GSSDP SP DFDGLAYLRD
T72	MFRFTEIKKT LCMDRTRDCA VRFHVYLQSL DLGSSDPHSP DFDGLAYLRD
DL10	MFRFTEIEKT LCMDRTRDCA VRFHVYLQSL DMGSSDPHSP DFDGLAYLRD
DL20	MFRFTEIEKT LCMDRTRDCA VRFHVYLQSL DLGSSDPHSP DFDGLAYLRD
GA	MFRFREIEKT LCMDRTRDCA VRFHVYLQSL DLGSSDPLSP DFDGLAYLRD
KU1	MFRFTEIEKT LCMDRTRDCA VRFHVYLQSL DLGSSDPHSP DFDGLAYLRD

	60 70 80 90 100
Consensus	ECLTKHPSLG SNSDA RKE LAYAKLMDS QRCKIQNSNG YD SHI V
T72	ECLTKHPSLG DSNSDALRKE LAYAKLMDS QRCKIQNSNG YDLSHIDSGV
DL10	ECLTKHPSLG NSNSDARRKE LAYAKLMDS QRCKIQNSNG YDYSHIESV
DL20	ECLTKHPSLG DSNSDARRKE LAYAKLMDS QRCKIQNSNG YDYSHIESV
GA	ECLTKHPSLG DSNSDARRKE LAYAKLMDS QRCKIQNSNG YDYSHIESV
KU1	ECLTKHPSLG DSNSDALRKE LAYAKLMDS QRCKIQNSNG YDLSHIDAGV

	110 120 130 140 150
Consensus	L GIL TA A A LL GFE SHFLNDCSFS NGASQGFKL DAAPFKKIAG
T72	LNGILLTAKA SIAKLLMGFE SHFLNDCSFS NGASQGFKLQ DAAPFKKIAG
DL10	LSGILKTAQA LVANLLTGFE SHFLNDCSFS NGASQGFKLR DAAPFKKIAG
DL20	LSGILKTAQA LVANLLTGFE SHFLNDCSFS NGASQGFKLR DAAPFKKIAG
GA	LSGILKTAQA LVANLLTGFE SHFLNDCSFS NGASQGFKLR DAAPFKKIAG
KU1	LNGILLTAKA LIAKLLIGFE SHFLNDCSFS NGASQGFKLR DAAPFKKIAG

	160 170 180 190 200
Consensus	QATVTAPAY AV AVKTC PW YMQETY GDET WFRRV YGNGLFSVPK
T72	QATVTAPAYD LAVHAVKTCG PWLRYMQETY GDETRWFRRV YGNGLFSVPK
DL10	QATVTAPAYN IAAVAVKTC PWAYMQETY GDETRWFRRV YGNGLFSVPK
DL20	QATVTAPAYD IAAVAVKTC PWAYMQETY GDETKWFRRV YGNGLFSVPK
GA	QATVTAPAYD IAAVAVKTC PWAYMQETY GDETKWFRRV YGNGLFSVPK
KU1	QATVTAPAYD LAVLAVKTCG PWLRYMQETY GDETRWFRRV YGNGLFSVPK

	210 220 230 240 250
Consensus	NNKIDRAACK EPDMNMYLQK GAGSFIR RL RSV IDLNDQ T NQELARLG
T72	NNKIDRAACK EPDMNMYLQK GAGSFIRRL RSVNIDLNDQ TRNQELARLG
DL10	NNKIDRAACK EPDMNMYLQK GAGSFIRKRL RSVGIDLNDQ TCNQELARLG
DL20	NNKIDRAACK EPDMNMYLQK GAGSFIRKRL RSVGIDLNDQ TRNQELARLG
GA	NNKIDRAACK EPDMNMYLQK GAGSFIRKRL RSVGIDLNDQ TRNQELARLG
KU1	NNKIDRAACK EPDMNMYLQK GAGSFIRRL RSVNIDLNDQ TRNQELARLG

	260 270 280 290 300
Consensus	SIDGSLATID LSSASDS SD RLVWDLPPH VSYL RIR SFTMIDG LH
T72	SIDGSLATID LSSASDSVSD RLVWDLPPH VSYLHRIRS SFTMIDGQLH
DL10	SIDGSLATID LSSASDSVSD RLVWDLPPH VSYLARIRS SFTMIDGRLH
DL20	SIDGSLATID LSSASDSISD RLVWDLPPH VSYLARIRS SFTMIDGRLH
GA	SIDGSLATID LSSASDSISD RLVWDLPPH VSYLARIRT SFTMIDGRLH
KU1	SIDGSLATID LSSASDSVSD RLVWDLPPH VSYLHRIRS SFTMIDGRLH

	310 320 330 340 350
Consensus	KW LFSTMG N GFTFELES MI FWALS M GVTG LG YGDDIIVP
T72	KWNLFSTMG N GFTFELES MI FWALSKSVMS YLGVTG LLI YGDDIIVPTK
DL10	KWGLFSTMG N GFTFELES MI FWALSKSVML SMGVTGSLGI YGDDIIVPVE
DL20	KWGLFSTMG N GFTFELES MI FWALSKSVML SMGVTGSLGV YGDDIIVPVE
GA	KWGLFSTMG N GFTFELES MI FWALSKSIML SMGVTGSLGI YGDDIIVPVE
KU1	KWNLFSTMG N GFTFELES MI FWALSNTVMS YLGVTG LLI YGDDIIVPTK

	360 370 380 390 400
Consensus	C P LL VLS AVNFLPN K TFFTGYFRES CGAHFFK A KPFYCKRPM
T72	CAPLLLQVLS AVNFLPNQKK TFFTGYFRES CGAHFFKGAS VKPFYCKRPM
DL10	CAPTLLKVLS AVNFLPNKKK TFFTGYFRES CGAHFFKGAD MKPFYCKRPM
DL20	CAPTLLKVLS AVNFLPNQKK TFFTGYFRES CGAHFFKGAD MKPFYCKRPM
GA	CRPTLLKVLS AVNFLPNEEK TFFTGYFRES CGAHFFKAD MKPFYCKRPM
KU1	CAPLLLQVLS AVNFLPNQKK TFFTGYFRES CGAHFFKGAS VKPFYCKRPM

	410 420 430 440 450
Consensus	ETLPD LLC NRIRGW T G G SDPRLFPI WKEFADMIPP KFKGGCNLDR
T72	ETLPDIMLLC NRIRGWGTIG GISDPRLFPI WKEFADMIPP KFKGGCNLDR
DL10	ETLPDVMLLC NRIRGWQTVG GMSDPRLFPI WKEFADMIPP KFKGGCNLDR
DL20	ETLPDVMLLC NRIRGWQTVG GMSDPRLFPI WKEFADMIPP KFKGGCNLDR
GA	ETLPDVMLLC NRIRGWQTVG GMSDPRLFPI WKEFADMIPP KFKGGCNLDR
KU1	ETLPDVLLC NRIRGWGTIG GISDPRLFPI WKEFADMIPP KFKGGCNLDR

	460 470 480 490 500
Consensus	DTYLVSPDKP G LVR A RSGFN F ENGRY HW LHMGSGEV E
T72	DTYLVSPDKP GKTLVRVAKK RSGFNHKFRS DYENGRYIHW LHMGSGEVLE
DL10	DTYLVSPDKP GVTLVRVAKV RSGFNHSFPY GHENGRYVHW LHMGSGEVLE
DL20	DTYLVSPDKP GVTLVRIAKV RSGFNHAFPY GYENGRYVHW LHMGSGEVLE
GA	DTYLVSPDKP GVSLVRIAKV RSGFNHAFPY GHENGRYVHW LHMGSGEVLE
KU1	DTYLVSPDKP GVTLVRLATV RSGFNYKFRR RQENGRYIHW LHMGSGEVSE

	510 520 530
Consensus	TISSAR RCK PNSEWR QIP LFPQE EACV LS
T72	TISSARFRCK PNSEWRTQIP LFPQEIEACV LS
DL10	TISSARFRCK PNSEWRTQIP LFPQEEACV LS
DL20	TISSARFRCK PNSEWRTQIP LFPQEEACV LS

GA
KU1

TISSARYRCK PNSEWRTQIP LFPQELEACV LS
TISSARFRCK PNSEWRIQIP LFPQEVEACV LS

Appendix B3 Group III

Amino acid sequences of *Leviviridae* Group III. Note the YGDD motif in all *Leviviridae* replicase proteins.

Group III isolates BR12, BZ1, HL4-9, TW18, VK, QB, MX1 and M11. The most conserved stretch of amino acids are highlighted in red. Gaps indicate degenerative sites.

Group III maturation protein:

Alignment: Align Group III maturation.

	10 20 30 40 50
Consensus	MP LPR LRF G E L DF QELW P S P YT G
BR12 matur	MPKLPRGLRF GADNEILNDF QELWFPDLFI ESSDTHPWYT LKGRVLNAHM
BZ1 matura	MPKLPRGLRF GADNEILNDF QELWFPDLFI ESSDTHPWYT LKGRVLNAHL
HL49 assem	MPKLPRGLRF GADNEILNDF QELWFPDLFI ESSDTHPWYT LKGRVLNAHL
TW18 matur	MPKLPRGLRF GADNEILNDF QELWFPDLFI ESSDTHPWYT LKGRVLNAHL
VK assembl	MPKLPRGLRF GADNEILNDF QELWFPDLFI ESSDTHPWYT LKGRVLNAHI
QB assembl	MPKLPRGLRF GADNEILNDF QELWFPDLFI ESSDTHPWYT LKGRVLNAHL
MX1 assemb	MPRLPRALRF GPNMEVLSDF QELWYPESII DSDVKYPLYT FRGSIGGSFF
M11 assemb	MPRLPRGLRF GANMEVLNDF QELWYPESRV DSDTIFPLYT FKGNMGGSFF

	60 70 80 90 100
Consensus	D N R RRTPH T VPIAS GLRP T V YDP L F R
BR12 matur	DDRLPNVSGR QVRRTPHRAT VPIASTGLRP VTTVQYDPAA LSF-LLNARV
BZ1 matura	DDRLPNVGGR QIRRTPHRAT VPIASSGLRP VTTVQYDPTA LSF-LLNARV
HL49 assem	DDRLPNVGGR QIRRTPHRAT VPIASSGLRP VTTVQYDPTA LSF-LLNARV
TW18 matur	DDRLPNVGGR QVRRTPHRVT VPIASSGLRP VTTVQYDPAA LSF-LLNARV
VK assembl	DDRLPNVSGR QVRRTPHRVT VPIASSGLRP VTTVQYDPAA LSF-LLNARV
QB assembl	DDRLPNVGGR QVRRTPHRVT VPIASSGLRP VTTVQYDPAA LSF-LLNARV
MX1 assemb	DSYGTNNIVR EIRRTPHCAT VPIASSGLRP CTSVWYDPTS LLFRIPEMRA
M11 assemb	DTYGTNNIVR QVRRTPHRAT VPIASSGLRP CTSVWYDPSS LLFRIPEMRA

	110 120 130 140 150
Consensus	WD G GD V DF F T APK FDFS NSL PRY A FSAFNAKYG
BR12 matur	DWDFGNGDSA DLVIKDFVFR TFAPKDFDFS NSLAPRYTQA FSAFNAKYGV
BZ1 matura	DWDFGNGDSA DLVINDFLFR TFAPKEFDFS NSLAPRYTQA FSAFNAKYGT
HL49 assem	DWDFGNGDSA NLVINDFLFR TFAPKEFDFS NSLVPRYTQA FSAFNAKYGT
TW18 matur	DWDFGNGDSA NLVINDFLFR TSAPKEFDFS NSLVPRYTQA FSAFNAKYGT
VK assembl	DWDFGNGDSA DLVIKDFVFR TFAPKDFDFS NSLVPRYTQA FSAFNAKYGV
QB assembl	DWDFGNGDSA NLVINDFLFR TFAPKEFDFS NSLVPRYTQA FSAFNAKYGT
MX1 assemb	EWDNGMGDAG DIVYKDFLFS TPAPKEFDFS NSLAPRYSNA FSAFNAKYGV
M11 assemb	VWDNGMGDTG DIVYNDFLFN TPAPKEFDFS NSLAPRYSNA FSAFNAKYGV

	160 170 180 190 200
Consensus	IGEG ET K Y LLLRRL RAV GD LR LR S Y G WKP T
BR12 matur	MIGEGLETIK YLGILLRRLR EGYRAVKRGD LRALRRVIQS YHKGKWKPTT
BZ1 matura	MIGEGLETIK YLGILLRRLR EGFRVVKHGD LRALRRVIQS YHKGKWKPTT
HL49 assem	MIGEGLETIK YLGILLRRLR EGYRAVKRGD LRALRRVIQS YHKGKWKPTT
TW18 matur	MIGEGLETIK YLGILLRRLR EGYRAVKRGD LRALRRVIQS YHNGKWKPTT
VK assembl	MIGEGLETIK YLGILLRRLR EGYRAVKRGD LRALRRVIQS YHKGKWKPTT
QB assembl	MIGEGLETIK YLGILLRRLR EGYRAVKRGD LRALRRVIQS YHNGKWKPAT
MX1 assemb	IIGEGHETLK YFALLLRRLH KAVRAVRHGD LRGLRKILDS YNKGKWKPAT
M11 assemb	IIGEGRETILK YFALLLRRLH KAVRAVRHGD LRGLRRILDS YHKGHWKPAT

	210 220 230 240 250
Consensus	AGNLWLEFRY GL PLF DI VM DW D IQ RFS VGHGED
BR12 matur	AGNLWLEFRY GLMPLFYDIR DVMLDWQSRH DKIQRLLRFS VGHGEDYVVK
BZ1 matura	AGNLWLEFRY GLMPLFYDIR DVMLDWQNRH DKIQRLLRFS VGHGEDYVVE
HL49 assem	AGNLWLEFRY GLMPLFYDIK DVMLDWQKRH DRIQRLLRFS VGHGEDYVVK
TW18 matur	AGNLWLEFRY GLMPLFYDIK DVMLDWQNRH DKIQRLLRFS VGHGEDCVVE
VK assembl	AGNLWLEFRY GLMPLFYDIR DVMLDWQNRH DKIQRLLRFS VGHGEDYVVK
QB assembl	AGNLWLEFRY GLMPLFYDIR DVMLDWQNRH DKIQRLLRFS VGHGEDYVVE
MX1 assemb	AGNLWLEFRY GLTPLFHDIK SVMDDWNRIN DKIQKLRRFS VGHGEDFKLS
M11 assemb	AGNLWLEFRY GLVPLFHDIK DVMNDWTRIN DKIQKYRRFS VGHGEDFKLS

	260 270 280 290 300
Consensus	D LYP F L GEIT RRHR GI YA NR GYA FDN GS RPSVSDWK
BR12 matur	FDNLYPALAY FKLKGEITLE RRHRHGISA NREGYAVFDN GSLRPVSDWK
BZ1 matura	FDNLYPAVAY FKLKGEITLE RRHRHGISA NREGYAVFDN GSLRPVSDWK
HL49 assem	FDSLPAVSF FKLKGEITLE RRHRHGISA NREGYAVFDN GSLRPVSDWK
TW18 matur	FDNLYPAVAY FKLKGEITLE RRHRHGISA NREGYAVFDN GSLRPVSDWK
VK assembl	FDNLYPALAY FKLKGEITLE RRHRHGISA NREGYAVFDN GSLRPVSDWK
QB assembl	FDNLYPAVAY FKLKGEITLE RRHRHGISA NREGYAVFDN GSLRPVSDWK
MX1 assemb	IDGLYPGLTH FRLSGEITVQ RRHRWGITYA NREGYATFDN GSIRPVSDWK
M11 assemb	IDGLYPGLTH FRLSGEITVQ RRHRWGIVYA NREGYATFDN GSIRPVSDWK

	310 320 330 340 350
Consensus	ELA AFINP EVAWELTPYS F DWF NVG DI QQ Q Y NI IVDG
BR12 matur	ELAVAFINPH EVAWELTPYS FVADWFLNVG DILAQQGQLY HNIEIVDGFD
BZ1 matura	ELATAFINPQ EVAWELTPYS FVVDWFLNVG DILAQQGQLY HNIDIVDGFD
HL49 assem	ELATAFINPH EVAWELTPYS FVVDWFLNVG DILAQQGQLY HNIDIVDGFD
TW18 matur	ELATAFINPH EVAWELTPYS FVVDWFLNVG DILAQQGQLY HNIDIVDGFD
VK assembl	ELAVAFINPH EVAWELTPYS FVVDWFLNVG DILAQQGQLY HNIDIVDGFD
QB assembl	ELATAFINPH EVAWELTPYS FVVDWFLNVG DILAQQGQLY HNIDIVDGFD
MX1 assemb	ELANAFINPG EVAWELTPYS FIVDWFINV G DIIEQKQWY QNIDIVDGYQ
M11 assemb	ELANAFINPG EVAWELTPYS FVVDWFINV G DIIEQKQLY QNIDIVDGYQ

	360 370 380 390 400
Consensus	RRDIR S KG RNG PV V D FY R H P ATL DT
BR12 matur	RRDIRFKSFT IKGERNQPV NVSADLSTID TFYSRLHASN IPFATLDLDT
BZ1 matura	RRDIRFKSFT IKGERNQPV NVSADLSAVD TFYSRLHTSS IPFATLDLDT
HL49 assem	RRDIRLKSFT IKGERNRPV NVSADLSAVD SFYSRLHTTK LPFATLDLDT
TW18 matur	RRDIRLKSFT IKGERNRPV NVSADLSAVD LFYSRLHTSS LPFATLDLDT
VK assembl	RRDIRFKSFT IKGERNQPV NVSADLSAID TFYSRLHASN IPFATLDLDT
QB assembl	RRDIRLKSFT IKGERNRPV NVSASLSAVD LFYSRLHTSN LPFATLDLDT
MX1 assemb	RRDIRMRSVS LKGVRNGIPV RVTGSVELVD SFYNRSHTTR IPQATLAIDT
M11 assemb	RRDIRMRSVT LKGVRNGIPV RVTGSVELVD SFYNRSHTTR IPQATLELDT

	410 420
Consensus	FSS KHV D S L TQ K R
BR12 matur	TFSSFKHVLD SIFLLTQRIK R
BZ1 matura	TFSSFKHVLD SIFLLTQRIK R
HL49 assem	TFSSYKHVLD SIFLLTQRIK R
TW18 matur	TFSSYKHVLD SIFLLTQRIK R
VK assembl	TFSSFKHVLD SVALLTQRIK R
QB assembl	TFSSFKHVLD SIFLLTQRVK R
MX1 assemb	SFSSIKHVMD SISLITQRIK R
M11 assemb	SFSSIKHVLD SISLITQXIK R

Group III capsid protein:

Alignment: Align Group III capsid.

	10 20 30 40 50
Consensus	MAKL TL IGK G TL LNPRGVNPT NGVA LS AG AVPALEKRV
BR12 coat	MAKLETVTLS NIGKDGKQTL VLNPRGVNPT NGVASLSEAG AVPALEKRV
BZ1 capsid	MAKLETVTLS NIGKDGKQTL VLNPRGVNPT NGVASLSEAG AVPALEKRV
HL49 capsid	MAKLETVTLS NIGKDGKQTL VLNPRGVNPT NGVAALSQAG AVPALEKRV
TW18 capsid	MAKLETVTLS NIGKDGKQTL VLNPRGVNPT NGVAALSQAG AVPALEKRV
VK capsid	MAKLETVTLS NIGKDGKQTL VLNPRGVNPT NGVASLSEAG AVPALEKRV
QB coat	MAKLETVTLG NIGKDGKQTL VLNPRGVNPT NGVASLSQAG AVPALEKRV
MX1 capsid	MAKLQAITLS GIGKNGDVTI VLNPRGVNPT NGVAALSEAG AVPALEKRV
M11 capsid	MAKLQAITLS GIGKKGDVTI DLNPRGVNPT NGVAALSEAG AVPALEKRV

	60 70 80 90 100
Consensus	SVSQPSRNR KNYKVQVKIQ NPT C A G CDPSVTR AY DVTFSFTQY
BR12 coat	VSVSQPSRNR KNYKVQVKIQ NPTACPANGS CDPSVTRQAY ADVTFSFTQY
BZ1 capsid	ISVSVSQPSRNR KNYKVQVKIQ NPTACSANGS CDPSVTRQAY ADVTFSFTQY
HL49 capsid	VSVSQPSRNR KNYKVQVKIQ NPTACTANGS CDPSVTRQAY ADVTFSFTQY
TW18 capsid	VSVSQPSRNR KNYKVQVKIQ NPTACTANGS CDPSVTRQAY ADVTFSFTQY
VK capsid	VSVSQPSRNR KNYKVQVKIQ NPTACSANGS CDPSVTRQAY ADVTFSFTQY
QB coat	VSVSQPSRNR KNYKVQVKIQ NPTACTANGS CDPSVTRQAY ADVTFSFTQY
MX1 capsid	ISVSVSQPSRNR KNYKVQVKIQ NPTSCTASGT CDPSVTRSAY ADVTFSFTQY
M11 capsid	ISVSVSQPSRNR KNYKVQVKIQ NPTSCTASGT CDPSVTRSAY SDVTFSFTQY

	110 120 130 ...
Consensus	ST EERA VR TEL ALLA P L AID LN PAY
BR12 coat	STDEERAFVR TELAALLAGP LLIDAIDRLN PAY
BZ1 capsid	STDEERAFVR TELAALLADP LLIDAIDQLN PAY
HL49 capsid	STDEERAFVR TELVALLASP LLIDAIDQLN PAY
TW18 capsid	STDEERAFVR TELIALASP LLIDAIDQLN PAY
VK capsid	STDEERAFVR TELAALLAGP LLIDAIDRLN PAY
QB coat	STDEERAFVR TELAALLASP LLIDAIDQLN PAY
MX1 capsid	STDEERALVR TELKALLADP MLIDAIDNLN PAY
M11 capsid	STVEERALVR TELQALLADP MLVNAIDNLN PAY

Group III read-through protein:

Alignment: Align Group III read-through protein.

	10 20 30 40 50
Consensus	MAKL TL IGK G TL LNPRGVNPT NGVA LS AG AVPALEKRV
Tw18 readt	MAKLETVTLS NIGKDGQQT VLNPRGVNPT NGVAALSQAG AVPALEKRV
HL49 readt	MAKLETVTLS NIGKDGKQT VLNPRGVNPT NGVAALSQAG AVPALEKRV
BR12 readt	MAKLETVTLS NIGKDGKQT VLNPRGVNPT NGVASLSEAG AVPALEKRV
BZ1 readth	MAKLETVTLS NIGKDGKQT VLNPRGVNPT NGVASLSEAG AVPALEKRV
VK readthr	MAKLETVTLS NIGKDGKQT VLNPRGVNPT NGVASLSEAG AVPALEKRV
QB readthr	MAKLETVTLG NIGKDGKQT VLNPRGVNPT NGVASLSQAG AVPALEKRV
M11 readth	MAKLQAITLS GIGKKGDVTL DLNPRGVNPT NGVAALSEAG AVPALEKRV
MX1 readth	MAKLQAITLS GIGKNGDVTL NLNPRGVNPT NGVAALSEAG AVPALEKRV

	60 70 80 90 100
Consensus	SVSQPSRNR KNYKVQVKIQ NPT C A G CDPSVTR AY DVTFSFTQY
Tw18 readt	VSVSQPSRNR KNYKVQVKIQ NPTACTANGS CDPSVTRQAY ADVTFSFTQY
HL49 readt	VSVSQPSRNR KNYKVQVKIQ NPTACTANGS CDPSVTRQAY ADVTFSFTQY
BR12 readt	VSVSQPSRNR KNYKVQVKIQ NPTACPANGS CDPSVTRQAY ADVTFSFTQY
BZ1 readth	ISVSVQPSRNR KNYKVQVKIQ NPTACSANGS CDPSVTRQAY ADVTFSFTQY
VK readthr	VSVSQPSRNR KNYKVQVKIQ NPTACSANGS CDPSVTRQAY ADVTFSFTQY
QB readthr	VSVSQPSRNR KNYKVQVKIQ NPTACTANGS CDPSVTRQAY ADVTFSFTQY
M11 readth	ISVSVQPSRNR KNYKVQVKIQ NPTSCTASGT CDPSVTRSAY SDVTFSFTQY
MX1 readth	ISVSVQPSRNR KNYKVQVKIQ NPTSCTASGT CDPSVTRSAY ADVTFSFTQY

	110 120 130 140 150
Consensus	ST EERA VR TEL ALLA P L AID LN PAYT L G SG P
Tw18 readt	STDEERAFVR TELIALLASP LLIDAIDQLN PAYTLLIAGG GSGSNPDS-V
HL49 readt	STDEERAFVR TELVALLASP LLIDAIDQLN PAYTLLIAGG GSGSKPDPVI
BR12 readt	STDEERAFVR TELAALLAGP LLIDAIDRLN PAYTLLIAGG GSGENPDP-V
BZ1 readth	STDEERAFVR TELAALLADP LLIDAIDQLN PAYTLLIAGG GSGENPDP-V
VK readthr	STDEERAFVR TELAALLAGP LLIDAIDRLN PAYTLLIAGG GSGENPDP-V
QB readthr	STDEERAFVR TELAALLASP LLIDAIDQLN PAYTLLIAGG GSGSKPDP-V
M11 readth	STVEERALVR TELQALLADP MLVNAIDNLN PAYTALLGVG -SGPSPGPGP
MX1 readth	STDEERALVR TELKALLADP MLIDAIDNLN PAYTALLGDG -SGPSPVPGP

	160 170 180 190 200
Consensus	PDPP PPP G G YTCPF IW L YE W IY A EL R F
Tw18 readt	IPDPPIDPPP GTGKYTCPFA IWSLEEVYEP PTRNRPWPIY NAVELRPRKF
HL49 readt	IPDPPIDPPP GTGKYTCPFA IWSLEEVYEP PTKNRPWPIY NAIELQPRKF
BR12 readt	DPDPPIDPPP GTGSYTCPFA IWSLEEVYEP PTIDRPWPIY RAVELSSRKF
BZ1 readth	DPDPPIDPPP GSGSYTCPFA IWSLEEVYEP PTSDRPWPIY YAVELQPRDF
VK readthr	NPDPPIIDPPP GTGTYTCPFA IWSLEEVYEP PTSDRPWPIY YAVELPPRDF
QB readthr	IPDPPIDPPP GTGKYTCPFA IWSLEEVYEP PTKNRPWPIY NAVELQPRF
M11 readth	DPDPPPEPPP GTGSYTCPFR IWDLSSVYEA ANSSHSDIY NAVELSPRNF

MX1 readth NPDPPLEPPP GTGSYTCPFR IWLSSIYEA ANSSHWDIY NAVELSPRKF

	210 220 230 240 250
Consensus	DV L DLLGN T WRDWD RL YTTFRG R NGYIDLDA L D
TW18 readt	DVALEDLLGN TNWRDWD SRL SYTTFRGCRG NGYIDLDATY LATDQAMRDQ
HL49 readt	DVALKDLLGN TKWRDWD SRL SYTTFRGCRS NGYIDLDASY LATDQAMRDQ
BR12 readt	DVALDDLLGN TKWRDWD SRL RYTTFRGCRG NGYIDLDATY LATDQAMLDQ
BZ1 readth	DVALDDLLGN TKWRDWD SRL SYTTFRGCRG NGYIDLDATY LATDQAMLDQ
VK readthr	DVALDDLLGN TEWRDWD SRL RYTTFRGCRG NGYIDLDATY LATDQAMLDQ
QB readthr	DVALKDLLGN TKWRDWD SRL SYTTFRGCRG NGYIDLDATY LATDQAMRDQ
M11 readth	DVALDDLLGN TNWRDWDGRL RYTTFRGCRG NGYIDL DATS LMKDEYLTSS
MX1 readth	DVTLDDLLGN TDWRDWDGRL RYTTFRGSRG NGYIDL DATS LMQDEYLTSS

	260 270 280 290 300
Consensus	KY R GK P G FG E F YLKSINAYCS LSDI AY D GV VGFWRDP
TW18 readt	KYDIREGKKP GAFGNIERFI YLKSINAYCS LSDIAAYHAD GVIVGFWRDP
HL49 readt	KYDIREGKKP GAFGNVERFI YLKSINAYCS LSDIAAYHAD GVIVGFWRDP
BR12 readt	KYDIRTGKRP GAFGKIEQFI YLKSINAYCS LSDIAAYHAD GVIVGFWRDP
BZ1 readth	KYDIRTGKRP GAFGNIEQFI YLKSINAYCS LSDIAAYHAD GVIVGFWRDP
VK readthr	KYDIRTGKRP GAFGNIEQFI YLKSINAYCS LSDIAAYHAD GVIVGFWRDP
QB readthr	KYDIREGKKP GAFGNIERFI YLKSINAYCS LSDIAAYHAD GVIVGFWRDP
M11 readth	KYLVREGKRP GVFGNIERFV YLKSINAYCS LSDITAYRTD GVIVGFWRDP
MX1 readth	KYLVREGKRP GAFGSIERFV YLKSINAYCS LSDITAYHSD GVVVGFWRDP

	310 320
Consensus	SSGGAIPFDF FD KCPI QAVIVVPR
TW18 readt	SSGGAIPFDF TKFDKTKCPI QAVIVVPRA
HL49 readt	SSGGAIPFDF TKFDKTKCPI QAVIVVPRA
BR12 readt	SSGGAIPFDF TQFDKTKCPI QAVIVVPRA
BZ1 readth	SSGGAIPFDF TEFDKTKCPI QAVIVVPRA
VK readthr	SSGGAIPFDF TEFDKTKCPI QAVIVVPRA
QB readthr	SSGGAIPFDF TKFDKTKCPI QAVIVVPRA
M11 readth	SSGGAIPFDF NEFDSNKCPI QAVIVVPRL
MX1 readth	SSGGAIPFDF SEFDSNKCPI QAVIVVPRL

Group III replicase:

Alignment: Align Group III replicase.

	10	20	30	40	50
Consensus	MSKT S SLS LRR AANTRI VE NLALSIANDL A F					
BR12 repli	MKCMSKTASS HNSLSAQLRR AANTRIEVEG NLALSIANDL MLAYGQSPFS					
BZ1 replic	MKCMSKTASS RNSLSAQLRR AANTRIEVEG NLALSIANDL MLAYGQSPFN					
HL49 repli	MKCMSKTASS HNSLSAQLRR AANTRIEVEG NLALSIANDL LLAYGQSPFN					
TW18 repli	MKCMSKTASS HNSLSAQLRR AANTRIEVEG NLALSIANDL LLAYGQSPFN					
VK replica	MKCMSKTASS HNSLSAQLRR AANTRIEVEG NLALSIANDL MLAYGQSPFS					
QB replica	MKCMSKTASS RNSLSAQLRR AANTRIEVEG NLALSIANDL LLAYGQSPFN					
MX1 replic	---MSKTLQS RKSLSGKLRR AANTRIVVEG NLALSIANDL LSALDVEPFN					
M11 replic	---MSKTSQS RKSLSGKLRR AANTRIVVED NLALSIANDL LSALDVESFS					
	60	70	80	90	100
Consensus	SE CIS P F D FR YL AE MS KYD FSLGI TEA AW KFL					
BR12 repli	SESECISLSP KFDGTPDNFR INYLKAEIMS KYDDFSLGID TEAAAWKKFL					
BZ1 replic	SESECISLGP KFDETPDNFR INYLKAEIMS KYDDFSLGID TEAAAWKKFL					
HL49 repli	SEAECISLSP RFDGTPDHFR INYLKAEVMS KYDDFSLGID TEAAAWKKFL					
TW18 repli	SEAECISLSP RFDGTPDDFR INYLKAEVMS KYDDFSLGID TEAAAWKKFL					
VK replica	SESECISLSP KFDGTPDNFR INYLKAEIMS KYDDFSLGID TEAAAWKKFL					
QB replica	SEAECISFSP RFDGTPDDFR INYLKAEIMS KYDDFSLGID TEAAAWKKFL					
MX1 replic	SEEDCISRSP KFGISPDQFR NSYLRAEIMS KYDSFSLGIN TEAAAWKKFL					
M11 replic	SEEDCISRSP KFDLSADQFR NSYLAEEIMS KYDSFSLGIN TEAAAWKKFL					
	110	120	130	140	150
Consensus	AAEAECA TN RLYRP Y E DFNFSLGE C HMARRKI K L GD E					
BR12 repli	AAEAECALTN ARLYRPNYSE DFNFSLGESC LHMARRKIVK LIGDAPSVEG					
BZ1 replic	AAEAECALTN ARLYRPNYSE DFNFSLGESC IHMARRKIVK LIGDAPSVEG					
HL49 repli	AAEAECALTN ARLYRPDYSE DFNFSLGESC IHMARRKIVK LIGDAPSVEG					
TW18 repli	AAEAECALTN ARLYRPDYSE DFNFSLGESC IHMARRKIVK LIGDAPSVEG					
VK replica	AAEAECALTN ARLYRPNYSE DFNFSLGESC LHMARRKIVK LIGDAPSVEG					
QB replica	AAEAECALTN ARLYRPDYSE DFNFSLGESC IHMARRKIVK LIGDAPSVEG					
MX1 replic	AAEAECAKTN LRLYRPNYNE DFNFSLGETC IHMARRKIVK LLGDSVPFEA					
M11 replic	AAEAECAITN QRLYRPNYNE DFNFSLGEAC IHMARRKIVK LLGDSVPFEA					
	160	170	180	190	200
Consensus	LRHCRFSGG ATTTN R G HPSFKFAL Q CTPRA YV AL A T D					
BR12 repli	MLRHCRFSGG ATTTNNRSYG HPSFKFALPQ ACTPRALKYV LALRASTHFD					
BZ1 replic	MLRHCRFSGG ATTTNNRSYG HPSFKFALPQ ACTPRALKYV LALRASTHFD					
HL49 repli	MLRHCRFSGG ATTTNNRSHG HPSFKFALPQ ACTPRALKYV LALRASTHFD					
TW18 repli	MLRHCRFSGG ATTTNNRSYG HPSFKFALPQ ACTPRALKYV LALRASTHFD					
VK replica	MLRHCRFSGG ATTTNNRSYG HPSFKFALPQ ACTPRALKYV LALRASTHFD					
QB replica	MLRHCRFSGG ATTTNNRSYG HPSFKFALPQ ACTPRALKYV LALRASTHFD					
MX1 replic	VLRHCRFSGG ATTTNSRLYG HPSFKFALAQ ECTPRAVPYV QALKACTNMD					

M11 replic MLRHCRFSGG ATTTNNRSYG HPSFKFALTQ ECTPRAVPYV QALKACTGMD

	210 220 230 240 250
Consensus	SPFN KAVTVPKNSK TDRCIAIEPG WNMFFQLGIG G R L W
BR12 repli	IRVSDISPFN KAVTVPKNSK TDRCIAIEPG WNMFFQLGIG GILRDRLRCW
BZ1 replic	IRVSDISPFN KAVTVPKNSK TDRCIAIEPG WNMFFQLGIG GILRDRLRCW
HL49 repli	IRVSDISPFN KAVTVPKNSK TDRCIAIEPG WNMFFQLGIG GILRDRLRCW
TW18 repli	IRISDISPFN KAVTVPKNSK TDRCIAIEPG WNMFFQLGIG GILRDRLRCW
VK replica	IRVSDISPFN KAVTVPKNSK TDRCIAIEPG WNMFFQLGIG GILRDRLRCW
QB replica	IRISDISPFN KAVTVPKNSK TDRCIAIEPG WNMFFQLGIG GILRDRLRCW
MX1 replic	LGITKVSPFN KAVTVPKNSK TDRCIAIEPG WNMFFQLGIG GVIREKLHLW
M11 replic	LGITKVSPFN KAVTVPKNSK TDRCIAIEPG WNMFFQLGIG GVIREKLRLW

	260 270 280 290 300
Consensus	IDLNDQTIN Q RA GS LATVDLS ASD ISLAL ELL PP WF
BR12 repli	GIDLNDQTIN QHRAHEGSVT NDLATVDLSA ASDSISLALC ELLLP PGWFE
BZ1 replic	GIDLNDQTIN QHRAHEGSVT NNLATVDLSA ASDSISLALC ELLLP PGWFE
HL49 repli	GIDLNDQTIN QHRAHEGSVT NNLATVDLSA ASDSISLALC ELLLP PGWFE
TW18 repli	GIDLNDQTIN QHRAHEGSVT NNLATVDLSA ASDSISLALC ELLLP PGWFE
VK replica	GIDLNDQTIN QHRAHEGSVT NDLATVDLSA ASDSISLALC ELLLP PGWFE
QB replica	GIDLNDQTIN QHRAHEGSVT NNLATVDLSA ASDSISLALC ELLLP PGWFE
MX1 replic	NIDLNDQTIN QVRAYSGSCS NELATVDLSS ASDTISLALV ELLLP PAWFK
M11 replic	GIDLNDQTIN QTRAYLGSRD DNLATVDLSR ASDTISLALV ELLMPPEWFK

	310 320 330 340 350
Consensus	VL LRS G LP G Y EKISSMGNG TFEESLIFA LARS CE L
BR12 repli	VLTDLRSPKG QLPDGSVITY EKISSMGNGY TFEESLIFA SLARSVCEIL
BZ1 replic	VLMDLRSPKG RLPNGSVVITY EKISSMGNGY TFEESLIFA SLARSVCEIL
HL49 repli	VLMDLRSPKG RLPDGSVITY EKISSMGNGY TFEESLIFA SLARSVCEIL
TW18 repli	VLMDLRSPKG RLPDGSVITY EKISSMGNGY TFEESLIFA SLARSVCEIL
VK replica	VLMDLRSPKG RLPDGSVITY EKISSMGNGY TFEESLIFA SLARSVCEIL
QB replica	VLMDLRSPKG RLPDGSVITY EKISSMGNGY TFEESLIFA SLARSVCEIL
MX1 replic	VLTDLRSRG MLPDGRIITY EKISSMGNGF TFEESLIFA ALARSLCELL
M11 replic	VLLALRSPKG ILPDGTVITY EKISSMGNGY TFEESLIFA ALARSLCELL

	360 370 380 390 400
Consensus	L S VTVY GDDIILPS A L EVF Y VGF TN KKT F GPFRESC
BR12 repli	DLDSSEVTY GDDIILPSRA VPALQEVFKY VGFTTNTKKT FSEGPFRESC
BZ1 replic	DLDSSEVTY GDDIILPSCA VPALQEVFKY VGFTTNTKKT FSEGPFRESC
HL49 repli	DLDSSEVTY GDDIILPSCA VPALREVFKY VGFTTNTKKT FSEGPFRESC
TW18 repli	DLDSSEVTY GDDIILPSCA VPALREVFKY VGFTTNTKKT FSEGPFRESC
VK replica	NLDSSEVTY GDDIILPSRA VPALQEVFKY VGFTTNTKKT FSEGPFRESC
QB replica	DLDSSEVTY GDDIILPSCA VPALREVFKY VGFTTNTKKT FSEGPFRESC
MX1 replic	NLQPSSVTY GDDIILPSDA CSSLIEVFSY VGFRNTNEKKT FFDGPFRESC
M11 replic	GLRPSDVTY GDDIILPSDA CSPLVEVFSY VGFRNTNKKKT FSSGPFRESC

	410 420 430 440 450
Consensus	GKHY GVDV TPFYIR RIV P DLILVLN YRWATIDG VWDPR VY
BR12 repli	GKHYYSGVDV TPFYIRRRIV SPADLILVLN NLYRWATIDG VWDPRAHSVY
BZ1 replic	GKHYYSGVDV TPFYIRRRIV SPADLILVLN NLYRWATIDG VWDPRAHSVY
HL49 repli	GKHYYSGVDV TPFYIRHRIV NPTDLILVLN NLYRWATIDG VWDPRAHSVY
TW18 repli	GKHYYSGVDV TPFYIRHRIV TPADLILVLN NLYRWATIDG VWDPRAHPVY
VK replica	GKHYYSGVDV TPFYIRRRIV SPADLILVLN NLYRWATIDG VWDPRAHSVY
QB replica	GKHYYSGVDV TPFYIRHRIV SPADLILVLN NLYRWATIDG VWDPRAHSVY
MX1 replic	GKH YFMGVDV TPFYIRHRIV SPSDLILVLN QMYRWATIDG VWDPRVYPVY
M11 replic	GKH YFLGVDV TPFYIRRRIV SPSDLILVLN QMYRWATIDG VWDPRVYPVY

	460 470 480 490 500
Consensus	KYR LP L RN PDGY GDGALVGSVL PFA NRGW R VP I D
BR12 repli	LKYRKLLPKQ LQRNTIPDGY GDGALVGSVL INPFAKNRGW IRYVPVIIDH
BZ1 replic	LKYRKLLPKQ LQRNTIPDGY GDGALVGSVL INPFAKNRGW IRYVPVIIDH
HL49 repli	LKYRKLLPKQ LQRNTIPDGY GDGALVGSVL INPFAKNRGW IRYVPVIIDH
TW18 repli	LKYRKLLPKQ LQRNTIPDGY GDGALVGSVL INPFAKNRGW IRYVPVITDH
VK replica	LKYRKLLPKQ LQRNTIPDGY GDGALVGSVL INPFAKNRGW IRYVPVIIDH
QB replica	LKYRKLLPKQ LQRNTIPDGY GDGALVGSVL INPFAKNRGW IRYVPVITDH
MX1 replic	TKYRRLLPDI LRRNVVPDGY GDGALVGSVL TSPFAENRGW VRRVPMIIDK
M11 replic	TKYRRYLPEI LRRNVVPDGY GDGALVGSVL ISPFAENRGW VRRVPMIIDK

	510 520 530 540 550
Consensus	DR R EG SYLY L S E P G
BR12 repli	TRDRERTESG SYLYDLFSRC FSESNDGLPL RGPSSCDPVY PLAIDQLICK
BZ1 replic	TRDRERTEPG SYLYDLFSRC FSESNDGLPL RGPSSCDSIY RFAVDQLICK
HL49 repli	TRDRERVESG SYLYDLFSRC FSEGNDGLPL RGPSSCDSVD LSAIDQLICR
TW18 repli	TRDRERAELG SYLYDLFSRC LPESNDGLPL RGPSSCDSVN LSAVDQLICR
VK replica	TRDRERTESG SYLYDLFSRY FSESNDGLPL RGPSSCDSIY PLAIDQLICK
QB replica	TRDRERAELG SYLYDLFSRC LSESNDGLPL RGPSGCDSAD LFAIDQLICR
MX1 replic	KKDRVRDERG SYLYELWSLQ QLECDSEFPF NGSLVVGTTND GVCTYRHRER
M11 replic	RKDRVRDEYG SYLYELWSLQ QLECDSEFPF NGSLVVGSTD GTLAYAHRER

	560 570 580 590
Consensus	T S D I C SRVLAPYG F L
BR12 repli	SNPTKISRST GKFQVQYIAC SSRVLAPYGV FQGTKVASLH EA
BZ1 replic	SNPTKISRST GKFQVQYIAC SSRVLAPYGV FQGTKVMSPLH EA
HL49 repli	SNPTKISRST GKFQVQYIAC GSRVLAPYGV FQGTKVTSLSH EV
TW18 repli	SNPTKISRST GKFQVQYIAC SSRVLAPYGV FQGTKVTSLSH EV
VK replica	SNPTKISRST GKFQVQYIAC SSRVLAPYGV FQGTKVASLH EA
QB replica	SNPTKISRST GKFQVQYIAC SSRVLAPYGV FQGTKVASLH EA
MX1 replic	-VSTAISDSV GAYDIVWIPC SSRVLAPYGD FRRHEGSILK --
M11 replic	-LPTVISDAV SAFDIMWIPC SSRVLAPYGD FRRHEGSILK MG

Appendix B4 Group IV

Amino acid sequences of *Leviviridae* Group IV. Note the YGDD motif in all *Leviviridae* replicase proteins.

Group IV isolates BR1, BR8, HB-P22, HB-P24, SP, NL95 and FI. The most conserved stretch of amino acids are highlighted in red. Gaps indicate degenerative sites.

Group IV maturation protein:

Alignment: Align Group IV maturation.

	10	20	30	40	50
Consensus	MP LP GLRF GS GE NDF LWFP G L GY					
BR1 assemb	MPTLPRGLRF GSNGEVLNDF EALWFPERHT VDLSNGTCKL TGYITNLPGY					
BR8 assemb	MPTLPRGLRF GSNGEVLNDF EALWFPERHT VDLSNGTCKL TGYITNLPDY					
SP assembl	MPTLPRGLRF GSNGEVLNDF EALWFPERHT VDLSNGTCKL TGYITNLPGY					
HB-P22 asm	MPALPRGLRF GSNGEILNDF NELWFPELVS SELNLGTYNL TGYISNLPGY					
HB-P24 asm	MPTLPRGLRF GSNGEVLNDF NALWFPERET FDSGLGSYEL TGYVSNQPGY					
NL95 assem	MPTLPRGLRF GSNGEIVNDF NALWFPEREA FDLELGSYTL TGYVSNQPGY					
FI assembl	MPTLPIGLRF GSKGEILNDF SALWFPKRVS FDSQLGRYEL SGYLNQ--F					

	60	70	80	90	100
Consensus	N TP R TV PVNH GYRPV TTVEY P GT RLDG V					
BR1 assemb	SNIFPNKGVV VARTPYRSTV PVNHLGYRPV TTVEYIPDGT YVRLDGHVKF					
BR8 assemb	SNTFPNKGVT VARTPYRSTV PVNHLGYRPV TTVEYIPDGT YVRLDGHVKF					
SP assembl	SDIFPNKGVV AARTPYRSTV PVNHLGYRPV TTVEYIPDGT YVRLDGHVKF					
HB-P22 asm	EVKERNKGSH VLRTPYRSTV PVNHLGYRPV TTVEYNPLGT FIRLDGDVKF					
HB-P24 asm	ETMRNPSMH CIRTTPHRSTV PVNHLGYRPV TTVEYTPNGT FIRLDGDVKF					
NL95 assem	TTRMRNPRMH CVRTPHRSTV PVNHFGYRPV TTVEYVPNGT FIRLDGDVKF					
FI assembl	SDYGRNPALQ VSKTPHRATV PVNHLGYRPV TTVEYVPNGT FVRLDGTVRI					

	110	120	130	140	150
Consensus	G V L N LA Q GFDYQSVI GPRFS F A FSTKYG LLG					
BR1 assemb	EGGLVNGSVD LTNYVISLAA QGGFDYQSVI GPRFSAHFSA FSTKYGVLLG					
BR8 assemb	EGGLVNGSVD LTNFVISLAA QGGFDYQSVI GPRFSAHFSA FSTKYGVLLG					
SP assembl	EGDLVNGSVD LTNFVISLAA QGGFDYQSVI GPRFSARFSA FSTKYGVLLG					
HB-P22 asm	SGGLVSASLR LDNYVVGLAS QAGFDYQSVI GPRFSSQFSA FSTKYGTLLG					
HB-P24 asm	SGGAVSGSLK LSNYVVNLAA QGGFDYQSVI GPRFSAQFAA FSTKYGTLLG					
NL95 assem	SGGSVSGSLK LNNFVVNLAS QGGFDYQSVI GPRFSSQFSA FSTKYGALLG					
FI assembl	SGELVNGTVR LDNYIVNLAA QGGFDYQSVI GPRFSSQFSA FSTKYGTLLG					

	160 170 180 190 200
Consensus	EGRETL YLL L RR REG AV GD KR LRN EP G
BR1 assemb	EGRETLKYLL LLLRRMREGY RAVRRGDLKR LRNVISTFEP TSSKGRKARS
BR8 assemb	EGRETLKYLL LLLRRMREGY RAVRRGDFKR LRNVISTFEP ISPRGKMARS
SP assembl	EGRETLKYLL LVVRRMREGY RAVRRGDLKR LRNVISTFEP STIKGKRARA
HB-P22 asm	EGRETLSYLL LLFRRVREGI RAVRRGDLKR LRNLVRSYEP TSVNGRRQAA
HB-P24 asm	EGRETLSYLL LLFRRVREGI RAVKHGDLKR LRNLIRTFEP RSVPGKRQRA
NL95 assem	EGRETLSYLL LLFRRMREGI RAVRRGDLKR LRNLIRTFEP RSISGRRQRT
FI assembl	EGRETLSYLL LLFRRMREGF LAVKRGDLKR LRNVIRTFEP RSQAGKRQRS

	210 220 230 240 250
Consensus	F Y D W SSAS LWLEFRYGL MPLFYDI S
BR1 assemb	EFSQTYRDKL TGNKVEVKPS EGKWNSSSAS DLWLEFRYGL MPLFYDIQSV
BR8 assemb	EFSQTYRDKL TGNKVVKVPS EGKWNSSSAS DLWLEFRYGL MPLFYDIQSV
SP assembl	EFSQTYRDKL TGNKVEVRPS EGKWNSSSAS DLWLEFRYGL MPLFYDIQSV
HB-P22 asm	SFSGTYRDEL AN----- -GKWKDSSAS DLWLEFRYGL MPLFYDIQSL
HB-P24 asm	SFSEAYRDR T AN----- -GSWKDSSAS DLWLEFRYGL MPLFYDIQSV
NL95 assem	TFSETYRDR L VN----- -DGWKDSSAS DLWLEFRYGL MPLFYDIKSV
FI assembl	SFNKSYCDKL AS----- -GEWKNSSAS NLWLEFRYGL MPLFYDIQSV

	260 270 280 290 300
Consensus	MEDFMR HK IAK QRFSAG HGKL V F P F E VTAVLQRRHR
BR1 assemb	MEDFMRVHKK IAKIQRFSAG HGKLETVSSR FYPDVHFALE VTAVLQRRHR
BR8 assemb	MEDFMRVHKK IAKIQRFSAG HGKLETVSSR FYPDVHFALE VTAVLQRRHR
SP assembl	MEDFMRVHKK IAKIQRFSAG HGKLETVSSR FYPDVHFSLE VTAVLQRRHR
HB-P22 asm	MEDFMRVHKK IAKIQRFSAG HGKLVEVSGT FYPDVHFGLE VTAVLQRRHR
HB-P24 asm	MEDFMRVHKK IAKLQRFSAG HGKLVEVKGK FFPDPHFALE VTAVLQRRHR
NL95 assem	MEDFMR IHKK IAKLQRFSAG HGKLVTVKGR FFPDPHFAIE VTAVLQRRHR
FI assembl	MEDFMRVHKR IAKIQRFSAG HGKLEKVSDI FYPSTYFQLE VTAVLQRRHR

	310 320 330 340 350
Consensus	WGV YQDT F NG L PV DW TAA A LNPAE AW E TP SFV D
BR1 assemb	WGVIIYQDTGS YATFNNGRLV PVKDWKTAAF ALLNPAEVAW EVTPYSFVVD
BR8 assemb	WGVVYQDTGS YATFNNGRLV PVKDWKTAAF ALLNPAEVAW EITPYSFVVD
SP assembl	WGVIIYQDTGS FATFNNGRLV PVKDWKTAAF ALLNPAEVAW EVTPYSFVVD
HB-P22 asm	WGVIIYQDTGS YATFDNGRLV PVKDWQTAAL ALLNPAETAW ELTPYSFVAD
HB-P24 asm	WGVIIYQDTGS FATFNNGRLI PVRDWQTAAL ALLNPAETAW ELTPYSFVAD
NL95 assem	WGVIIYQDTGS LPPFNNGQLI PVRDWQTAAL ALLNPAETAW ELTPYSFVAD
FI assembl	WGVIIYQD TDT FATFDNGRLI PVRDWQTAAL AFLNPAETAW ELTPLSFVAD

	360 370 380 390 400
Consensus	WFFNVGDMLE Q L V DVVDGFDRKD L S SVRV A
BR1 assemb	WFFNVGDMLE QMG-QLYRHV DVVDGFDRKD IKLKSVSVRV LTGDSAHVAK
BR8 assemb	WFFNVGDMLE QMG-QLYRHV DVVDGFDRKD IRLKSISVRV LTSDSAHVAS
SP assembl	WFFNVGDMLE QMG-QLYRHV DVVDGFDRKD IKLKSVSVRV LTNDVAHVAS
HB-P22 asm	WFFNVGDMLE QIG-QLYRHV DVVDGFDRKD VKLRSVSVRV IA-NGATNTQ
HB-P24 asm	WFFNVGDMLE QIG-QLYRHV DVVDGFDRKD VKLKSVSVRV IR-DDATHTS

NL95 assem	WFVNVGDMLE	QTRPALSVNV	DVVDGFDRKD	VKLRSVSVRV	IR-DDATTTS
FI assembl	WFVNVGDMLE	QIG-QLYRYV	DVVDGFDRKD	VKLKSVSVRV	IA-PSADSAT

	410 420 430 440 450
Consensus	F L A LLH YSRVHTVA FPQISPQ D E RS KHVID S ALLTQR K
BR1 assemb	FTLRQARLLH SYYSRVHTVA FPQISPQLDT EIRSVKHVID SIALLTQRVK
BR8 assemb	FTLRQARLLH SYYSRVHTVA FPQISPQLDT EIRSVKHVID SIALLTQRVK
SP assembl	FQLRQAKLLH SYYSRVHTVA FPQISPQLDT EIRSVKHVID SIALLTQRVK
HB-P22 ass	FVLRRASLLH SFYSRVHTVA FPQISPQLDT EVRSVKHVID SIALLTQRFK
HB-P24 ass	FSLAGAVLLH SFYSRVHTVA FPQISPQLDT EIRSVKHVID SIALLTQRFK
NL95 assem	FNLSGAKLLH SFYSRVHTVA FPQISPQLDT EVRSFKHVID SVALLTQRFK
FI assembl	FELRRAELLH GFYSRVHTVA FPQISPQIDA EVRSVKHVID SLALLTQRFK

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Consensus	
BR1 assemb	R-
BR8 assemb	R-
SP assembl	R-
HB-P22 asm	R-
HB-P24 asm	RR
NL95 assem	KR
FI assembl	R-

Group IV capsid protein:

Alignment: Align Group IV capsid.

	10 20 30 40 50
Consensus	MAKLN VTL GK T TLTPRGVNPT NGVA LSEAG AVPALEKRVT
BR1 coat	MAKLNQVTLS KIGKNGDQTL TLTPRGVNPT NGVASLSEAG AVPALEKRVT
BR8 coat	MAKLNQVTLS KLGKNGDQTL TLTPRGVNPT NGVASLSEAG AVPALEKRVT
SP coat	MAKLNQVTLS KIGKNGDQTL TLTPRGVNPT NGVASLSEAG AVPALEKRVT
HB-P22 coa	MAKLNQVTLT KLGKAGDQTL TLTPRGVNPT NGVASLSEAG AVPALEKRVT
HB-P24 coa	MAKLNKVTLT GIGKAGDQTL TLTPRGVNPT NGVASLSEAG AVPALEKRVT
NL95 coat	MAKLNKVTLT GIGKAGNQTL TLTPRGVNPT NGVASLSEAG AVPALEKRVT
FI coat	MAKLNTVTLT KLGKEANKTM TLTPRGVNPT NGVATLSEAG AVPALEKRVT

	60 70 80 90 100
Consensus	VSVAQPSRNR KN K QIKLQ NPTACT DA DPSVTRS D TLSFTSYS
BR1 coat	VSVAQPSRNR KNFKVQIKLQ NPTACTKDA- DPSVTRSAFA DVTLSFTSYS
BR8 coat	VSVAQPSRNR KNFKVQIKLQ NPTACTKDAC DPSVTRSAFA DVTLSFTSYS
SP coat	VSVAQPSRNR KNFKVQIKLQ NPTACTRDAC DPSVTRSAFA DVTLSFTSYS
HB-P22 coa	VSVAQPSRNR KNYKVQIKLQ NPTACTKDAC DPSVTRSAFA DLTLSFTSYS
HB-P24 coa	VSVAQPSRNR KNYKIQIKLQ NPTACTKDAC DPSVTRSAFA DLTLSFTSYS
NL95 coat	VSVAQPSRNR KNYKVQIKLQ NPTACTKDAC DPSVTRSGSR DVTLSFTSYS
FI coat	VSVAQPSRNR KNFKVQIKLQ NPTACTKDAC DPSVTRSAFA DLTLSFTSYS

	110 120 130 ..
Consensus	T ERAL RT ELAALL D L I DAIDNLNP AY
BR1 coat	TDEERALIRT ELAALLQDNL IIDDAIDNLNP AY
BR8 coat	TDAERALIRT ELAALLQDPL IVDAIDNLNP AY
SP coat	TDEERALIRT ELAALLADPL IVDAIDNLNP AY
HB-P22 coa	TDAERALIRT ELAALLQDPL IVDAIDNLNP AY
HB-P24 coa	TDVERALVRT ELAALLKDDL IVDAIDNLNP AY
NL95 coat	TERERALIRT ELAALLKDDL IVDAIDNLNP AY
FI coat	TDEERALIRT ELAALLADPL ITDAIDNLNP AY

Group IV read-through protein:

Alignment: Align Group IV read-through.

	10 20 30 40 50
Consensus	MAKLN VTL GK T TLTPRGVNPT NGVA LSEAG AVPALEKRVT
BR1 readth	MAKLNQVTLS KIGKNGDQTL TLTPRGVNPT NGVASLSEAG AVPALEKRVT
BR8 readth	MAKLNQVTLS KLGKNGDQTL TLTPRGVNPT NGVASLSEAG AVPALEKRVT
SP readthr	MAKLNQVTLS KIGKNGDQTL TLTPRGVNPT NGVASLSEAG AVPALEKRVT
HB-P22 rea	MAKLNQVTLT KLGKAGDQTL TLTPRGVNPT NGVASLSEAG AVPALEKRVT
HB-P24 rea	MAKLNKVTLT GIGKAGDQTL TLTPRGVNPT NGVASLSEAG AVPALEKRVT
NL95 readt	MAKLNKVTLT GIGKAGNQTL TLTPRGVNPT NGVASLSEAG AVPALEKRVT
FI readthr	MAKLNTVTLT KLGKEANKTM TLTPRGVNPT NGVATLSEAG AVPALEKRVT

	60 70 80 90 100
Consensus	VSVAQPSRNR KN K QIKLQ NPTACT DAC DPSVTRS D TLSFTSYS
BR1 readth	VSVAQPSRNR KNFKVQIKLQ NPTACTKDAC DPSVTRSAFA DVTLSFTSYS
BR8 readth	VSVAQPSRNR KNFKVQIKLQ NPTACTKDAC DPSVTRSAFA DVTLSFTSYS
SP readthr	VSVAQPSRNR KNFKVQIKLQ NPTACTRDAC DPSVTRSAFA DVTLSFTSYS
HB-P22 rea	VSVAQPSRNR KNYKVQIKLQ NPTACTKDAC DPSVTRSAFA DLTLSFTSYS
HB-P24 rea	VSVAQPSRNR KNYKIQIKLQ NPTACTKDAC DPSVTRSAFA DLTLSFTSYS
NL95 readt	VSVAQPSRNR KNYKVQIKLQ NPTACTKDAC DPSVTRSGSR DVTLSFTSYS
FI readthr	VSVAQPSRNR KNFKVQIKLQ NPTACTKDAC DPSVTRSAFA DLTLSFTSYS

	110 120 130 140 150
Consensus	T ERAL RT ELAALL D L I DAIDNLNP AY AALL S G P P
BR1 readth	TDEERALIRT ELAALLQDNL IIDAIDNLNP AY-AALLVAS SGGGDNPSPMP
BR8 readth	TDAERALIRT ELAALLQDPL IVDAIDNLNP AY-AALLVAS SGGGDNPSPK
SP readthr	TDEERALIRT ELAALLADPL IVDAIDNLNP AY-AALLVAS SGGGDNPSPD
HB-P22 rea	TDAERALIRT ELAALLQDPL IVDAIDNLNP AY-AALLAAS SSGGDNP-KP
HB-P24 rea	TDVERALVRT ELAALLKDDL IVDAIDNLNP AY-AALLAAS PGGGNNP-NP
NL95 readt	TERERALIRT ELAALLKDDL IVDAIDNLNP AY-AALLAAS PGGGNNP-YP
FI readthr	TDEERALIRT ELAALLADPL ITDAIDNLNP AYWAALLVSS SGGEVKKPIP

	160 170 180 190 200
Consensus	VP P VKP P GTG CP F CYR G Y G E V
BR1 readth	DVPDLPDVKP PDGTGRYTCP FSCYRLGSIY EEGKDGSPDI YERGDEVSVT
BR8 readth	DVPDLPDVKP PDGTGRYTCP FACYRLGSIY EVGKDGSPDI YERGDEVSVT
SP readthr	DVPVVPDVKP PDGTGRYKCP FACYRLGSIY EVGKEGSPDI YERGDEVSVT
HB-P22 rea	DVPDRPDVKP PGGTGSYRCP FTCYRLGNII EVGQNGSPDI YARGDEVQVM
HB-P24 rea	GVPDSPNVKP PGGTGTYRCP FACYRRGELI TEAKDGACAL YALGSEAIVE
NL95 readt	GVPDSPNVKP PGGTGTYRCP FACYRRGELI TEAKDGACAL YACGSEALVE
FI readthr	DVPDVPSVKP PGGTGSFTCP FSCYRLDTII EAGKDGVPDL YEQGPEVTVT

	210 220 230 240 250
Consensus	F YA EDFLG N WRNWD R LS YD R RCRGNGY DL A MQ D
BR1 readth	FDYALEDFLG NTNWRNWDQR LSSYDLANRR RCRGNGYIDL DATVMQSDEF
BR8 readth	FDYALEDFLG NTNWRNWDQR LSSYDLANRR RCRGNGYIDL NATAMQSDEC
SP readthr	FDYALEDFLG NTNWRNWDQR LSDYDIANRR RCRGNGYIDL DATAMQSDDF
HB-P22 rea	FDYALEDFLG NTNWRNWDQR LSNYDIANRR RCRGNGYVDL DATAMQTDSF
HB-P24 rea	FDYALEDFLG NVFWRNWDGR LSTYDIDTHR RCRGNGYVDL DATMMQSDAY
NL95 readt	FEYALEDFLG NEFWRNWDGR LSKYDIETHR RCRGNGYVDL DASVMQSDEY
FI readthr	FDYAVEDFLG NTNWRNWDGR LSNYDIGNLR RCRGNGYVDL DATAMQSDSY

	260 270 280 290 300
Consensus	VLSG Y V K P F Y L DL VTAY SYGMVIGFW
BR1 readth	VLSGRYPVRK VKFPGAFGSI KYLLNIQGDA WDLSEVTAY RSYGMVIGFW
BR8 readth	VLSGRYPVRK VKFPGAFGSI KYLLNIQGDA WDLSEVTAY RSYGMVIGFW
SP readthr	VLSGRYGVK VKFPGAFGSI KYLLNIQGDA WDLSEVTAY RSYGMVIGFW
HB-P22 rea	VLSGKYPVRK VKFPGAFGAL KYLLNIKDDA WVDLSEVTAY RSYGMVIGFW
HB-P24 rea	VLSGAYDVVK MQPPSIFDSP RYYLHLMDGI YVDLAEVTAY HSYGMVIGFW
NL95 readt	VLSGAYDVVK MQPPGTFDSP RYYLHLMDGI YVDLAEVTAY RSYGMVIGFW
FI readthr	VLSGKYRVRK GLPPGIFASP RYYLELQDGA WVDLAAVTAY RSYGMVIGFW

	310 320 330
Consensus	TDSKSPQ P DFT F C PVQTVI IPS L
BR1 readth	TDSKSPQLPT DFTQFNSANC PVQTVIIIPS L--
BR8 readth	TDSKSPQLPT DFTQFNSANC PVQTVIIIPS L--
SP readthr	TDSKSPQLPT DFTQFNSANC PVQTVIIIPS L--
HB-P22 rea	TDSKSPQLPS DFTQFDSTNC PVQTVIVIPS L--
HB-P24 rea	TDSKSPQLPT DFTRFNHRNC PVQTVIVIPS L--
NL95 readt	TDSKSPQLPT DFTRFNHRNC PVQTVIVIPS LAT
FI readthr	TDSKSPQIPN DFTRFDSTKC PVQTVIVIPS LD-

Group IV replicase:

Alignment: Align Group IV replicase.

	10	20	30	40	50
Consensus	M KTA IT L KVDI FEDDIH SI ANDL A G L AE CI					
BR1 replic	MSKTASRRRE ITQLLGKVDI NFEDDIHMSI ANDLFEAYGI PKLQNAEECI					
BR8 replic	MSKTASRRRE ITQLLGKVDI NFEDDIHMSI ANDLCEAYGI PMLRDAEECI					
SP replica	MPKTASRRRE ITQLLGKVDI NFEDDIHMSI ANDLFEAYGI PKLDSAEECI					
HB-P22 rep	MSKTASHRKK ITHLLSKVDI DFEDDIHMSI ANDLFKAFGV APLTSAEQCI					
HB-P24 rep	MSKTASHKKK ITQALSKVDI NFEDDIHMSI ANDLFEAYGI APLASAEQCI					
NL95 repli	MSKTACHKKK ITQTLSKVDI NFEDDIHMSI ANDLLQACGV APLASAEQCI					
FI replica	MSKTASRRRE ITHLLGKVDI SFEDDIHLSI ANDLFRAYGV GELSSAEECI					
	60	70	80	90	100
Consensus	T FP D FR YLR EILSK H PLG TE A WEKFLAAEEG					
BR1 replic	NTAFPSLDQG ADTFRVEYLR AEILSKFDGH PLGTDTEKTA WEKFLAAEEG					
BR8 replic	NTAFPSLDQS ADTFRVEYLR AEILSKFDGH PLGINTEEAA WEKFLAAEEG					
SP replica	NTAFPSLDQG VDTFRVEYLR AEILSKFDGH PLGIDTEAAA WEKFLAAEEG					
HB-P22 rep	NTPFPDTSMT ADAFRIAYLR SEILSKYSAH PLGIDTEAVA WEKFLAAEEG					
HB-P24 rep	STPFPDTSMD ADAFRIHYLR SEILSKFSAH PLGIDTEAAA WEKFLAAEEG					
NL95 repli	NTPFPDTSMN PDEFRIQYLR SEILSKFSAH PLGIDTEAAA WEKFLAAEEG					
FI replica	NTAFPRLDQS PDTFRTEYLR SEILSKFNAH PLGIDTEAVA WEKFLAAEEG					
	110	120	130	140	150
Consensus	CR TN RL KYHDNSILS WGERVIHTAR RKILKLIGE P GDVALR					
BR1 replic	CRLTNERLSQ VKYHDNSILS WGERVIHTAR RKILKLIGES VPFGDVALRC					
BR8 replic	CRRTNERLSQ AKYHDNSILS WGERVIHTAR RKILKLIGES VPFGDVALRC					
SP replica	CRQTNERLSL VKYHDNSILS WGERVIHTAR RKILKLIGES VPFGDVALRC					
HB-P22 rep	CRQTNERLSQ AKYHDNSILS WGERVIHTAR RKILKLIGES VPLGDVALRC					
HB-P24 rep	CRQTNERLTK VKYHDNSILS WGERVIHTAR RKILKLIGET VPLGDVALRA					
NL95 repli	CRLTNERLTK VKYHDNSILS WGERVIHTAR RKILKLIGEA APLGDVALRA					
FI replica	CRLTNARLSS CKYHDNSILS WGERVIHTAR RKILKLIGES VPFGDVALRC					
	160	170	180	190	200
Consensus	RFSGGATTSV NRLHGHPSWK HACPDVTKR A KYL A K ACGD LR					
BR1 replic	RFSGGATTSV NRLHGHPSWK HACPDVTKR AFKYLQAFKR ACGDVVDLRV					
BR8 replic	RFSGGATTSV NRLHGHPSWK HACPDVTKR ALKYLQAFKR ACGDVVDLRV					
SP replica	RFSGGATTSV NRLHGHPSWK HACPDVTKR AFKYLQAFKR ACGDVVDLRV					
HB-P22 rep	RFSGGATTSV NRLHGHPSWK HACPDVTKR AFKYLQAYKM ACGDIVDLRV					
HB-P24 rep	RFSGGATTSV NRLHGHPSWK HACPDVTKR ALKYL MAYKK ACGDVVDLRV					
NL95 repli	RFSGGATTSV NRLHGHPSWK HACPDVTKR ALKYLIAYKK ACGDVVDLRV					
FI replica	RFSGGATTSV NRLHGHPSWK HACPDVTKR ALKYLLAYKK ACGDTDELRI					

	210 220 230 240 250
Consensus	EVRTSNKAV TVPKNSKTD R CIAIEPGWNM FFQLGVGAVL RDRLRLW ID
BR1 replic	NEVRTSNKAV TVPKNSKTD R CIAIEPGWNM FFQLGVGAVL RDRLRLWKID
BR8 replic	NEVRTSNKAV TVPKNSKTD R CIAIEPGWNM FFQLGVGAVL RDRLRLWKID
SP replica	NEVRTSNKAV TVPKNSKTD R CIAIEPGWNM FFQLGVGAVL RDRLRLWKID
HB-P22 rep	NEVRTSNKAV TVPKNSKTD R CIAIEPGWNM FFQLGVGAVL RDRLRLWGID
HB-P24 rep	NEVRTSNKAV TVPKNSKTD R CIAIEPGWNM FFQLGVGAVL RDRLRLWQID
NL95 repli	NEVRTSNKAV TVPKNSKTD R CIAIEPGWNM FFQLGVGAVL RDRLRLWHID
FI replica	GEVRTSNKAV TVPKNSKTD R CIAIEPGWNM FFQLGVGAVL RDRLRLWSID

	260 270 280 290 300
Consensus	LNDQS NQRL ARD S L HL AT DLSAASD SIS LVELL PP W LT
BR1 replic	LNDQSTNQRL ARDGSLNHL ATIDLSAASD SISLKLVELL LPPEWYDLLT
BR8 replic	LNDQSTNQRL ARDGSLNHL ATIDLSAASD SISLKLVELL LPPEWYDLLT
SP replica	LNDQSTNQRL ARDGSLNHL ATIDLSAASD SISLKLVELL MPPEWYDLLT
HB-P22 rep	LNDQSINQRL ARDASQLDHL ATVDLSAASD SISLKLVELL LPPDWFVGLT
HB-P24 rep	LNDQSVNQRL ARDASQLDHL ATVDLSAASD SISLRLVELL MPPAWFDLLT
NL95 repli	LNDQSVNQRL ARDASQLDHL ATVDLSAASD SISLRLVELL MPPAWFDLLT
FI replica	LNDQSLNQRL ARDASQLDHL ATVDLSAASD SISIKLVELL LPPAWFELLT

	310 320 330 340 350
Consensus	DLRSD G LP G VTYEKI SSMGNGYTFE LESLIFAA A RSVCELL D
BR1 replic	DLRSDEGVLP DGRVVITYEKI SSMGNGYTFE LESLIFAALA RSVCELLEID
BR8 replic	DLRSDQGVLP DGRVVITYEKI SSMGNGYTFE LESLIFAALA RSVCELLEID
SP replica	DLRSDEGILP DGRVVITYEKI SSMGNGYTFE LESLIFAAIA RSVCELLEID
HB-P22 rep	DLRSDQGILP DGRAVITYEKI SSMGNGYTFE LESLIFAAIA RSVCELLDLD
HB-P24 rep	DLRSDQGVLP DGRVVITYEKI SSMGNGYTFE LESLIFAALA RSVCELLDLD
NL95 repli	DLRSDQGILP DGRVVITYEKI SSMGNGYTFE LESLIFAALA RSVCELLDLD
FI replica	DLRSDQGVLP NGEVVITYEKI SSMGNGYTFE LESLIFAAIA RSVCELLDLD

	360 370 380 390 400
Consensus	QS VSVYGDD III AA LM VFEYVGF T N KKT F GPFRESCGKH
BR1 replic	QSTVSVYGDD IIIDTRAAAP LMDVFEYVGF TPNKKKTFCD GPFRESCGKH
BR8 replic	QSTVSVYGDD IIIDTRAAAP LMDVFEYVGF TPNKKKTFCD GPFRESCGKH
SP replica	QSTVSVYGDD IIIDTRAAAP LMDVFEYVGF TPNRKKTFCD GPFRESCGKH
HB-P22 rep	QSTVSVYGDD IIIDSRAAAT LMDVFEYVGF TPNRKKTFVS GPFRESCGKH
HB-P24 rep	QSTVSVYGDD IIIDSRAADV LMAVFEYVGF TPNRKKTFIK GPFRESCGKH
NL95 repli	QSTVSVYGDD IIIDSRAADV LMAVFEYVGF TPNRKKTFIK GPFRESCGKH
FI replica	QSAVSVYGDD IIIPSDAAQT LMDVFEYVGF TANRKKTFIT GPFRESCGKH

	410 420 430 440 450
Consensus	W GVDVTPF YIRRP L DMILVLN Y RWGT DG WD PR L VY KY
BR1 replic	WFQGVDPVTPF YIRRPICLA DMILVLNSIY RWGTVDGVD PRALTVYEKY
BR8 replic	WFQGVDPVTPF YIRRPICLA DMILVLNSIY RWGTVDGVD PRALTVYEKY
SP replica	WFQGVDPVTPF YIRRPICLA DMILVLNSIY RWGTVDGIWD PRALTVYEKY
HB-P22 rep	WHSQGVDPVTPF YIRRPICLV DMILVLNSIY RWGTIDGVVD PRVLPVYQKY
HB-P24 rep	WHSQGVDPVTPF YIRRPICLA DMILVLNSIY RWGTIDGVVD PRVLPVYQKY

NL95 repli	W	H	S	G	V	D	V	T	P	F	Y	I	R	R	P	I	R	C	L	A	D	M	I	L	V	L	N	S	I	Y	R	W	G	T	I	D	G	V	W	D	P	R	V	L	P	V	Y	Q	K	Y
FI replica	W	F	L	G	V	D	V	T	P	F	Y	I	R	R	P	I	R	S	L	A	D	M	I	L	V	L	N	N	L	Y	R	W	G	T	V	D	G	V	W	D	P	R	A	L	T	V	Y	Q	K	Y

	460 470 480 490 500
Consensus	LPR WRR N IPDGYGDG ALVG A TNP FV V N R YPVLVEVQ D
BR1 replic	LKLLPRNWRR NRIPDGYGDG ALVGLATTNP FVIVKNYSRL YPVLVEVQRD
BR8 replic	LKLLPRNWRR NRIPDGYGDG ALVGLATTNP FVIVKNYSRL YPVLVEVQRD
SP replica	LKLLPRNWRR NRIPDGYGDG ALVGLATTNP FVIVKNYSRL YPVLVEVQRD
HB-P22 rep	VNMLPRNWRR NTIPDGYGDG ALVGLATTNP FVIVKNFSRL YPVLVEVQKD
HB-P24 rep	VKLLPRDWRR NTIPDGYGDG ALVGLATTNP FVIVRNYSRW YPVLVEVQRD
NL95 repli	VKLLPRDWRR NTIPDGYGDG ALVGLATTNP FVIVRNYSRW YPVLVEVQRD
FI replica	VKLLPRNWRR NTIPDGYGDG ALVGSALTNP FVLVRNFQRE YPVLVEVQKD

	510 520 530 540 550
Consensus	R E G YL Y LR R R PFL D FDE PLAT LRRKTGRYK
BR1 replic	VKRSEVGSYL YALLRDRETR YSPFLRDADR TGFDEAPLAT SLRRKTGRYK
BR8 replic	VKRSEVGSYL YSLLRNRETR YSPFLRDADR TGFDEAPLAT SLRRKTGRYK
SP replica	VKRSEEGSYL YALLRDRETR YSPFLRDADR TGFDEAPLAT SLRRKTGRYK
HB-P22 rep	VKRHEYGSYL YAMLRDRETR YNPFLRVADG TGFDEAPLAT SLRRKTGRYK
HB-P24 rep	AKRHEFGSYL YALLRDREAR YNPFLRTADG SGFDETPLAT SLRRKTGRYK
NL95 repli	AKRHEFGSYL YALLRDRDAR YSPFLRVADG TGFDEAPLAT SLRRKTGRYK
FI replica	TPRSEKGAYL YHLLRDREAR HNPFLYDTDW VRFDEAPLAT RLRRKTGRYK

	560 570 580
Consensus	VAWIQDSAFI RPPY G P EVK A
BR1 replic	VAWIQDSAFI RPPYFITGIP EVKLAS-----
BR8 replic	VAWIQDSAFI RPPYFITGIP EVKLAS-----
SP replica	VAWIQDSAFI RPPYLITGIP EVKLAS-----
HB-P22 rep	VAWIQDSAFI RPPYFLTGLP EVKLAS-----
HB-P24 rep	VAWIQDSAFI RPPYFIKGIP EVKLAS-----
NL95 repli	VAWIQDSAFI RPPYFIKGIP EVKLAS-----
FI replica	VAWIQDSAFI RPPYS-TGLP EVKFARKTLV RNGKGAR

Appendix C

Group I and JS Nucleotides

Alignment: Align Group I and JS nucleotide sequences.

	10	20	30	40	50
Consensus	GGGTGGGACC	CCTTTCGGGG	TCCTGCTCAA	CTTCCTGTCTG	AGCTAAATGC	
DL1 I	
DL2 I	-----	-----	-----	-----	-----	
DL13 I	-----	-----	-----	-----	-----	
DL16 I-	
ST4 I-	
R17 I-	
J20 I-	
MS2 I-	
M12 I-	
DL52	-----	-----	-----	-----	-----	
DL54	-----	-----	-----	-----	-----	

	60	70	80	90	100
Consensus	CATTTTAAAT	GTCTTTAGCG	AGACGCTACC	WTGGCTATCG	CTGTAGGTAG	
DL1 I	A.....	
DL2 I	-----	-----	-----	A.....	
DL13 I	-----	-----	-----	A.....	
DL16 I	A.....	
ST4 I	A.....	
R17 I	A.....	
J20 I	A.....	
MS2 I	A.....	
M12 I	T.....	
DL52	-----	-----	-----	-----	-----	
DL54	-----	-----	-----	-----	-----	

	110	120	130	140	150
				ORF1		
Consensus	CCGGAATTCC	ATTCCTAGGA	GGTTTGACY	RTGCGAGCTT	TYAGTRYCT	
DL1 ITC	A.....	.C...GTC..	
DL2 ITC	A.....	.T...GTC..	
DL13 ITC	A.....	.T...GTC..	
DL16 ITC	A.....	.T...GTC..	
ST4 ICT	G.....	.T...ACC..	
R17 ICT	A.....	.T...GCC..	
J20 ITC	A.....	.T...GTC..	
MS2 ICT	G.....	.T...ACC..	
M12 ITC	A.....	.T...GTT..	
DL52	-----	-----	-----TC	A.....	.T...GTC..	
DL54	-----	-----	-----TC	A.....	.T...GTC..	

	160 170 180 190 200
Consensus	TGATMRGGAR WYRARACCT TYGTSCCBHB CRTYCGCRY YAYGCKRACG
DL1 IAA...G TCTG.G.... .T..C..GCT .G.C...ACC ..T..TG...
DL2 ICA...G TCCG.G.... .C..C..TAG .G.C...GTT ..T..GG...
DL13 ICA...G TCCG.G.... .C..C..TAG .G.C...GTT ..T..GG...
DL16 ICA...G TCCG.G.... .C..C..TAG .G.C...GTT ..T..GG...
ST4 IAG...G AACG.G.... .C..C..CTC .G.T...GTT ..C..GG...
R17 IAA...G AGCA.G.... .C..C..TTC .A.T...GTT ..C..GA...
J20 IAG...A TCCG.G.... .C..C..CTC .G.T...GTT ..T..TG...
MS2 IAG...G AACG.G.... .C..C..CTC .G.T...GTT ..C..GG...
M12 ICA...G AACG.A.... .C..G..CTC .G.T...GTT ..T..GG...
DL52CA...G TCCG.G.... .C..C..TAG .G.C...GTT ..T..GG...
DL54CA...G TCCG.G.... .C..C..TAG .G.C...GTT ..T..GG...

	210 220 230 240 250
Consensus	GBSAGRYGGA RGATAACTCR TTYTCYYTVA AATAYCGYTC GAACTGGAC Y
DL1 I	.CG..GTT.. A.....G ..C..CT.G.C..T..C
DL2 I	.GC..GTC.. A.....G ..T..CC.A.C..T..T
DL13 I	.GC..GTC.. A.....G ..T..CC.A.C..C..T
DL16 I	.GC..GTC.. A.....G ..T..CC.A.C..C..T
ST4 I	.TG..ACT.. A.....A ..C..TT.A.T..T..T
R17 I	.TG..ACC.. A.....A ..C..TT.A.T..C..T
J20 I	.GG..GTC.. G.....A ..T..CT.A.C..T..T
MS2 I	.TG..ACT.. A.....A ..C..TT.A.T..T..T
M12 I	.CG..ACT.. A.....G ..T..CC.C.T..C..C
DL52	.GC..GTC.. A.....G ..T..CC.A.C..C..T
DL54	.GC..GTC.. A.....G ..T..CC.A.C..C..T

	260 270 280 290 300
Consensus	CCBGGYCGWT TTAAYTCGAC YGGGDCYARA ACGRADCAGT GGCACTAYCC
DL1 I	..T..T..T.C..... T...G.T.G. ...A.A....T..
DL2 I	..C..T..A.T..... C...T.T.G. ...A.A....T..
DL13 I	..C..T..A.T..... C...T.T.G. ...G.A....T..
DL16 I	..C..T..A.T..... C...T.T.G. ...G.A....T..
ST4 I	..C..T..T.C..... T...G.C.A. ...A.A....C..
R17 I	..C..T..T.C..... T...G.C.G. ...A.G....T..
J20 I	..T..C..A.C..... T...A.C.G. ...A.T....T..
MS2 I	..C..T..T.C..... T...G.C.A. ...A.A....C..
M12 I	..G..T..A.C..... T...G.C.G. ...A.A....T..
DL52	..C..T..A.T..... C...T.T.G. ...G.A....T..
DL54	..C..T..A.T..... C...T.T.G. ...G.A....T..

	310320330340350
Consensus	STCYICKTAY TCDMGRGGDG CGYTNAGYGT YACDKCGRTV GATCAAGGTK
DL1 I	G..CT.T..T ..TA.A..T. ..C.T..C.. C..AT..G.GG
DL2 I	G..CC.T..T ..TA.G..T. ..C.C..T.. C..AT..G.AG
DL13 I	G..CC.T..T ..TA.G..T. ..C.C..T.. C..AT..G.AG
DL16 I	G..CC.T..T ..TA.G..T. ..C.C..T.. C..AT..G.AG
ST4 I	C..TC.G..T ..AC.G..G. ..T.A..T.. C..AT..A.AG
R17 I	C..CC.G..T ..GC.G..G. ..T.A..T.. C..GT..A.AG
J20 I	G..CC.T..C ..TA.G..A. ..C.T..T.. C..TT..G.CT
MS2 I	C..TC.G..T ..AC.G..G. ..T.A..T.. C..AT..A.AG
M12 I	C..CC.T..C ..TC.G..A. ..T.G..T.. T..TG..A.AG
DL52	G..CC.T..T ..TA.G..T. ..C.C..T.. C..AT..G.AG
DL54	G..CC.T..T ..TA.G..T. ..C.C..T.. C..AT..G.AG

	360370380390400
Consensus	CYTAYAAGCG MWSTGGGTCTC TCGTGGGGTC GYCCGTACGA GGAGAAARCY
DL1 I	.T..T..... CTC.....AC.....G.C
DL2 I	.C..C..... CTC.....AC.....G.C
DL13 I	.C..C..... CTC.....AC.....G.C
DL16 I	.C..C..... CTC.....AC.....G.C
ST4 I	.C..C..... AAG.....AC.....G.C
R17 I	.C..T..... CAG.....AC.....G.C
J20 I	.C..C..... CTC.....AT.....G.T
MS2 I	.C..C..... AAG.....AC.....G.C
M12 I	.C..T..... AAG.....GC.....A.C
DL52	.C..C..... CTC.....AC.....G.C
DL54	.C..C..... CTC.....AC.....G.C

	410420430440450
Consensus	GGTTWYGGYT TCTCNCTCGA CGCACGYTCC TGCTAYAGCC TCTTCCCTGT
DL1 IAT..C.A.....C... ..C....
DL2 ITT..C.A.....T... ..T....
DL13 ITT..C.A.....T... ..T....
DL16 ITT..C.A.....T... ..T....
ST4 ITC..C.C.....C... ..C....
R17 ITT..C.T.....C... ..C....
J20 ITT..C.G.....T... ..C....
MS2 ITC..C.C.....C... ..C....
M12 ITT..T.A.....C... ..C....
DL52TT..C.A.....T... ..T....
DL54TT..C.A.....T... ..T....

	460 470 480 490 500
Consensus	HAGYCARAAY HTGACTTACA TYGAAGTGCC GCAGAACGTT GCGAAYCGGG
DL1 I	T..T..G..C C..... .T.....
DL2 I	T..T..G..T C..... .C.....
DL13 I	T..T..G..T C..... .C.....
DL16 I	T..T..G..T C..... .C.....
ST4 I	A..C..G..C T..... .C.....
R17 I	A..C..G..C T..... .C.....
J20 I	C..T..A..T A..... .C.....
MS2 I	A..C..A..C T..... .C.....
M12 I	A..C..G..C C..... .C.....
DL52	T..T..G..T C..... .C.....
DL54	T..T..G..T C..... .C.....

	510 520 530 540 550
Consensus	CKWCGACCGA AGTCCTGCAR AAGGTYACYC ARGGNAAYTT YAACCTTGGB
DL1 I	.GT.....
DL2 I	.GT.....
DL13 I	.GT.....
DL16 I	.GT.....
ST4 I	.GT.....
R17 I	.GT.....
J20 I	.GT.....
MS2 I	.GT.....
M12 I	.TA.....
DL52	.GT.....
DL54	.GT.....

	560 570 580 590 600
Consensus	GTNGCYITWG CAGAGGCVAG RTCGACAGCC TCACAACCTCG CGACGCAAAC
DL1 I	..A..CT.A.G.. G.....
DL2 I	..G..CC.A.C.. G.....
DL13 I	..G..CC.A.C.. G.....
DL16 I	..G..CC.A.C.. G.....
ST4 I	..T..TT.A.C.. G.....
R17 I	..C..CC.A.C.. A.....
J20 I	..A..TT.A.C.. G.....
MS2 I	..T..TT.A.C.. G.....
M12 I	..G..CC.T.A.. G.....
DL52	..G..CC.A.C.. G.....
DL54	..G..CC.A.C.. G.....

	610620630640650
Consensus	CATTGCGCTC GTGAAGGCGT ACACTGCCGC TCGTCGCGGY AAYTGGCGCC
DL1 IC ..C.....
DL2 IC ..T.....
DL13 IC ..T.....
DL16 IC ..T.....
ST4 IT ..T.....
R17 IT ..T.....
J20 IT ..T.....
MS2 IT ..T.....
M12 IT ..T.....
DL52C ..T.....
DL54C ..T.....

	660670680690700
Consensus	AGVCGSTCCG CTAYCTYGCC CTWAACGAAG AYCGRARTT YCGRTCRAAA
DL1 I	..A..C.... ..T..C... ..T..... .C..A..A.. T..G..A...
DL2 I	..A..C.... ..T..T... ..A..... .T..G..G.. C..A..G...
DL13 I	..A..C.... ..T..T... ..A..... .T..G..G.. C..A..G...
DL16 I	..A..C.... ..T..T... ..A..... .T..G..G.. C..A..G...
ST4 I	..G..C.... ..C..T... ..A..... .T..A..G.. T..A..A...
R17 I	..G..C.... ..C..C... ..A..... .T..A..A.. T..A..A...
J20 I	..G..C.... ..T..T... ..A..... .T..G..A.. C..G..A...
MS2 I	..G..C.... ..C..T... ..A..... .T..A..G.. T..A..A...
M12 I	..C..G.... ..C..C... ..A..... .T..A..G.. T..A..A...
DL52	..A..C.... ..T..T... ..A..... .T..G..G.. C..A..G...
DL54	..A..C.... ..T..T... ..A..... .T..G..G.. C..A..G...

	710720730740750
Consensus	CACGTGGCVG GYAGRTGGTT GGAGTTGCAG TTCGGHTGGY TACCRCTHAT
DL1 IG. .C..G.....C...TG..C..
DL2 IA. .C..G.....C...CG..C..
DL13 IA. .C..G.....C...CG..C..
DL16 IA. .C..G.....C...CG..C..
ST4 IC. .C..G.....T...TA..A..
R17 IC. .C..G.....A...TA..A..
J20 IA. .T..A.....T...CG..T..
MS2 IC. .C..G.....T...TA..A..
M12 IC. .C..G.....C...TA..T..
DL52A. .C..G.....C...CG..C..
DL54A. .C..G.....C...CG..C..

	760770780790800
Consensus	GAGYGATATC CARGGYGCAT AYGAGATGCT TACGAAGGTT CAYCTTCAAG
DL1 I	...C..... ..G..C.... .T..... ..C.....
DL2 I	...C..... ..G..C.... .T..... ..C.....
DL13 I	...C..... ..G..C.... .T..... ..C.....
DL16 I	...C..... ..G..C.... .T..... ..C.....
ST4 I	...T..... ..G..T.... .T..... ..C.....
R17 I	...T..... ..A..T.... .T..... ..C.....
J20 I	...C..... ..A..T.... .T..... ..T.....
MS2 I	...T..... ..G..T.... .T..... ..C.....
M12 I	...T..... ..G..C.... .C..... ..C.....
DL52	...C..... ..G..C.... .T..... ..C.....
DL54	...C..... ..G..C.... .T..... ..C.....

	810820830840850
Consensus	AGTTTCTYCC TATGAGRGCC GTRCGNCARG TNGGYACTAA CRTYAAGTTA
DL1 IC..A... ..G..C..G. .A..C..... .G.C.....
DL2 IT..G... ..A..G..A. .G..C..... .A.T.....
DL13 IT..G... ..A..G..A. .G..C..... .A.T.....
DL16 IT..G... ..A..G..A. .G..C..... .A.T.....
ST4 IT..A... ..A..T..G. .C..T..... .A.C.....
R17 IT..A... ..A..T..A. .T..T..... .A.T.....
J20 IT..A... ..A..A..G. .G..T..... .G.T.....
MS2 IT..A... ..A..T..G. .C..T..... .A.C.....
M12 IC..A... ..A..T..G. .C..C..... .A.T.....
DL52T..G... ..A..G..A. .G..C..... .A.T.....
DL54T..G... ..A..G..A. .G..C..... .A.T.....

	860870880890900
Consensus	RATGGCCGYT TKKCGTATCC AGCTGCAAAC TWCCARACWA CGTGCAACAT
DL1 I	G.....CC .GT..... ..A...A..T.
DL2 I	G.....TT .GT..... ..A...G..T.
DL13 I	G.....TT .GT..... ..A...G..T.
DL16 I	G.....TT .GT..... ..A...G..T.
ST4 I	A.....TC .GT..... ..T...G..A.
R17 I	G.....CT .GG..... ..T...G..A.
J20 I	G.....CT .GT..... ..A...G..T.
MS2 I	G.....TC .GT..... ..T...G..A.
M12 I	G.....CC .TT..... ..A...G..A.
DL52	G.....TT .GT..... ..A...G..T.
DL54	G.....TT .GT..... ..A...G..T.

	910920930940950
Consensus	ATCRCGACGW ATCGTGATAT GGTTTTACAT AAACGATGCA CGWTTGGCHT
DL1 I	...G.....TT.....T.
DL2 I	...A.....AT.....C.
DL13 I	...A.....AT.....C.
DL16 I	...A.....AT.....C.
ST4 I	...G.....TT.....A.

R17 I	...	A	...	T	T	A
J20 I	...	G	...	A	T	C
MS2 I	...	G	...	T	T	A
M12 I	...	G	...	T	A	C
DL52	...	A	...	A	T	C
DL54	...	A	...	A	T	C
<div style="text-align: center;"> <div style="display: flex; justify-content: space-around; width: 100%;"> 960 970 980 990 1000 </div> </div>												
Consensus	GGTTGTCGTC	TCTRGGTATC	TTGAACCCAC	TAGGTATAGT	GTGGGAAAAG							
DL1 I	A
DL2 I	A
DL13 I	A
DL16 I	A
ST4 I	A
R17 I	A
J20 I	A
MS2 I	A
M12 I	G
DL52	A
DL54	A
<div style="text-align: center;"> <div style="display: flex; justify-content: space-around; width: 100%;"> 1010 1020 1030 1040 1050 </div> </div>												
Consensus	GTGCCYTTCT	CATTCGTTGT	CGACTGGCTC	CTDCCTGTDG	GDAACATGCT							
DL1 I	T	T	A	..	A
DL2 I	T	T	T	..	G
DL13 I	T	T	T	..	G
DL16 I	T	T	T	..	G
ST4 I	T	A	A	..	T
R17 I	T	A	A	..	T
J20 I	T	T	A	..	A
MS2 I	T	A	A	..	T
M12 I	C	G	G	..	G
DL52	T	T	T	..	G
DL54	T	T	T	..	G
<div style="text-align: center;"> <div style="display: flex; justify-content: space-around; width: 100%;"> 1060 1070 1080 1090 1100 </div> </div>												
Consensus	MGAGGGCCTH	ACRGCYCCMG	TDGGATGYTC	YTACATGTCD	GGRACAGTTA							
DL1 I	C.....	..	A	..	G	..	C	..	C	..	A
DL2 I	C.....	..	C	..	A	..	C	..	C	..	A
DL13 I	C.....	..	C	..	A	..	C	..	C	..	A
DL16 I	C.....	..	C	..	A	..	C	..	C	..	A
ST4 I	C.....	..	T	..	G	..	C	..	C	..	A
R17 I	C.....	..	T	..	G	..	T	..	C	..	A
J20 I	C.....	..	T	..	A	..	C	..	C	..	A
MS2 I	C.....	..	T	..	G	..	C	..	C	..	A
M12 I	A.....	..	T	..	G	..	T	..	A	..	A
DL52	C.....	..	C	..	A	..	C	..	C	..	A
DL54	C.....	..	C	..	A	..	C	..	C	..	A

	1110 1120 1130 1140 1150
Consensus	CTGACGTAAT AACGGGTGAG TCCATCATAA GCGTYGACGC TCCCTAYGGG
DL1 IT.....T...
DL2 IC.....T...
DL13 IC.....T...
DL16 IC.....T...
ST4 IT.....C...
R17 IT.....T...
J20 IT.....T...
MS2 IT.....C...
M12 IT.....C...
DL52C.....T...
DL54C.....T...

	1160 1170 1180 1190 1200
Consensus	TGGACDGTGG AKAGACAGGG CACTGCTAAG GCCCADRTYT CRGCCATGCA
DL1 IA......G.....GA.C..A.....
DL2 IG......T.....AG.C..A.....
DL13 IG......T.....AG.C..A.....
DL16 IG......T.....AG.C..A.....
ST4 IT......G.....AA.C..A.....
R17 IT......G.....TG.T..A.....
J20 IT......G.....GA.C..G.....
MS2 IT......G.....AA.C..A.....
M12 IT......G.....AG.C..A.....
DL52G......T.....AG.C..A.....
DL54G......T.....AG.C..A.....

	1210 1220 1230 1240 1250
Consensus	TCGAGGGGTR CARTCCGTAT GSCCAACWAC TGGCGYRTAC GTRAARTCWC
DL1 IA..G......G.....A..TA...A..G..A.
DL2 IG..A......G.....A..TA...G..G..A.
DL13 IG..A......G.....A..TA...G..G..A.
DL16 IG..A......G.....A..TA...G..G..A.
ST4 IA..A......G.....A..CG...A..G..T.
R17 IA..A......G.....A..CA...A..G..T.
J20 IG..A......G.....A..TA...G..A..A.
MS2 IA..A......G.....A..CG...A..G..T.
M12 IA..A......C.....T..TA...A..G..T.
DL52G..A......G.....A..TA...G..G..A.
DL54G..A......G.....A..TA...G..G..A.

	1260 1270 1280 1290 1300
Consensus	CYTTYTCGAT KGTCCAYACY TTAGAYGCGT TRGCATTAAT CAGGCAACGG
DL1 I	.T..C.....G.....C..T.....T......G.....
DL2 I	.T..C.....G.....T..C.....C......G.....
DL13 I	.T..C.....G.....T..C.....C......G.....
DL16 I	.T..C.....G.....T..C.....C......G.....
ST4 I	.T..C.....G.....T..C.....T......A.....

R17 I	.T..C.....	G.....T..CT....	.A.....
J20 I	.T..C.....	T.....T..CT....	.A.....
MS2 I	.T..C.....	G.....T..CT....	.A.....
M12 I	.C..T.....	G.....T..CT....	.G.....
DL52	.T..C.....	G.....T..CC....	.G.....
DL54	.T..C.....	G.....T..CC....	.G.....

		STOP 1		ORF2	
				
	1310	1320	1330	1340	1350
Consensus	CTCTYWARAT	AGAGCCCTCA	ACCGGAGTTY	GAAGCATGGC	TTCTAACTTT
DL1 ICA.G..C
DL2 ITA.G..C
DL13 ITA.G..C
DL16 ITA.G..C
ST4 ICT.G..T
R17 ICT.A..T
J20 ICA.G..T
MS2 ICT.G..T
M12 ICT.A..T
DL52TA.G..C
DL54TA.G..C

				
	1360	1370	1380	1390	1400
Consensus	ACTCAGTTYG	TTCTCGTCGA	CAATGGCGGA	ACYGGYGACG	TGACTGTCGC
DL1 IC.T..T....
DL2 IC.T..C....
DL13 IC.T..C....
DL16 IC.T..C....
ST4 IC.T..C....
R17 IT.T..C....
J20 IT.C..C....
MS2 IC.T..C....
M12 IT.T..C....N.
DL52C.T..C....
DL54C.T..C....

				
	1410	1420	1430	1440	1450
Consensus	BCCAAGCAAC	TTCGCTAACG	GGGTCGCTGA	ATGGATYAGC	TCYAACTCVC
DL1 I	C.....T...	..T.....G.
DL2 I	T.....T...	..T.....A.
DL13 I	T.....T...	..T.....A.
DL16 I	T.....T...	..T.....A.
ST4 I	C.....C...	..T.....G.
R17 I	T.....C...	..T.....G.
J20 I	T.....C...	..T.....A.
MS2 I	C.....C...	..T.....G.
M12 I	G.....C...	..C.....C.
DL52	T.....T...	..T.....A.
DL54	T.....T...	..T.....A.

	14601470148014901500
Consensus	GYTCWCARGC TTACAAAGTA ACYTGTAGYG TKCGTCAGAG CTCTGCGCAG
DL1 I	.C..T..G.. ..C.....C. .T.....
DL2 I	.T..T..G.. ..C.....C. .T.....
DL13 I	.T..T..G.. ..C.....C. .T.....
DL16 I	.T..T..G.. ..C.....C. .T.....
ST4 I	.T..A..G.. ..C.....C. .T.....
R17 I	.C..A..G.. ..C.....C. .T.....
J20 I	.C..T..G.. ..C.....T. .T.....
MS2 I	.T..A..G.. ..C.....C. .T.....
M12 I	.C..A..A.. ..T.....C. .G.....
DL52	.T..T..G.. ..C.....C. .T.....
DL54	.T..T..G.. ..C.....C. .T.....

	15101520153015401550
Consensus	AAYCGCAART ACACYATYAA RGTYGARGTR CCDAARGTGG CWACYCARAC
DL1 I	..T.....G.T..C.. G..C..A..G ..G..A.... .T..C..G..
DL2 I	..C.....G.C..T.. G..T..G..A ..A..A.... .T..C..A..
DL13 I	..C.....G.C..T.. G..T..G..A ..A..A.... .T..C..A..
DL16 I	..C.....G.C..T.. G..T..G..A ..A..A.... .T..C..A..
ST4 I	..T.....A.C..C.. A..C..G..G ..T..A.... .A..C..G..
R17 I	..T.....A.C..T.. A..C..G..G ..T..G.... .A..T..G..
J20 I	..C.....G.C..C.. A..C..A..G ..A..A.... .T..T..G..
MS2 I	..T.....A.C..C.. A..C..G..G ..T..A.... .A..C..G..
M12 I	..C.....A.C..C.. G..C..G..G ..G..A.... .A..C..A..
DL52	..C.....G.C..T.. G..T..G..A ..A..A.... .T..C..A..
DL54	..C.....G.C..T.. G..T..G..A ..A..A.... .T..C..A..

	15601570158015901600
Consensus	YGTYYGGYGGY GTASAGCTTC CTGTAGCCGC RTGGCGTTTCG TAYYTRAATA
DL1 I	C..T..T..C ...C..... G..... ..TC.G....
DL2 I	C..C..T..T ...C..... A..... ..CT.G....
DL13 I	C..C..T..T ...C..... A..... ..CT.G....
DL16 I	C..C..T..T ...C..... A..... ..CT.G....
ST4 I	T..T..T..T ...G..... A..... ..CT.A....
R17 I	T..T..T..T ...G..... A..... ..CT.A....
J20 I	C..C..T..C ...C..... A..... ..CT.A....
MS2 I	T..T..T..T ...G..... A..... ..CT.A....
M12 I	C..T..C..T ...G..... A..... ..CC.G....
DL52	C..C..T..T ...C..... A..... ..CT.G....
DL54	C..C..T..T ...C..... A..... ..CT.G....


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      ....|....| ....|....| ....|....| ....|....| ....|....|
      1610      1620      1630      1640      1650
Consensus  TGGAAAYTRAC YATTCCDATY TTCGCDACGA AYKMCGAYTG CGMGCTWATT
DL1 I      .....T.G.. T.....A..C .....A.... .CGA...C.. ..C...A...
DL2 I      .....T.A.. T.....T..T .....A.... .CGA...T.. ..C...T...
DL13 I     .....T.A.. T.....T..T .....A.... .CGA...T.. ..C...T...
DL16 I     .....T.A.. T.....T..C .....A.... .CGA...T.. ..C...T...
ST4 I      .....C.A.. C.....A..T .....T.... .TTC...C.. ..A...T...
R17 I      .....T.A.. T.....A..T .....T.... .CTC...T.. ..A...T...
J20 I      .....T.A.. T.....A..T .....G.... .CGA...T.. ..C...T...
MS2 I      .....C.A.. C.....A..T .....T.... .TTC...C.. ..A...T...
M12 I      .....T.A.. T.....G..T .....T.... .CTC...T.. ..C...T...
DL52       .....T.A.. T.....T..T .....A.... .CGA...T.. ..C...T...
DL54       .....T.A.. T.....T..T .....A.... .CGA...T.. ..C...T...

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ORF3

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      ....|....| ....|....| ....|....| ....|....| ....|....|
      1660      1670      1680      1690      1700
Consensus  GTYAARGCRA TGCAAGGTCT CCTRAAAGAT GGAAACCCRA TYCCYTCRGC
DL1 I      ..T..G..G. ......... ..A..... ..G.... .C..C..A..
DL2 I      ..T..G..G. ......... ..A..... ..G.... .C..C..A..
DL13 I     ..T..G..G. ......... ..A..... ..G.... .C..C..A..
DL16 I     ..T..G..G. ......... ..A..... ..G.... .C..C..A..
ST4 I      ..T..G..A. ......... ..A..... ..G.... .T..C..A..
R17 I      ..T..G..A. ......... ..A..... ..G.... .T..C..A..
J20 I      ..T..G..G. ......... ..A..... ..A.... .C..C..A..
MS2 I      ..T..G..A. ......... ..A..... ..G.... .T..C..A..
M12 I      ..C..A..A. ......... ..G..... ..G.... .T..T..G..
DL52       ..T..G..G. ......... ..A..... ..G.... .C..C..A..
DL54       ..T..G..G. ......... ..A..... ..G.... .C..C..A..

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STOP 2

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      ....|....| ....|....| ....|....| ....|....| ....|....|
      1710      1720      1730      1740      1750
Consensus  AATCGCAGCM AACTCCGGM A TCTAYTAATA GAYDYCGKCC ATTCMAACAT
DL1 I      .....A .....A. ....C..... .TTT..G.. ....A.....
DL2 I      .....A .....A. ....C..... .TTC..T.. ....C.....
DL13 I     .....A .....A. ....C..... .TTC..T.. ....C.....
DL16 I     .....A .....A. ....C..... .TTC..T.. ....C.....
ST4 I      .....A .....C. ....C..... .CGC..G.. ....A.....
R17 I      .....A .....C. ....C..... .TGC..G.. ....A.....
J20 I      .....C .....A. ....T..... .TTC..G.. ....C.....
MS2 I      .....A .....C. ....C..... .CGC..G.. ....A.....
M12 I      .....A .....C. ....C..... .TAC..G.. ....A.....
DL52       .....A .....A. ....C..... .TTC..T.. ....C.....
DL54       .....A .....A. ....C..... .TTC..T.. ....C.....

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ORF4

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      ....|....| ....|....| ....|....| ....|....| ....|....|
      1760      1770      1780      1790      1800
Consensus  GAGGATTACC CATGTCGAAG ACAACAAAGA AGTTCAACTC TTTATGTATT
DL1 I      .....
DL2 I      .....
DL13 I     .....
DL16 I     .....
ST4 I      .....

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R17 I
J20 I
MS2 I
M12 IR
DL52
DL54

	18101820183018401850
Consensus	GATCTTCCTY GCGATCTTTC TCTCGAAATT TACCAATCAA TTGCTTCTGT
DL1 IC.....
DL2 IC.....
DL13 IC.....
DL16 IT.....
ST4 IC.....
R17 IC.....
J20 IC.....
MS2 IC.....
M12 IC.....
DL52C.....
DL54C.....

	18601870188018901900
Consensus	CGCTACTGGA WCGGGTRATC CGCACAGTRR MGACTTTWCR GCAATTGCTT
DL1 IA.....G... ..AG A.....A.A
DL2 IA.....G... ..AG A.....A.A
DL13 IA.....G... ..AG A.....A.A
DL16 IA.....G... ..AG A.....A.A
ST4 IA.....G... ..GA C.....A.A
R17 IA.....A... ..GA C.....A.A
J20 IA.....G... ..AG A.....A.G
MS2 IA.....G... ..GA C.....A.A
M12 IT.....A... ..GA C.....T.A
DL52A.....G... ..AG A.....A.A
DL54A.....G... ..AG A.....A.A

	STOP 3
		19101920193019401950
Consensus	ACYTAAGRGA YGARTTGCTH ACWAAGCAYC CVWCNTTRGG HWMYGGTAAT	
DL1 I	..C....G.. C..G....A ..T....T. .CT.C..A.. CAAT.....	
DL2 I	..C....A.. C..A....A ..T....T. .GT.C..A.. AAAT.....	
DL13 I	..C....A.. C..A....A ..T....T. .GT.C..A.. AAAT.....	
DL16 I	..C....A.. C..A....A ..T....T. .GT.C..A.. AAAT.....	
ST4 I	..T....G.. C..A....C ..A....T. .GA.C..A.. TTCT.....	
R17 I	..C....G.. C..A....T ..A....T. .GA.T..A.. CTCT.....	
J20 I	..C....A.. C..G....C ..A....T. .GT.T..A.. AAAT.....	
MS2 I	..T....G.. C..A....C ..A....T. .GA.C..A.. TTCT.....	
M12 I	..C....G.. C..G....T ..T....C. .GA.G..A.. TTCC.....	
DL52	..C....A.. C..A....A ..T....T. .GT.C..A.. AAAT.....	
DL54	..C....A.. T..G....T ..T....T. .AT.A..G.. CAAT.....	

	19601970198019902000
Consensus	GACGAGGCGA CCCGYCGNRC YYTAGCTATY GCTAAGCTNC GGGAGGCGAA
DL1 IC..AG. TT.....TT.
DL2 IC..GG. TC.....CC.
DL13 IC..GG. TC.....CC.
DL16 IC..GG. TC.....CC.
ST4 IT..TA. CT.....CA.
R17 IT..CA. CT.....TG.
J20 IC..GG. TT.....CC.
MS2 IT..TA. CT.....CA.
M12 IC..TG. TT.....TA.
DL52C..GG. TC.....CC.
DL54C..GG. TC.....TC.

	20102020203020402050
Consensus	TGRDSRDYGY GGYCAGATHA AYAGRGARGG TTTCTTACAY GAYAAATCCT
DL1 I	..AACGAT.T ..C.....C. .T..A..G..T ..C.....
DL2 I	..AGCGGT.C ..C.....T. .T..G..A..C ..T.....
DL13 I	..AGCGGT.C ..C.....T. .T..G..A..C ..T.....
DL16 I	..AGCGGT.C ..C.....T. .T..G..A..C ..T.....
ST4 I	..ATCGGT.C ..T.....A. .T..A..A..T ..C.....
R17 I	..ATCGGT.C ..C.....A. .T..A..A..T ..C.....
J20 I	..AACGAT.T ..C.....T. .C..A..A..T ..C.....
MS2 I	..GTGATC.C ..T.....A. .T..A..A..T ..C.....
M12 I	..ATCGAT.T ..C.....T. .C..A..A..T ..C.....
DL52	..AGCGGT.C ..C.....T. .T..G..A..C ..T.....
DL54	..AGCGAT.T ..C.....T. .T..A..A..T ..C.....

	20602070208020902100
Consensus	TRTCRTGGGA TCCGGATGTT TTACAAACCA GCATCCGTAG CCTWATHGGC
DL1 I	.G..A.....T..T...
DL2 I	.A..G.....T..T...
DL13 I	.A..G.....T..T...
DL16 I	.A..G.....T..T...
ST4 I	.G..A.....T..T...
R17 I	.G..A.....T..C...
J20 I	.G..G.....T..T...
MS2 I	.G..A.....T..T...
M12 I	.G..G.....A..A...
DL52	.A..G.....T..T...
DL54	.G..A.....T..T...

	21102120213021402150
Consensus	AAYCTYCTCT CTGGYTAYCR HTCCTCGTTG TTTGGGCAAT GCACGTTYTC
DL1 I	..C..T....C..T.G T.....C..
DL2 I	..C..T....C..C.G C.....C..
DL13 I	..C..T....C..C.G C.....C..
DL16 I	..C..T....C..C.G C.....C..
ST4 I	..C..C....C..C.G A.....C..

R17 I	..T..T....C..C.A	A.....C..
J20 I	..C..T....C..T.G	T.....T..
MS2 I	..C..C....C..C.G	A.....C..
M12 I	..C..C....T..C.G	A.....C..
DL52	..C..T....C..C.G	C.....C..
DL54	..C..C....C..T.G	T.....C..

	
	2160 2170 2180 2190 2200	
Consensus	CAACGGTGCY YCDATGGGGC ACAAGTTGCA GGATGCAGCG CCTTACAAGA	
DL1 IC T.G.....	
DL2 IC T.A.....	
DL13 IC T.A.....	
DL16 IC T.A.....	
ST4 IC T.T.....	
R17 IC T.T.....	
J20 IC T.T.....	
MS2 IT C.T.....	
M12 IC T.G.....	
DL52C T.A.....	
DL54C T.G.....	

	
	2210 2220 2230 2240 2250	
Consensus	AGTTCGCTGA ACAAGCAACC GTTACCCCCC GCGCTCTRAG AGCGGCNYTR	
DL1 IA..CC.A
DL2 IG..AC.A
DL13 IG..AC.A
DL16 IG..AC.A
ST4 IG..TC.A
R17 IG..TC.A
J20 IG..GT.A
MS2 IG..TC.A
M12 IA..CC.G
DL52G..AC.A
DL54G..TC.A

	
	2260 2270 2280 2290 2300	
Consensus	YTGGTCMGAG ACCARTGTGY GCCGTGGATY AGACACGCGG TCCRCTAYAR	
DL1 I	C.....C... ..G....TT ..G...C.A
DL2 I	C.....C... ..G....TC ..G...C.A
DL13 I	C.....C... ..G....TC ..G...C.A
DL16 I	C.....C... ..G....TC ..G...C.A
ST4 I	T.....C... ..A....CC ..G...T.A
R17 I	T.....C... ..A....CC ..A...T.A
J20 I	C.....A... ..G....CC ..G...T.A
MS2 I	T.....C... ..A....CC ..G...T.A
M12 I	C.....A... ..G....CT ..G...C.A
DL52	C.....C... ..G....TC ..G...C.A
DL54	C.....C... ..G....CC ..A...C.G

	23102320233023402350
Consensus	YGARTCATAT RAATTTAGRC TCGTYGTAGG GAACGGAGTG TTYACAGTTC
DL1 I	T..G..... G.....G.T..... ..T.....
DL2 I	C..G..... G.....G.C..... ..T.....
DL13 I	C..G..... G.....G.C..... ..T.....
DL16 I	C..G..... G.....G.C..... ..T.....
ST4 I	C..G..... G.....G.T..... ..T.....
R17 I	C..G..... G.....G.T..... ..T.....
J20 I	C..A..... A.....A.T..... ..T.....
MS2 I	C..G..... G.....G.T..... ..T.....
M12 I	C..G..... G.....G.T..... ..C.....
DL52	C..G..... G.....G.C..... ..T.....
DL54	C..G..... G.....G.C..... ..C.....

	23602370238023902400
Consensus	CGAAGAATAA TAAATAGAT CGGGCTGCTT GYAARGAGCC HGATRYGAAY
DL1 IC. .T..G.... C...AT...T
DL2 IC. .T..G.... C...AT...T
DL13 IC. .T..G.... C...AT...T
DL16 IC. .T..G.... C...AT...T
ST4 IC. .T..G.... T...AT...T
R17 IC. .T..G.... T...AT...T
J20 IC. .T..G.... C...AT...T
MS2 IC. .T..G.... T...AT...T
M12 IT. .C..A.... C...AT...T
DL52C. .C..G.... T...GC...C
DL54C. .T..G.... A...GC...C

	24102420243024402450
Consensus	ATGTACCTYC AGAAAGGRGT HGGHGVYTTY ATHMGVCGYC GBCTCMRHTC
DL1 IT.G.. T..C.CC..C ..TA.A..C. .C...AAA..
DL2 IC.A.. T..C.CC..C ..AA.G..T. .C...AAA..
DL13 IC.A.. T..C.CC..C ..AA.G..T. .C...AAA..
DL16 IC.A.. T..C.CC..C ..AA.G..T. .C...AAA..
ST4 IC.G.. C..T.CC..T ..CA.A..C. .G...AAA..
R17 IC.G.. C..C.CT..T ..TA.A..C. .G...AAA..
J20 IT.A.. T..C.CT..T ..AC.G..T. .T...AGA..
MS2 IC.G.. C..T.CT..C ..CA.A..C. .G...AAA..
M12 IT.A.. A..A.CC..C ..CA.G..C. .T...AAA..
DL52T.A.. T..C.AT..T ..AC.C..T. .T...CGC..
DL54T.G.. T..T.GC..C ..AC.C..C. .C...CGT..

	24602470248024902500
Consensus	YRTYGGTATM GAYCTGAATG ATCARWCGAT CAAYCARCKT YTDGCWMARC
DL1 I	TG.C....A ..C.....AA.... ...T..G.G. C.A..TC.A.
DL2 I	TG.C....A ..C.....AA.... ...T..G.G. T.A..TC.A.
DL13 I	TG.C....A ..C.....AA.... ...T..G.G. T.A..TC.A.
DL16 I	TG.C....A ..C.....AA.... ...T..G.G. T.A..TC.A.

ST4 I	CG.T.....A ..T.....AT....	...C..G.T.	C.G..TC.G.
R17 I	CG.T.....A ..C.....GT....	...C..G.G.	C.A..TC.G.
J20 I	CG.T.....A ..C.....AA....	...C..G.G.	C.G..TC.A.
MS2 I	CG.T.....A ..C.....AT....	...C..G.G.	C.G..TC.G.
M12 I	CG.T.....A ..C.....AA....	...C..G.G.	C.G..TC.A.
DL52	TA.C.....C ..C.....AA....	...T..G.G.	C.T..AA.A.
DL54	TA.C.....C ..C.....AA....	...T..A.G.	C.T..AA.A.

	2510	2520	2530	2540	2550
Consensus	WNGGCAGYRY	MGATGGHTCD	YTDGCRACKA	TWGAYYTATC	GTCNGCNTCB
DL1 I	AG.....CAT	A.....T..T	T.A..G..G.	.A..CC....	...A..T..G
DL2 I	AG.....CAT	A.....C..T	T.A..A..G.	.A..TT....	...G..T..T
DL13 I	AG.....CAT	A.....C..T	T.A..A..G.	.A..TT....	...G..T..T
DL16 I	AG.....CAT	A.....C..T	T.A..A..G.	.A..TT....	...G..T..T
ST4 I	AG.....CGT	A.....T..G	C.T..G..G.	.A..CT....	...T..A..C
R17 I	AA.....CGC	A.....T..G	C.T..G..G.	.A..CT....	...C..G..C
J20 I	AA.....CGT	C.....C..A	T.G..G..G.	.A..CT....	...G..C..T
MS2 I	AG.....CGT	A.....T..G	C.T..G..G.	.A..CT....	...T..A..C
M12 I	AA.....TAT	A.....A..G	T.A..G..T.	.A..CT....	...T..A..C
DL52	TT.....CAT	C.....T..T	T.A..G..G.	.T..TT....	...T..T..T
DL54	TC.....TAT	C.....T..T	T.A..G..G.	.A..TT....	...T..T..T

	2560	2570	2580	2590	2600
Consensus	GAYTCYATHT	CBGAYCGCCT	NGTKTGRRN	TTYCTYCCAY	CKSARHTRTA
DL1 I	..T..C..C.	.T..C.....	G..G...AGC	..T..C...C	.TG.GC.A..
DL2 I	..T..T..C.	.T..C.....	G..G...AGT	..C..C...C	.TG.GT.A..
DL13 I	..T..T..C.	.T..C.....	G..G...AGT	..C..C...C	.TG.GT.A..
DL16 I	..T..T..C.	.T..C.....	G..G...AGT	..C..C...C	.TG.GT.A..
ST4 I	..T..C..C.	.C..T.....	G..G...AGT	..T..C...C	.TG.GC.A..
R17 I	..T..C..T.	.C..C.....	A..G...AAT	..C..T...C	.TG.GC.A..
J20 I	..C..C..C.	.T..C.....	G..G...AGT	..C..C...C	.TG.GC.A..
MS2 I	..T..C..C.	.C..T.....	G..G...AGT	..T..C...C	.TG.GC.A..
M12 I	..T..C..C.	.T..C.....	G..G...AGT	..C..C...C	.TG.GC.A..
DL52	..T..C..A.	.G..C.....	C..T...GAA	..C..C...T	.GC.AA.G..
DL54	..C..C..A.	.G..C.....	T..T...GAG	..T..C...C	.GC.AA.G..

	2610	2620	2630	2640	2650
Consensus	YKCRtayCTB	KMKMRWATYC	GYTCVYMMYR	HGGAATCRTW	GAYGGNSRKR
DL1 I	TT.A..T..C	GATCGT..C.	.C..CCACTA	C.....G.A	..T..CGAGA
DL2 I	TT.A..T..C	GATCGT..C.	.C..CCACTA	C.....G.A	..T..GGAGA
DL13 I	TT.A..T..C	GATCGT..C.	.C..CCACTA	C.....G.A	..T..GGAGA
DL16 I	TT.A..T..C	GATCGT..C.	.C..CCACTA	C.....G.A	..T..GGAGA
ST4 I	TT.A..T..C	GATCGT..C.	.C..ACACTA	C.....G.A	..T..CGAGA
R17 I	TT.A..T..T	GATCGT..C.	.C..GCACTA	C.....G.A	..T..CGAGA
J20 I	TT.A..T..C	GATCGT..C.	.C..CCACTA	T.....A.A	..T..AGAGA
MS2 I	TT.A..T..C	GATCGT..C.	.C..ACACTA	C.....G.A	..T..CGAGA
M12 I	CT.A..T..C	GATCGT..C.	.C..CCACTA	C.....G.A	..T..TGAGA
DL52	CG.G..C..G	TCGAAA..T.	.T..GTCACG	A.....G.T	..C..ACGTG
DL54	TG.G..C..G	TCTAAA..T.	.T..GCCACG	A.....G.T	..C..ACGTG

	2660 2670 2680 2690 2700
Consensus	YRRTMSRVTG GSAMCTATTT TCCACDATGG GWAAYGGDDT YACNTTYGAR
DL1 I	CGA.ACGG.. .G.A..... .A..T..G.. C..T..C..A
DL2 I	CGA.ACGA.. .G.A..... .A..T..G.. C..T..C..A
DL13 I	CGA.ACGA.. .G.A..... .A..T..G.. C..T..C..A
DL16 I	CGA.ACGA.. .G.A..... .A..T..G.. C..T..C..A
ST4 I	CGA.ACGA.. .G.A..... .A..T..G.. C..G..T..G
R17 I	CGA.ACGA.. .G.A..... .G..... .A..T..G.. T..A..T..G
J20 I	CGA.ACGA.. .G.A..... .A..T..G.. C..T..T..A
MS2 I	CGA.ACGA.. .G.A..... .A..T..G.. C..A..T..G
M12 I	CGA.ACGA.. .G.A..... .A..C..G.. C..T..T..G
DL52	TAG.CGAC.. .C.C..... .A..T..T..T.. T..C..T..G
DL54	TGA.CGAC.. .C.C..... .T..... .T..T..A.. T..C..T..A

	2710 2720 2730 2740 2750
Consensus	CTAGAGTCCA TGATMTTYTG GGCWATAGTN AARGCRACYM WRAYYCATT
DL1 IA..T.. ...T....G ..A..G..CC AG.TC.....
DL2 IC..T.. ...T....G ..A..A..CC AG.TC.....
DL13 IC..T.. ...T....G ..A..A..CC AG.TC.....
DL16 IC..T.. ...T....G ..A..A..CC AG.TC.....
ST4 IA..C.. ...A....C ..A..G..CC AA.TC.....
R17 IA..C.. ...A....C ..A..A..CC AA.TC.....
J20 IA..T.. ...T....A ..A..A..CC AG.TC.....
MS2 IA..C.. ...A....C ..A..G..CC AA.TC.....
M12 IA..T.. ...T....C ..A..G..CC AA.TC.....
DL52A..T.. ...A....C ..G..A..TA TG.TC.....
DL54C..T.. ...A....T ..G..A..CA TG.CT.....

	2760 2770 2780 2790 2800
Consensus	TGGTAACSYG GGAACMATWG GCATCTAYGG GGACGATATY ATATGTCCCA
DL1 IGCCC..A.C.. ..T.....
DL2 IGCCC..A.C.. ..T.....
DL13 IGCCC..A.C.. ..T.....
DL16 IGCCC..A.C.. ..T.....
ST4 IGCCC..A.C.. ..T.....
R17 IGCCC..A.C.. ..C.....
J20 IGCCC..A.C.. ..T.....
MS2 IGCCC..A.C.. ..T.....
M12 IGCCC..A.C.. ..T.....
DL52CTTA..T.T.. ..C.....
DL54CTTA..T.T.. ..C.....

	28102820283028402850
Consensus	SWGAGATTGC ACCYCGTGTG CTRGAGGCDC THRSCTWCTA CGGTTTTYAAA
DL1 I	GT.....C.....A.....A..TGC..A... ..T...
DL2 I	GT.....C.....A.....A..TGC..A... ..C...
DL13 I	GT.....C.....A.....A..TGC..A... ..C...
DL16 I	GT.....C.....A.....A..TGC..A... ..C...
ST4 I	GT.....C.....G.....A..TGC..A... ..C...
R17 I	GT.....C.....A.....A..TGC..A... ..T...
J20 I	GT.....C.....A.....G..AGC..A... ..C...
MS2 I	GT.....C.....A.....A..TGC..A... ..T...
M12 I	GT.....C.....A.....A..TGC..A... ..T...
DL52	CA.....T.....A.....T..CAG..T... ..T...
DL54	CA.....T.....A.....T..CAG..T... ..T...

	28602870288028902900
Consensus	CCGAATCWBY SKAARACGTT CRTSWCVGGK CKCTTTTCGCG AGWSCTGYRG
DL1 ITTC GT..A.....G.GT.G..G.T..... ..AG...CG.
DL2 ITTC GT..A.....G.GT.A..G.T..... ..AG...TG.
DL13 ITTC GT..A.....G.GT.A..G.T..... ..AG...TG.
DL16 ITTC GT..A.....G.GT.A..G.T..... ..AG...TG.
ST4 ITCC GT..A.....G.GT.C..G.T..... ..AG...CG.
R17 ITTC GT..A.....G.GT.C..G.T..... ..AG...CA.
J20 ITTC GT..A.....G.GT.A..G.T..... ..AG...TG.
MS2 ITTC GT..A.....G.GT.C..G.T..... ..AG...CG.
M12 ITTC GT..A.....G.GT.C..G.T..... ..AG...CG.
DL52AGT CG..G.....A.CA.G..T.G..... ..TC...TG.
DL54AGT CG..G.....A.CA.G..T.G..... ..TC...TG.

	29102920293029402950
Consensus	YGCRCAYTWY TWCSGYGGTG YYGATKKCAA ACCGWTYTAY ATCARGAAAC
DL1 I	C..G..C.TT .A.C.T.... TC...GT... ..T.T..C ..A.....
DL2 I	C..G..C.TT .A.C.T.... TC...GT... ..T.C..C ..A.....
DL13 I	C..G..C.TT .A.C.T.... TC...GT... ..T.C..C ..A.....
DL16 I	C..G..C.TT .A.C.T.... TC...GT... ..T.C..C ..A.....
ST4 I	C..G..C.TT .A.C.T.... TC...GT... ..T.T..C ..A.....
R17 I	C..G..C.TT .A.C.T.... TC...GT... ..T.T..C ..A.....
J20 I	C..G..C.TT .A.C.T.... TC...GT... ..T.T..C ..G.....
MS2 I	C..G..C.TT .A.C.T.... TC...GT... ..T.T..C ..A.....
M12 I	C..G..C.TT .A.C.T.... TC...GT... ..T.C..C ..A.....
DL52	T..A..T.AC .T.G.C.... CT...TG... ..A.T..T ..A.....
DL54	T..A..T.AC .T.G.C.... CT...TG... ..A.T..T ..A.....

	29602970298029903000
Consensus	CHGTYRACAA YCTCTTYKCC STBWKKCTGW TMHTNAAYMG RCTDMGSGGN
DL1 I	.A..TG.... C.....CT.. C.TATG...A .AA.G..TC. G..TA.G..G
DL2 I	.A..TG.... T.....CT.. C.TATG...A .CA.G..TC. G..TA.G..G
DL13 I	.A..TG.... T.....CT.. C.TATG...A .CA.G..TC. G..TA.G..G
DL16 I	.A..TG.... T.....CT.. C.TATG...A .CA.G..TC. G..TA.G..G
ST4 I	.T..TG.... T.....CG.. C.TATG...A .AT.G..TC. G..AC.G..T

R17 I	.T..TG....	C.....TG..	C.GATG...A	.AT.A..TC.	G..AC.G..T
J20 I	.A..CG....	T.....CT..	C.TATG...A	.CA.G..TC.	G..TA.G..A
MS2 I	.T..TG....	T.....CG..	C.GATG...A	.AT.A..TC.	G..AC.G..T
M12 I	.A..TG....	C.....TT..	C.TATG...A	.AC.G..TC.	G..AC.G..T
DL52	.C..TA....	C.....CG..	G.CTGT...T	.AC.C..CA.	G..AC.C..G
DL54	.C..TA....	C.....CG..	G.CTGT...T	.AC.T..CA.	A..GC.C..C
<div style="text-align: center;"> </div> <div style="text-align: center;"> 3010 3020 3030 3040 3050 </div>					
Consensus	TGGGGNGTTG	TSRRMGGWRT	GTCAGATCCD	CGCCTHTWYR	AGRYKTGGRW
DL1 IC....	.CGGA..TA.GT.ATA	..GTT...GT
DL2 IC....	.CGGA..TA.GT.ATA	..GTT...GT
DL13 IC....	.CGGA..TA.GT.ATA	..GTT...GT
DL16 IC....	.CGGA..TA.GT.ATA	..GTT...GT
ST4 IA....	.CGGA..TA.AT.ACA	..GTG...GT
R17 IG....	.CGGA..TA.AT.ACA	..GTG...GT
J20 IT....	.CGGA..TA.TA.ATA	..GTT...GT
MS2 IA....	.CGGA..TA.AC.ATA	..GTG...GT
M12 IG....	.CGGA..TA.GT.ACA	..GTT...GT
DL52T....	.GAAC..AG.TC.TCG	..ACT...AA
DL54T....	.GAAC..TG.TC.TCG	..ACT...AA
<div style="text-align: center;"> </div> <div style="text-align: center;"> 3060 3070 3080 3090 3100 </div>					
Consensus	AYGRCTVTCC	KMMCDKGTDC	CHTCGATRYT	YTTCGGTSSS	WCDRACCTYG
DL1 I	.C.A..G...	TCC.TG..T.	.A.....GT.	C.....GGG	A.GG....T.
DL2 I	.C.A..A...	TCC.TG..T.	.A.....GT.	C.....GGG	A.GG....T.
DL13 I	.C.A..A...	TCC.TG..T.	.A.....GT.	C.....GGG	A.GG....T.
DL16 I	.C.A..A...	TCC.TG..T.	.A.....GT.	C.....GGG	A.GG....T.
ST4 I	.C.A..C...	TCC.AG..G.	.T.....GT.	T.....GGG	A.GG....C.
R17 I	.C.A..C...	TCC.AG..A.	.T.....GT.	C.....GGG	A.GG....C.
J20 I	.C.A..G...	TCC.TG..T.	.A.....GT.	C.....GGG	A.GG....T.
MS2 I	.C.G..C...	TCC.AG..G.	.T.....GT.	C.....GGG	A.GG....C.
M12 I	.C.A..G...	TCC.TG..T.	.A.....GT.	C.....CCC	A.GG....C.
DL52	.T.G..A...	GAA.GT..T.	.T.....AC.	T.....GGG	T.TA....T.
DL54	.T.G..A...	GAA.GT..T.	.C.....AC.	T.....GGG	T.AA....T.

	31103120313031403150
Consensus	MTGCHGACTA YTACGTRGTY AGYCSBCMNR MNGCYADRWK KCRKTDYGAB
DL1 I	C...C..... T.....A..C ..C.CC.CGA CC...--AGTC T.AG.TT-.T
DL2 I	C...C..... C.....A..T ..C.CT.CGA AT...--TGTC T.GG.TT-.T
DL13 I	C...C..... C.....A..T ..C.CT.CGA AT...--TGTC T.GG.TT-.T
DL16 I	C...C..... C.....A..T ..C.CT.CGA AT...--TGTC T.GG.TT-.T
ST4 I	C...C..... C.....A..C ..C.CG.CCA CG...--AGTC T.GG.AT-.T
R17 I	C...C..... C.....A..C ..C.CG.CTA CG...--GGTC T.GG.AT-.T
J20 I	C...T..... C.....A..C ..C.CC.CGA CA...--GGTC T.AG.TT-.T
MS2 I	C...C..... C.....A..C ..C.CG.CTA CG...--AGTC T.GG.AT-.C
M12 I	C...T..... C.....A..C ..C.CT.CAA CT...--TGTC T.AG.AT-.T
DL52	A...A..... C.....G..T ..T.GT.AGG AA..T.AAAA G.GT.AC..G
DL54	A...A..... T.....G..C ..C.GC.AGG AA..C.GGAA G.GT.GC..G

	31603170318031903200
Consensus	RVYMANMCHB YNTAYGGRMG BHYVCTYKCK SAYDYCRYA SCWCKSSTYW
DL1 I	ACTA.AA.TG CC..C..GA. GTTA..CG.G G.TACC.GT. C.T.GGG.TT
DL2 I	ACCA.AA.TG CA..T..GA. GTTA..CG.G G.CGCC.GT. C.T.GGG.TT
DL13 I	ACCA.AA.TG CA..T..GA. GTTA..CG.G G.CGCC.GT. C.T.GGG.TT
DL16 I	ACCA.AA.TG CA..T..GA. GTTA..CG.G G.CGCC.GT. C.T.GGG.TT
ST4 I	ACCA.GA.TC CG..T..GC. GCTA..CG.G G.TACC.GT. C.T.GGG.TT
R17 I	ACCA.GA.TC CG..T..AC. GCTG..CG.G G.TACC.GT. C.T.GGG.TT
J20 I	ACTA.GA.CG CA..T..GA. GTTG..CG.G G.CGCC.GT. C.T.GGG.TT
MS2 I	ACCA.GA.TC CG..C..GC. GCTG..CG.G G.TACC.GT. C.T.GGG.TT
M12 I	ACTA.GA.CG CG..T..GA. GCTG..CG.G G.TACC.GT. C.T.GGG.TT
DL52	AGTC.CC.AT CT..T..GC. CACC..CT.T C.CTTT.AT. G.A.TCC.CA
DL54	GATC.TC.AG TA..T..AC. TACC..TG.T C.TTTC.AC. G.T.TCC.CA

	32103220323032403250
Consensus	YCGKYTKKMY MGHRYRCCCG YNHVWRARCG HRARYDYTCT YMRCGHRAAG
DL1 I	C..TC.TGCT C.TAT---- CAAAAG.G.. AA.GCGC.-. TAG..AG...
DL2 I	C..TC.TGCT C.TAT---- CGAAAG.G.. AA.GCAC.-. TAG..AG...
DL13 I	C..TC.TGCT C.TAT---- CGAAAG.G.. AA.GCAC.-. TAG..AG...
DL16 I	C..TC.TGCT C.TAT---- CGAAAG.G.. AA.GCAC.-. TAG..AG...
ST4 I	C..TC.TGCT C.TAT---- CTCGAG.A.. CA.GTTC.-. CAG..AA...
R17 I	C..TC.TGCT C.TAT---- CTCGAG.A.. CA.GTTC.-. CAG..AA...
J20 I	C..TC.TGCT C.TAT---- CGAAAG.G.. TA.GCAC.-. TAG..AG...
MS2 I	C..TC.TGCT C.TAT---- CTCGAG.A.. CA.GTTC.-. CAG..AA...
M12 I	C..TC.TGCT A.AAT---- CAAAAG.G.. TA.GCGC.-. TAG..AG...
DL52	T..GC.TTAC C.CGTG.... TATCTA.A.. TG.ATTT... CCA..TG...
DL54	T..GT.GTAC C.CGTA.... CCTCTA.A.. CG.ATTT... TCG..CG...

	32603270328032903300
Consensus	MRYGRCASYG GYCGCYWCAT MRCDTGGTWC CATAMTGGHG GTSARRTYAY
DL1 I	CAT.A..GT. .T...TA... AG.A....T.C...A. ..G.GA.C.C
DL2 I	CAT.A..GT. .T...TA... AG.A....T.C...A. ..G.AA.C.C
DL13 I	CAT.G..GT. .T...TA... AG.A....T.C...A. ..G.AA.C.C
DL16 I	CAT.A..GT. .T...TA... AG.A....T.C...A. ..G.AA.C.C
ST4 I	CAT.A..GT. .C...TA... AG.G....T.C...A. ..G.AG.C.C

R17 I	CAT.A..GT. .T...TA... AG.G....T.C...A. ..G.AA.C.C
J20 I	CAT.A..GT. .T...TA... AG.A....T.C...A. ..G.AA.C.C
MS2 I	CAC.A..GT. .T...TA... AG.G....T.C...A. ..G.AA.C.C
M12 I	CAT.A..GT. .T...TA... AG.G....T.C...A. ..G.AA.C.C
DL52	AAT---CC. .T...CT... CA.T....A.A...T. ..C.AG.T.T
DL54	AGT---CC. .T...CT... CA.T....A.A...C. ..C.AA.T.T

	3310 3320 3330 3340 3350
Consensus	YGAYASYAYK AMGWCCSCHR GSGTKCGYVT HVTRCGMACD TCGGARTGGC
DL1 I	C..T.GT.TG .A.T..G.TG .C..G..CG. AA.G..C..TG....
DL2 I	C..T.GT.TG .A.T..G.TG .C..G..TG. AA.A..C..TG....
DL13 I	C..T.GT.TG .A.T..G.TG .C..G..TG. AA.A..C..TG....
DL16 I	C..T.GT.TG .A.T..G.TG .C..G..TG. AA.A..C..TG....
ST4 I	C..C.GT.TG .A.T..G.CG .C..G..TA. TA.G..C..TG....
R17 I	C..C.GT.TG .A.T..G.CG .C..G..CA. CA.G..C..TG....
J20 I	C..T.GT.TG .A.T..G.CG .C..G..TG. TA.G..C..TA....
MS2 I	C..C.GC.TG .A.T..G.CG .C..G..CG. TA.A..C..TG....
M12 I	C..T.GT.TG .A.T..G.TG .C..G..CG. AC.G..C..TG.--
DL52	T..C.CC.CT .C.A..C.AA .G..T..CC. TG.G..A..GA....
DL54	T..C.CC.CT .C.A..C.AA .G..T..CC. TG.G..A..AA....

	3360 3370 3380 3390 3400
Consensus	TRRCRSYRGT KCCMHYMTTC CCKCAGGARK RTGRCRMMHG CGAGCTCTCC
DL1 I	.GA.GCCG.. T..CACA... ..T.....GT G..G-GCCA.
DL2 I	.GA.GCCG.. T..CATA... ..T.....AT G..G-GCCA.
DL13 I	.GA.GCCG.. T..CATA... ..T.....AT G..G-GCCA.
DL16 I	.GA.GCCG.. T..CATA... ..T.....AT G..G-GCCA.
ST4 I	.AA.GCCG.. T..CACA... ..T.....GT G..G-GCCA.
R17 I	.AA.GCCG.. T..CACA... ..T.....GT G..G-GCCA.
J20 I	.AA.GCCG.. T..CACA... ..G.....GT G..G-GCCA.
MS2 I	.AA.GCCG.. T..CACA... ..T.....GT G..G-GCCA.
M12 I	-----
DL52	.AA.AGTG.. G..ACTC... ..T.....AG A..G.AACT.
DL54	.AG.GGTA.. G..ATCC... ..T.....AG A..A.ACAC.

	STOP 4 Grp I	STOP 4 JS	
		
	3410 3420 3430 3440 3450		
Consensus	TMGKYAGCWS RSCKAGGGAC CCCCCTAAWC GGGGTGGGTG TGCYCGMRAR		
DL1 I	.C.GT...TG AC.G..... ..A..... ..T..AA.G		
DL2 I	.C.GT...TG AC.G..... ..A..... ..T..AA.G		
DL13 I	.C.GT...TG AC.G..... ..A..... ..T..AA.G		
DL16 I	.C.GT...TG AC.G..... ..A..... ..T..AA.G		
ST4 I	.C.GT...TG AC.G..... ..A..... ..T..AA.G		
R17 I	.C.GT...TG AC.G..... ..A..... ..T..AA.G		
J20 I	.C.GT...TG AC.G..... ..A..... ..T..AA.G		
MS2 I	.C.GT...TG AC.G..... ..A..... ..T..AA.G		
M12 I	-----		
DL52	.A.TC...AC GG.T..... ..T..... ---C..CG.A		
DL54	.A.TC...AC GG.T..... ..T..... ---C..CG.A		

	
	34603470348034903500	
Consensus	AGCRSSKRTY	CRYSDWARCR RYCCGGMTSS AYMGRARKRW BSKCSKRMTT
DL1 I	...ACGGG.C	.GCGAA.G.G GT--..C.CC .CC.A.AGGT GGG.GGGC..
DL2 I	...ACGGG.C	.GCGAA.G.G GT--..C.CC .CC.A.AGGT GGG.GGGC..
DL13 I	...ACGGG.C	.GCGAA.G.G GT--..C.CC .CC.A.AGGT GGG.GGGC..
DL16 I	...ACGGG.C	.GCGAA.G.G GT--..C.CC .CC.A.AGGT GGG.GGGC..
ST4 I	...ACGGG.C	.GCGAA.G.G GT--..C.CC .CC.A.AGGT GGG.GGGC..
R17 I	...ACGGG.C	.GCGAA.G.G GT--..C.CC .CC.A.AGGT GGG.GGGC..
J20 I	...ACGGG.C	.GCGTA.G.G GT--..C.CC .CC.A.AGGT GGG.GGGC..
MS2 I	...ACGGG.-	-GCGAA.G.G GT....C.CC .CC.A.AGGT GGG.GGGC..
M12 I	-----	-----
DL52	...GGCTA.T	.ATCGT.A.A AC---.A.GG .TA.G.GTAA TCT.CTAA..
DL54	...GGCTA.T	.ATCGT.A.A AC---.A.GG .TA.G.GTAA CCT.CTAA..

(See App C4) This 3' alignment of JS is not accurate.

	
	35103520353035403550	
Consensus	YGGYSMMKRR	MYYTSCYYBK VWAGWSASSR CCCRGGATTCT TCCCATTGTTG
DL1 I	C..CCCAGGG	ACC.C.CCCT AA..AG.GGA ...G.....
DL2 I	C..CCCAGGG	ACC.C.CCCT AA..AG.GGA ...G.....
DL13 I	C..CCCAGGG	ACC.C.CCCT AA..AG.GGA ...G.....
DL16 I	C..CCCAGGG	ACC.C.CCCT AA..AG.GGA ...G.....
ST4 I	C..CCCAGGG	ACC.C.CCCT GA..AG.GGG ...G.....
R17 I	C..CCCAGGG	ACC.C.CCCT GA..AG.GGA ...G.....
J20 I	C..CCCAGGG	ACC.C.CCCT AA..AG.GGA ...G.....
MS2 I	C..CCCAGGG	ACC.C.CCCT AA..AG.GGA ...G.....
M12 I	-----	-----
DL52	T..TGACTAA	CTT.G.TTGG CT..TC.CCA ...A-----
DL54	C..TGACTAA	CTT.G.TTGG CT..TC.CCA ...A-----

	
	356035703580	
Consensus	GTAAGTAGCT	GCTTGGCTAG TKACCACCCA
DL1 IT.....
DL2 IT.....
DL13 IT.....
DL16 IT.....
ST4 IT.....
R17 IG.....
J20 IT.....
MS2 IT.....
M12 I	-----	-----
DL52	-----	-----
DL54	-----	-----

Appendix C2 JS Amino Acids

Amino acid sequences of *Leviviridae* Group JS. Note the YGDD motif in all *Leviviridae* replicase proteins.

Group JS isolates DL52 and DL54. The amino acid composition is highly conserved among these two strains.

Alignment: Align JS strains maturation protein.

	10 20 30 40 50
Consensus	MRAFSVLDQE SETFVPSVRV YADGQVEDNS FSLKYRSNWT PGRFNSTGSR
DL52 matur	MRAFSVLDQE SETFVPSVRV YADGQVEDNS FSLKYRSNWT PGRFNSTGSR
DL54 matur	MRAFSVLDQE SETFVPSVRV YADGQVEDNS FSLKYRSNWT PGRFNSTGSR

	60 70 80 90 100
Consensus	TEQWHYPSPY SRGALSVTSV DQGAYKRSGS SWGRPYEEKA GFGFSLDARS
DL52 matur	TEQWHYPSPY SRGALSVTSV DQGAYKRSGS SWGRPYEEKA GFGFSLDARS
DL54 matur	TEQWHYPSPY SRGALSVTSV DQGAYKRSGS SWGRPYEEKA GFGFSLDARS

	110 120 130 140 150
Consensus	CYSLFPVSQN LTYIEVPQNV ANRASTEVLQ KVTQGNFNLG VALAEARSTA
DL52 matur	CYSLFPVSQN LTYIEVPQNV ANRASTEVLQ KVTQGNFNLG VALAEARSTA
DL54 matur	CYSLFPVSQN LTYIEVPQNV ANRASTEVLQ KVTQGNFNLG VALAEARSTA

	160 170 180 190 200
Consensus	SQLATQTIAL VKAYTAARRG NWRQTLRYLA LNEDRKFRSK HVAGRWLELQ
DL52 matur	SQLATQTIAL VKAYTAARRG NWRQTLRYLA LNEDRKFRSK HVAGRWLELQ
DL54 matur	SQLATQTIAL VKAYTAARRG NWRQTLRYLA LNEDRKFRSK HVAGRWLELQ

	210 220 230 240 250
Consensus	FGWLPLMSDI QGAYEMLTKV HLQEFLPMRA VRQVGNIKL DGRLSYPAAN
DL52 matur	FGWLPLMSDI QGAYEMLTKV HLQEFLPMRA VRQVGNIKL DGRLSYPAAN
DL54 matur	FGWLPLMSDI QGAYEMLTKV HLQEFLPMRA VRQVGNIKL DGRLSYPAAN

	260 270 280 290 300
Consensus	YQTTCNISRR IVIWFIYINDA RLAWLSSLGI LNPLGIVWEK VPFSFVVDWL
DL52 matur	YQTTCNISRR IVIWFIYINDA RLAWLSSLGI LNPLGIVWEK VPFSFVVDWL
DL54 matur	YQTTCNISRR IVIWFIYINDA RLAWLSSLGI LNPLGIVWEK VPFSFVVDWL

	310 320 330 340 350
Consensus	LPVGNMLEGL TAPVGCSYMS GTVTDVITGE SIISVDAPYG WTVDRQGTAK
DL52 matur	LPVGNMLEGL TAPVGCSYMS GTVTDVITGE SIISVDAPYG WTVDRQGTAK
DL54 matur	LPVGNMLEGL TAPVGCSYMS GTVTDVITGE SIISVDAPYG WTVDRQGTAK

	360 370 380 390
Consensus	AQVSAMHRGV QSVWPTTGVY VKSPFSMVHT LDALALIRQR LLR
DL52 matur	AQVSAMHRGV QSVWPTTGVY VKSPFSMVHT LDALALIRQR LLR
DL54 matur	AQVSAMHRGV QSVWPTTGVY VKSPFSMVHT LDALALIRQR LLR

Alignment: Align JS strains capsid.

	10 20 30 40 50
Consensus	MASNFTQFVL VDNGGTGDVT VAPSNFANGV AEWISSNSRS QAYKVTCSVR
DL52 capsid	MASNFTQFVL VDNGGTGDVT VAPSNFANGV AEWISSNSRS QAYKVTCSVR
DL54 coat	MASNFTQFVL VDNGGTGDVT VAPSNFANGV AEWISSNSRS QAYKVTCSVR

	60 70 80 90 100
Consensus	QSSAQNRKYT IKVEVPKVAT QTVGGVQLPV AAWRSYLNME LTIPIFATND
DL52 capsid	QSSAQNRKYT IKVEVPKVAT QTVGGVQLPV AAWRSYLNME LTIPIFATND
DL54 coat	QSSAQNRKYT IKVEVPKVAT QTVGGVQLPV AAWRSYLNME LTIPIFATND

	110 120 130
Consensus	DCALIVKAMQ GLLKDGNIPI SAIAANSIY
DL52 capsid	DCALIVKAMQ GLLKDGNIPI SAIAANSIY
DL54 coat	DCALIVKAMQ GLLKDGNIPI SAIAANSIY

Alignment: Align JS strains lysis protein.

	10 20 30 40 50
Consensus	METRSPQQSQ QTPESTNRFR PFQHEDYPCR RQQRSSTLYV LIFLAIFLSK
DL52 lysis	METRSPQQSQ QTPESTNRFR PFQHEDYPCR RQQRSSTLYV LIFLAIFLSK
DL54 lysis	METRSPQQSQ QTPESTNRFR PFQHEDYPCR RQQRSSTLYV LIFLAIFLSK

	60 70
Consensus	FTNQLLLSLL EAVIRTVETL QQLLT
DL52 lysis	FTNQLLLSLL EAVIRTVETL QQLLT
DL54 lysis	FTNQLLLSLL EAVIRTVETL QQLLT

Alignment: Align JS strains replicase protein.

	10	20	30	40	50
Consensus	MSKTTKKFNS	LCIDLPRDLS	LEIYQSIASV	ATGSGDPHSR	DFTAIAYLRD	
DL52 repli	MSKTTKKFNS	LCIDLPRDLS	LEIYQSIASV	ATGSGDPHSR	DFTAIAYLRD	
DL54 repli	MSKTTKKFNS	LCIDLPRDLS	LEIYQSIASV	ATGSGDPHSR	DFTAIAYLRD	
	60	70	80	90	100
Consensus	ELLTKHPSLG	NGNDEATRR	LAIAKLREAN	ERCGQINREG	FLHDKSLSWD	
DL52 repli	ELLTKHPSLG	NGNDEATRR	LAIAKLREAN	ERCGQINREG	FLHDKSLSWD	
DL54 repli	ELLTKHPSLG	NGNDEATRR	LAIAKLREAN	ERCGQINREG	FLHDKSLSWD	
	110	120	130	140	150
Consensus	PDVLQTSIRS	LIGNLLSGYR	SSLFGQCTFS	NGASMGHKLQ	DAAPYKKFAE	
DL52 repli	PDVLQTSIRS	LIGNLLSGYR	SSLFGQCTFS	NGASMGHKLQ	DAAPYKKFAE	
DL54 repli	PDVLQTSIRS	LIGNLLSGYR	SSLFGQCTFS	NGASMGHKLQ	DAAPYKKFAE	
	160	170	180	190	200
Consensus	QATVTPRALR	AALLVRDQC	PWIRHAV Y	ESYEFRLVVG	NGVFTVPKNN	
DL52 repli	QATVTPRALR	AALLVRDQCV	PWIRHAVRYN	ESYEFRLVVG	NGVFTVPKNN	
DL54 repli	QATVTPRALR	AALLVRDQCA	PWIRHAVHYS	ESYEFRLVVG	NGVFTVPKNN	
	210	220	230	240	250
Consensus	KIDRAACEKP	DANMYLQKGV	G FIRRRRLRS	IGIDLNDQTI	NQRLAKLGSI	
DL52 repli	KIDRAACEKP	DANMYLQKGV	GDFIRRRRLRS	IGIDLNDQTI	NQRLAKLGSI	
DL54 repli	KIDRAACEKP	DANMYLQKGV	GGFIRRRRLRS	IGIDLNDQTI	NQRLAKLGSI	
	260	270	280	290	300
Consensus	DGSLATIDLS	SASDSISDRL	VWEFLP QMY	AYLSKIRS R	GIVDGR DW	
DL52 repli	DGSLATIDLS	SASDSISDRL	VWEFLPSQMY	AYLSKIRSSR	GIVDGRVVDW	
DL54 repli	DGSLATIDLS	SASDSISDRL	VWEFLPPQMY	AYLSKIRSPR	GIVDGRMIDW	
	310	320	330	340	350
Consensus	HLFSTMGNGF	TFELES MIFW	AIVKATM HF	GNLGTIGIYG	DDIICPTEIA	
DL52 repli	HLFSTMGNGF	TFELES MIFW	AIVKATMIHF	GNLGTIGIYG	DDIICPTEIA	
DL54 repli	HLFSTMGNGF	TFELES MIFW	AIVKATMTHF	GNLGTIGIYG	DDIICPTEIA	
	360	370	380	390	400
Consensus	PRVLEALSFY	GFKPNQSKTF	ITGRFRESCG	AHYFGGADCK	PIYIKKPVNN	
DL52 repli	PRVLEALSFY	GFKPNQSKTF	ITGRFRESCG	AHYFGGADCK	PIYIKKPVNN	
DL54 repli	PRVLEALSFY	GFKPNQSKTF	ITGRFRESCG	AHYFGGADCK	PIYIKKPVNN	

	410 420 430 440 450
Consensus	LFAVCLLLNR LRGWGVVNGV SDPRLFETWK WLSERVPSIL FGGSNLDADY
DL52 repli	LFAVCLLLNR LRGWGVVNGV SDPRLFETWK WLSERVPSIL FGGSNLDADY
DL54 repli	LFAVCLLLNR LRGWGVVNGV SDPRLFETWK WLSERVPSIL FGGSNLDADY

	460 470 480 490 500
Consensus	YVVSQRQA K R E HP YGR TL HFHS PH RLYRVP SKR EFS REESGR
DL52 repli	YVVSQRQA K R E HP YGR TL HFHS PH RLYRVP SKR EFS REESGR
DL54 repli	YVVSQRQA K R E HP YGR TL HFHS PH RLYRVP SKR EFS REESGR

	510 520 530 540
Consensus	LITWYHNGGQ IDTTTTTPRV RLVRTSEWL VVP FPQED ELS
DL52 repli	LITWYHNGGQ VIDTTTTTPRV RLVRTSEWL VVPLFPQEDG NCELS
DL54 repli	LITWYHNGGQ IIDTTTTTPRV RLVRTSEWL VVPSFPQEDD TRELS

Appendix C3 Group I and JS Amino Acids

Amino acid sequences of *Leviviridae* Group I and JS strains (DL52, DL54). Group I strains DL1, DL2, DL13, DL16, J20, ST4, R17, M12, MS2, fr.

A. Align maturation protein.

	10	20	30	40	50		
Consensus	MR F	V	R	YA G	EDNS	L YRSNW	PG	STG
DL52 matur	MRAFSVLDQE	SETFVPSVRV	YADGQVEDNS	FSLKYRSNWT	PGRFNSTGSR			
DL54 MATUR	MRAFSVLDQE	SETFVPSVRV	YADGQVEDNS	FSLKYRSNWT	PGRFNSTGSR			
DL1 matura	MRAFSVLDKE	SETFVPLVRT	YADGEVEDNS	FSLKYRSNWT	PGRFNSTGAR			
DL2 matura	MRAFSVLDQE	SETFVPSVRV	YADGQVEDNS	FSLKYRSNWT	PGRFNSTGSR			
DL13 matur	MRAFSVLDQE	SETFVPSVRV	YADGQVEDNS	FSLKYRSNWT	PGRFNSTGSR			
DL16 matur	MRAFSVLDQE	SETFVPSVRV	YADGQVEDNS	FSLKYRSNWT	PGRFNSTGSR			
J20 matura	MRAFSVLDRE	SETFVPSVRV	YADGEVEDNS	FSLKYRSNWT	PGRFNSTGTR			
ST4 matura	MRAFSTLDRE	NETFVPSVRV	YADGETEDNS	FSLKYRSNWT	PGRFNSTGAK			
R17 matura	MRAFSALDKE	SKTFVPSIRV	YANGETEDNS	FSLKYRSNWT	PGRFNSTGAR			
M12 matura	MRAFSVLDQE	NETFVPSVRV	YADGETEDNS	FSLKYRSNWT	PGRFNSTGAR			
MS2 matura	MRAFSTLDRE	NETFVPSVRV	YADGETEDNS	FSLKYRSNWT	PGRFNSTGAK			
fr maturat	MRKFIPTERM	SKSHVSVRE	YADGELEDNS	LPLIYRSNWS	PGQYTSTGPR			

	60	70	80	90	100
Consensus	T WHYPS Y SRGA	DQG Y R G	SWGR	EE	G G S	DARS
DL52 matur	TEQWHYPSPY	SRGALSVTSV	DQGAYKRSGS	SWGRPYEEKA	GFGFSLDARS	
DL54 MATUR	TEQWHYPSPY	SRGALSVTSV	DQGAYKRSGS	SWGRPYEEKA	GFGFSLDARS	
DL1 matura	TKQWHYPSSY	SRGALSVTSV	DQGAYKRSGS	SWGRPYEEKA	GYGFSLDARS	
DL2 matura	TKQWHYPSPY	SRGALSVTSV	DQGAYKRSGS	SWGRPYEEKA	GFGFSLDARS	
DL13 matur	TEQWHYPSPY	SRGALSVTSV	DQGAYKRSGS	SWGRPYEEKA	GFGFSLDARS	
DL16 matur	TEQWHYPSPY	SRGALSVTSV	DQGAYKRSGS	SWGRPYEEKA	GFGFSLDARS	
J20 matura	TNQWHYPSPY	SRGALSVTSV	DQGSYKRSGS	SWGRPYEEKA	GFGFSLDARS	
ST4 matura	TKQWHYPSPY	SRGALSVTSI	DQGAYKRSGS	SWGRPYEEKA	GFGFSLDARS	
R17 matura	TKQWHYPSPY	SRGALSVTSI	DQGAYKRSGS	SWGRPYEEKA	GFGFSLDARS	
M12 matura	TKQWHYPSPY	SRGALSVTAI	DQGAYKRSGS	SWGRPYEEKT	GFGFSLDARS	
MS2 matura	TKQWHYPSPY	SRGALSVTSI	DQGAYKRSGS	SWGRPYEEKA	GFGFSLDARS	
fr maturat	TKEWHYPSSY	SRGAIGIKAL	DQGYARLGT	SWGREFEERA	GYGMSIDARS	

	110	120	130	140	150
Consensus	CYSLFPVSQN	T I VP NV	ANRA TEVL	KVTQGNFNLG	VALAEARSTA	
DL52 matur	CYSLFPVSQN	LTYIEVPQNV	ANRASTEVLQ	KVTQGNFNLG	VALAEARSTA	
DL54 MATUR	CYSLFPVSQN	LTYIEVPQNV	ANRASTEVLQ	KVTQGNFNLG	VALAEARSTA	
DL1 matura	CYSLFPVSQN	LTYIEVPQNV	ANRASTEVLQ	KVTQGNFNLG	VALAEARSTA	
DL2 matura	CYSLFPVSQN	LTYIEVPQNV	ANRASTEVLQ	KVTQGNFNLG	VALAEARSTA	
DL13 matur	CYSLFPVSQN	LTYIEVPQNV	ANRASTEVLQ	KVTQGNFNLG	VALAEARSTA	
DL16 matur	CYSLFPVSQN	LTYIEVPQNV	ANRASTEVLQ	KVTQGNFNLG	VALAEARSTA	
J20 matura	CYSLFPVSQN	MTYIEVPQNV	ANRASTEVLQ	KVTQGNFNLG	VALAEARSTA	
ST4 matura	CYSLFPVSQN	LTYIEVPQNV	ANRASTEVLQ	KVTQGNFNLG	VALAEARSTA	
R17 matura	CYSLFPVSQN	LTYIEVPQNV	ANRASTEVLQ	KVTQGNFNLG	VALAEARSTA	
M12 matura	CYSLFPVSQN	LTYIEVPQNV	ANRATTEVLQ	KVTQGNFNLG	VALAEARSTA	

MS2 matura	CYSLFPVSQN	LTYIEVPQNV	ANRASTEVLQ	KVTQGNFNLG	VALAEARSTA
fr maturat	CYSLFPVSQN	LTWIDVPTNV	ANRATTEVLG	KVTQGNFNLG	VALAEARSTA

	160	170	180	190	200	
Consensus	SQL TQTIAL	KAYTAARRG	NWRQ RYLA	LNE RKF SK	VA RWLELQ	
DL52 matur	SQLATQTIAL	VKAYTAARRG	NWRQTLRYLA	LNEDRKFRSK	HVAGRWLELQ	
DL54 MATUR	SQLATQTIAL	VKAYTAARRG	NWRQTLRYLA	LNEDRKFRSK	HVAGRWLELQ	
DL1 matura	SQLATQTIAL	VKAYTAARRG	NWRQTLRYLA	LNEDRKFRSK	HVAGRWLELQ	
DL2 matura	SQLATQTIAL	VKAYTAARRG	NWRQTLRYLA	LNEDRKFRSK	HVAGRWLELQ	
DL13 matur	SQLATQTIAL	VKAYTAARRG	NWRQTLRYLA	LNEDRKFRSK	HVAGRWLELQ	
DL16 matur	SQLATQTIAL	VKAYTAARRG	NWRQTLRYLA	LNEDRKFRSK	HVAGRWLELQ	
J20 matura	SQLATQTIAL	VKAYTAARRG	NWRQALRYLA	LNEDRKFRSK	HVAGRWLELQ	
ST4 matura	SQLATQTIAL	VKAYTAARRG	NWRQALRYLA	LNEDRKFRSK	HVAGRWLELQ	
R17 matura	SQLATQTIAL	VKAYTAARRG	NWRQALRYLA	LNEDRKFRSK	HVAGRWLELQ	
M12 matura	SQLATQTIAL	VKAYTAARRG	NWRQPVRILA	LNEDRKFRSK	HVAGRWLELQ	
MS2 matura	SQLATQTIAL	VKAYTAARRG	NWRQALRYLA	LNEDRKFRSK	HVAGRWLELQ	
fr maturat	SQLSTQTIAL	IKAYTAARRG	NWRQALRYLA	LNENRKFN SK	SVASRWLELQ	

	210	220	230	240	250
Consensus	FGW PL SDI	QGAYEMLTKV	HL F PMRA	VRQVG N L	GRL PAA
DL52 matur	FGWLPLMSDI	QGAYEMLTKV	HLQEFLPMRA	VRQVGTNIKL	DGRLSYPAAN
DL54 MATUR	FGWLPLMSDI	QGAYEMLTKV	HLQEFLPMRA	VRQVGTNIKL	DGRLSYPAAN
DL1 matura	FGWLPLMSDI	QGAYEMLTKV	HLQEFLPMRA	VRQVGTNVKL	DGRLSYPAAN
DL2 matura	FGWLPLMSDI	QGAYEMLTKV	HLQEFLPMRA	VRQVGTNIKL	DGRLSYPAAN
DL13 matur	FGWLPLMSDI	QGAYEMLTKV	HLQEFLPMRA	VRQVGTNIKL	DGRLSYPAAN
DL16 matur	FGWLPLMSDI	QGAYEMLTKV	HLQEFLPMRA	VRQVGTNIKL	DGRLSYPAAN
J20 matura	FGWLPLMSDI	QGAYEMLTKV	HLQEFLPMRA	VRQVGTNVKL	DGRLSYPAAN
ST4 matura	FGWLPLMSDI	QGAYEMLTKV	HLQEFLPMRA	VRQVGTNIKL	NGRLSYPAAN
R17 matura	FGWLPLMSDI	QGAYEMLTKV	HLQEFLPMRA	VRQVGTNIKL	DGRLAYPAAN
M12 matura	FGWLPLMSDI	QGAYEMLTKV	HLQEFLPMRA	VRQVGTNIKL	DGRLSYPAAN
MS2 matura	FGWLPLMSDI	QGAYEMLTKV	HLQEFLPMRA	VRQVGTNIKL	DGRLSYPAAN
fr maturat	FGWMPLLSDI	QGAYEMLTKV	HLKAFMPMRA	VRQVGQNVSL	SGRLTSPAAS

	260	270	280	290	300
Consensus	TCNISRR	IVIWFYINDA	RLAWLSSLGI	LNPLGIVWEK	VPFSF VDWL
DL52 matur	YQTTCNISRR	IVIWFYINDA	RLAWLSSLGI	LNPLGIVWEK	VPFSFVVDWL
DL54 MATUR	YQTTCNISRR	IVIWFYINDA	RLAWLSSLGI	LNPLGIVWEK	VPFSFVVDWL
DL1 matura	YQTTCNISRR	IVIWFYINDA	RLAWLSSLGI	LNPLGIVWEK	VPFSFVVDWL
DL2 matura	YQTTCNISRR	IVIWFYINDA	RLAWLSSLGI	LNPLGIVWEK	VPFSFVVDWL
DL13 matur	YQTTCNISRR	IVIWFYINDA	RLAWLSSLGI	LNPLGIVWEK	VPFSFVVDWL
DL16 matur	YQTTCNISRR	IVIWFYINDA	RLAWLSSLGI	LNPLGIVWEK	VPFSFVVDWL
J20 matura	YQTTCNISRR	IVIWFYINDA	RLAWLSSLGI	LNPLGIVWEK	VPFSFVVDWL
ST4 matura	FQTTCNISRR	IVIWFYINDA	RLAWLSSLGI	LNPLGIVWEK	VPFSFVVDWL
R17 matura	YQTTCNISRR	IVIWFYINDA	RLAWLSSLGI	LNPLGIVWEK	VPFSFVVDWL
M12 matura	YQTTCNISRR	IVIWFYINDA	RLAWLSSLGI	LNPLGIVWEK	VPFSFVVDWL
MS2 matura	FQTTCNISRR	IVIWFYINDA	RLAWLSSLGI	LNPLGIVWEK	VPFSFVVDWL
fr maturat	YKSTCNISRR	IVIWFYINDA	RLAWLSSLGI	LNPLGIVWEK	VPFSFLVDWL

	310 320 330 340 350
Consensus	LPVGNMLEGL TAP GCSY S GTVTDVI GE S I D YG W R TAK
DL52 matur	LPVGNMLEGL TAPVGCSYMS GTVTDVITGE SIISVDAPYG WTVDRQGTAK
DL54 MATUR	LPVGNMLEGL TAPVGCSYMS GTVTDVITGE SIISVDAPYG WTVDRQGTAK
DL1 matura	LPVGNMLEGL TAPVGCSYMS GTVTDVITGE SIISVDAPYG WTVDRQGTAK
DL2 matura	LPVGNMLEGL TAPVGCSYMS GTVTDVITGE SIISVDAPYG WTVDRQGTAK
DL13 matur	LPVGNMLEGL TAPVGCSYMS GTVTDVITGE SIISVDAPYG WTVDRQGTAK
DL16 matur	LPVGNMLEGL TAPVGCSYMS GTVTDVITGE SIISVDAPYG WTVDRQGTAK
J20 matura	LPVGNMLEGL TAPVGCSYMS GTVTDVITGE SIISVDAPYG WTVDRQGTAK
ST4 matura	LPVGNMLEGL TAPVGCSYMS GTVTDVITGE SIISVDAPYG WTVDRQGTAK
R17 matura	LPVGNMLEGL TAPVGCSYMS GTVTDVITGE SIISVDAPYG WTVDRQGTAK
M12 matura	LPVGNMLEGL TAPVGCSYMS GTVTDVITGE SIISVDAPYG WTVDRQGTAK
MS2 matura	LPVGNMLEGL TAPVGCSYMS GTVTDVITGE SIISVDAPYG WTVDRQGTAK
fr maturat	LPVGNMLEGL TAPIGCSYQS GTVTDVISGE STITADDIYG WDTVRPATAK

	360 370 380 390
Consensus	SA HRGV QSV PTTG Y VKSPFS VHT LDALAL RQR L
DL52 matur	AQVSAMHRGV QSVWPTTGvy VKSPFSMVHT LDALALIRQR LLR
DL54 MATUR	AQVSAMHRGV QSVWPTTGvy VKSPFSMVHT LDALALIRQR LLR
DL1 matura	AQISAMHRGV QSVWPTTGvy VKSPFSMVHT LDALALIRQR LSR
DL2 matura	AQVSAMHRGV QSVWPTTGvy VKSPFSMVHT LDALALIRQR LLR
DL13 matur	AQVSAMHRGV QSVWPTTGvy VKSPFSMVHT LDALALIRQR LLR
DL16 matur	AQVSAMHRGV QSVWPTTGvy VKSPFSMVHT LDALALIRQR LLR
J20 matura	AQISAMHRGV QSVWPTTGvy VKSPFSIVHT LDALALIRQR LSR
ST4 matura	AQISAMHRGV QSVWPTTGAY VKSPFSMVHT LDALALIRQR LSR
R17 matura	AHVSAMHRGV QSVWPTTGAY VKSPFSMVHT LDALALIRQR LSK
M12 matura	AQVSAMHRGV QSVCPPTGvy VKSPFSMVHT LDALALIRQR LSK
MS2 matura	AQISAMHRGV QSVWPTTGAY VKSPFSMVHT LDALALIRQR LSR
fr maturat	VQISAVHRGV QSVWPTTGvy VKSPFSMVHT LDALALFRQR LWK

B. Alignment of capsid proteins.

	
	10 20 30 40 50	
Consensus	MASNF FVL VDNGGTGDV V PSNFANGV AEWISSNSRS QAYKVTCSVR	
DL52 capsid	MASNFTQFVL VDNGGTGDVT VAPSNFANGV AEWISSNSRS QAYKVTCSVR	
DL54 coat	MASNFTQFVL VDNGGTGDVT VAPSNFANGV AEWISSNSRS QAYKVTCSVR	
DL1 coat p	MASNFTQFVL VDNGGTGDVT VAPSNFANGV AEWISSNSRS QAYKVTCSVR	
DL2 coat	MASNFTQFVL VDNGGTGDVT VAPSNFANGV AEWISSNSRS QAYKVTCSVR	
DL13 coat	MASNFTQFVL VDNGGTGDVT VAPSNFANGV AEWISSNSRS QAYKVTCSVR	
DL16 coat	MASNFTQFVL VDNGGTGDVT VAPSNFANGV AEWISSNSRS QAYKVTCSVR	
J20 coat	MASNFTQFVL VDNGGTGDVT VAPSNFANGV AEWISSNSRS QAYKVTCSVR	
ST4 coat p	MASNFTQFVL VDNGGTGDVT VAPSNFANGV AEWISSNSRS QAYKVTCSVR	
R17 coat p	MASNFTQFVL VDNGGTGDVT VAPSNFANGV AEWISSNSRS QAYKVTCSVR	
M12 coat	MASNFTQFVL VDNGGTGDVT VXPSNFANGV AEWISSNSRS QAYKVTCSVR	
MS2 coat p	MASNFTQFVL VDNGGTGDVT VAPSNFANGV AEWISSNSRS QAYKVTCSVR	
fr coat pr	MASNFEEFVL VDNGGTGDVK VAPSNFANGV AEWISSNSRS QAYKVTCSVR	

	
	60 70 80 90 100	
Consensus	QSSA NRKYT KVEVPKVAT Q GGV LPV AAWRSY NME LTIP FATN	
DL52 capsid	QSSAQNRKYT IKVEVPKVAT QTVGGVQLPV AAWRSYLNME LTIPIFATND	
DL54 coat	QSSAQNRKYT IKVEVPKVAT QTVGGVQLPV AAWRSYLNME LTIPIFATND	
DL1 coat p	QSSAQNRKYT IKVEVPKVAT QTVGGVQLPV AAWRSYLNME LTIPIFATND	
DL2 coat	QSSAQNRKYT IKVEVPKVAT QTVGGVQLPV AAWRSYLNME LTIPIFATND	
DL13 coat	QSSAQNRKYT IKVEVPKVAT QTVGGVQLPV AAWRSYLNME LTIPIFATND	
DL16 coat	QSSAQNRKYT IKVEVPKVAT QTVGGVQLPV AAWRSYLNME LTIPIFATND	
J20 coat	QSSAQNRKYT IKVEVPKVAT QTVGGVQLPV AAWRSYLNME LTIPIFATND	
ST4 coat p	QSSAQNRKYT IKVEVPKVAT QTVGGVELPV AAWRSYLNME LTIPIFATNS	
R17 coat p	QSSAQNRKYT IKVEVPKVAT QTVGGVELPV AAWRSYLNME LTIPIFATNS	
M12 coat	QSSAQNRKYT IKVEVPKVAT QTVGGVELPV AAWRSYLNME LTIPIFATNS	
MS2 coat p	QSSAQNRKYT IKVEVPKVAT QTVGGVELPV AAWRSYLNME LTIPIFATNS	
fr coat pr	QSSANNRKYT VKVEVPKVAT QVQGGVELPV AAWRSYMNME LTIPVFATND	

	
	110 120 130	
Consensus	DC LIVKA Q G K GNPI AIAANSIY	
DL52 capsid	DCALIVKAMQ GLLKDGNIPI SAIAANSIY	
DL54 coat	DCALIVKAMQ GLLKDGNIPI SAIAANSIY	
DL1 coat p	DCALIVKAMQ GLLKDGNIPI SAIAANSIY	
DL2 coat	DCALIVKAMQ GLLKDGNIPI SAIAANSIY	
DL13 coat	DCALIVKAMQ GLLKDGNIPI SAIAANSIY	
DL16 coat	DCALIVKAMQ GLLKDGNIPI SAIAANSIY	
J20 coat	DCALIVKAMQ GLLKDGNIPI SAIAANSIY	
ST4 coat p	DCELIVKAMQ GLLKDGNIPI SAIAANSIY	
R17 coat p	DCELIVKAMQ GLLKDGNIPI SAIAANSIY	
M12 coat	DCALIVKAMQ GLLKDGNIPI SAIAANSIY	
MS2 coat p	DCELIVKAMQ GLLKDGNIPI SAIAANSIY	
fr coat pr	DCALIVKALQ GTFKTGNPIA TAIAANSIY	

C. Alignment of lysis proteins.

	10 20 30 40 50
Consensus	M SQ T S PF HE YPC QQRSSTLYV LI LAIFLSK
DL52 lysis	METRSPQQSQ QTPESTNRFR PFQHEDYPCR RQRSSTLYV LIFLAIFLSK
DL54 lysis	METRSPQQSQ QTPESTNRFR PFQHEDYPCR RQRSSTLYV LIFLAIFLSK
DL2 lysis	METRSPQQSQ QTPESTNRFR PFQHEDYPCR RQRSSTLYV LIFLAIFLSK
DL13 lysis	METRSPQQSQ QTPESTNRFR PFQHEDYPCR RQRSSTLYV LIFLAIFLSK
DL16 lysis	METRSPQQSQ QTPESTNRFR PFQHEDYPCR RQRSSTLYV LIFLAIFLSK
J20 lysis	METQSPQQSQ PTPESINRFR PFQHEDYPCR RQRSSTLYV LIFLAIFLSK
DL1 lysis	METRSPQQSQ QTPESTNRFR PFKHEDYPCR RQRSSTLYV LIFLAIFLSK
ST4 lysis	METRFPQQSQ QTPASTNRRR PFKHEDYPCR RQRSSTLYV LIFLAIFLSK
R17 lysis	METRFPQQSQ QTPASTNRCR PFKHEDYPCR RQRSSTLYV LIFLAIFLSK
M12 lysis	METRFLRQSQ QTPASTNRYR PFKHEDYPCR XQRSSTLYV LIFLAIFLSK
MS2 lysis	METRFPQQSQ QTPASTNRRR PFKHEDYPCR RQRSSTLYV LIFLAIFLSK
fr lysis p	MQ----QPSQ PTRESTKKPV PFQHEEYPCQ NQRSSTLYV LICLAIFLSK

	60 70
Consensus	FTNQLL SLL IR V T QLLT
DL52 lysis	FTNQLLLSLL EAVIRTVETL QLLT
DL54 lysis	FTNQLLLSLL EAVIRTVETL QLLT
DL2 lysis	FTNQLLLSLL EAVIRTVETL QLLT
DL13 lysis	FTNQLLLSLL EAVIRTVETL QLLT
DL16 lysis	FTNQLLLSLL EAVIRTVETL QLLT
J20 lysis	FTNQLLLSLL EAVIRTVETL RQLLT
DL1 lysis	FTNQLLLSLL EAVIRTVETL QLLT
ST4 lysis	FTNQLLLSLL EAVIRTVTTL QLLT
R17 lysis	FTNQLLLSLL EAVIRTVTTL QLLT
M12 lysis	FTNQLLLSLL DAVIRTVTTF QLLT
MS2 lysis	FTNQLLLSLL EAVIRTVTTL QLLT
fr lysis p	FTNQLLASLL DLLIRIVTTL QLLT

D. Alignment of replicase.

	
	10 20 30 40 50	
Consensus	MSK TKKFNS LCIDL DLS LE YQSIASV ATGS PHS DFTAIAYLRD	
DL52 repli	MSKTTKKFNS LCIDLPRDLS LEIYQSIASV ATGSGDPHSR DFTAIAYLRD	
DL54 repli	MSKTTKKFNS LCIDLPRDLS LEIYQSIASV ATGSGDPHSR DFTAIAYLRD	
DL1 replic	MSKTTKKFNS LCIDLPRDLS LEIYQSIASV ATGSGDPHSR DFTAIAYLRD	
DL2 replic	MSKTTKKFNS LCIDLPRDLS LEIYQSIASV ATGSGDPHSR DFTAIAYLRD	
DL13 repli	MSKTTKKFNS LCIDLPRDLS LEIYQSIASV ATGSGDPHSR DFTAIAYLRD	
DL16 repli	MSKTTKKFNS LCIDLPCDLS LEIYQSIASV ATGSGDPHSR DFTAIAYLRD	
ST4 replic	MSKTTKKFNS LCIDLPRDLS LEIYQSIASV ATGSGDPHSD DFTAIAYLRD	
R17 replic	MSKTTKKFNS LCIDLPRDLS LEIYQSIASV ATGSGNPHSD DFTAIAYLRD	
J20 replic	MSKTTKKFNS LCIDLPRDLS LEIYQSIASV ATGSGDPHSR DFTAIAYLRD	
MS2 replic	MSKTTKKFNS LCIDLPRDLS LEIYQSIASV ATGSGDPHSD DFTAIAYLRD	
fr replica	MSKSTKKFNS LCIDLSRDLS LEVYQSIASV ATGSSDPHSD DFTAIAYLRD	

	
	60 70 80 90 100	
Consensus	ELLTKHP LG GNDEATRR LAIAKL EAN GQINR G FLHD SWD	
DL52 repli	ELLTKHPSLG NGNDEATRR LAIAKLREAN ERCGQINREG FLHDKSLSWD	
DL54 repli	ELLTKHPSLG NGNDEATRR LAIAKLREAN ERCGQINREG FLHDKSLSWD	
DL1 replic	ELLTKHPSLG NGNDEATRR LAIAKLREAN ERCGQINREG FLHDKSLSWD	
DL2 replic	ELLTKHPSLG NGNDEATRR LAIAKLREAN ERCGQINREG FLHDKSLSWD	
DL13 repli	ELLTKHPSLG NGNDEATRR LAIAKLREAN ERCGQINREG FLHDKSLSWD	
DL16 repli	ELLTKHPSLG NGNDEATRR LAIAKLREAN ERCGQINREG FLHDKSLSWD	
ST4 replic	ELLTKHPTLG SGNDEATRR LAIAKLREAN DRCGQINREG FLHDKSLSWD	
R17 replic	ELLTKHPTLG SGNDEATRR LAIAKLREAN DRCGQINREG FLHDKSLSWD	
J20 replic	ELLTKHPSLG NGNDEATRR LAIAKLREAN ERCGQINREG FLHDKSLSWD	
MS2 replic	ELLTKHPTLG SGNDEATRR LAIAKLREAN GDRGQINREG FLHDKSLSWD	
fr replica	ELLTKHPNLG DGNDEATRR LAIAKLLEAN DRCGQINRDG FLHDATAASWD	

	
	110 120 130 140 150	
Consensus	PDVLQTSIRS LIGNLLSGY S LF CTFS NGA MGHKLQ DAAPYKKFAE	
DL52 repli	PDVLQTSIRS LIGNLLSGYR SSLFGQCTFS NGASMGHKLQ DAAPYKKFAE	
DL54 repli	PDVLQTSIRS LIGNLLSGYR SSLFGQCTFS NGASMGHKLQ DAAPYKKFAE	
DL1 replic	PDVLQTSIRS LIGNLLSGYR SSLFGQCTFS NGASMGHKLQ DAAPYKKFAE	
DL2 replic	PDVLQTSIRS LIGNLLSGYR SSLFGQCTFS NGASMGHKLQ DAAPYKKFAE	
DL13 repli	PDVLQTSIRS LIGNLLSGYR SSLFGQCTFS NGASMGHKLQ DAAPYKKFAE	
DL16 repli	PDVLQTSIRS LIGNLLSGYR SSLFGQCTFS NGASMGHKLQ DAAPYKKFAE	
ST4 replic	PDVLQTSIRS LIGNLLSGYR SSLFGQCTFS NGASMGHKLQ DAAPYKKFAE	
R17 replic	PDVLQTSIRS LIGNLLSGYQ SSLFGQCTFS NGASMGHKLQ DAAPYKKFAE	
J20 replic	PDVLQTSIRS LIGNLLSGYR SSLFGQCTFS NGASMGHKLQ DAAPYKKFAE	
MS2 replic	PDVLQTSIRS LIGNLLSGYR SSLFGQCTFS NGAPMGHKLQ DAAPYKKFAE	
fr replica	PDVLQTSIRS LIGNLLSGYS SQLFRHCTFS NGASMGHKLQ DAAPYKKFAE	

	160 170 180 190 200
Consensus	QATVTPRAL AA LV DQC PWIRH ESY FRLV G NGVFTVPKNN
DL52 repli	QATVTPRALR AALLVRDQCV PWIRHAVRYN ESYEFRLVVG NGVFTVPKNN
DL54 repli	QATVTPRALR AALLVRDQCA PWIRHAVHYS ESYEFRLVVG NGVFTVPKNN
DL1 replic	QATVTPRALR AALLVRDQCV PWIRHAVRYN ESYEFRLVVG NGVFTVPKNN
DL2 replic	QATVTPRALR AALLVRDQCV PWIRHAVRYN ESYEFRLVVG NGVFTVPKNN
DL13 repli	QATVTPRALR AALLVRDQCV PWIRHAVRYN ESYEFRLVVG NGVFTVPKNN
DL16 repli	QATVTPRALR AALLVRDQCV PWIRHAVRYN ESYEFRLVVG NGVFTVPKNN
ST4 replic	QATVTPRALR AALLVRDQCA PWIRHAVRYN ESYEFRLVVG NGVFTVPKNN
R17 replic	QATVTPRALR AALLVRDQCA PWIRHAVHYN ESYEFRLVVG NGVFTVPKNN
J20 replic	QATVTPRALR AALLVRDQCA PWIRHAVRYN ESYKFRVLVG NGVFTVPKNN
MS2 replic	QATVTPRALR AALLVRDQCA PWIRHAVRYN ESYEFRLVVG NGVFTVPKNN
fr replica	QATVTPRALK AAVLVKDQCS PWIRHSHVFP ESYTFRLVGG NGVFTVPKNN

	210 220 230 240 250
Consensus	KIDRAACEP D NMYLQKGV G FIRRL GIDLNDQ I NQ LA GS
DL52 repli	KIDRAACEP DANMYLQKGV GDFIRRLRS IGIDLNDQTI NQRLAKLGS
DL54 repli	KIDRAACEP DANMYLQKGV GGFIRRLRS IGIDLNDQTI NQRLAKLGS
DL1 replic	KIDRAACEP DMNMYLQKGV GAFIRRLKS VGIDLNDQTI NQRLAQQGS
DL2 replic	KIDRAACEP DMNMYLQKGV GAFIRRLKS VGIDLNDQTI NQRLAQQGS
DL13 repli	KIDRAACEP DMNMYLQKGV GAFIRRLKS VGIDLNDQTI NQRLAQQGS
DL16 repli	KIDRAACEP DMNMYLQKGV GAFIRRLKS VGIDLNDQTI NQRLAQQGS
ST4 replic	KIDRAACEP DMNMYLQKGV GAFIRRLKS VGIDLNDQSI NQLLAQQGSV
R17 replic	KIDRAACEP DMNMYLQKGV GAFIRRLKS VGIDLNDQSI NQRLAQQGS
J20 replic	KIDRAACEP DMNMYLQKGV GAFIRRLRS VGIDLNDQTI NQRLAQQGSV
MS2 replic	KIDRAACEP DMNMYLQKGV GAFIRRLKS VGIDLNDQSI NQRLAQQGSV
fr replica	KIDRAACEP DMNMYLQKGV GGFIRRLKT VGIDLNDQTI NQRLAQQGS

	260 270 280 290 300
Consensus	DGSLATIDLS SASDSISDRL VW FLP Y YL IRS G G W
DL52 repli	DGSLATIDLS SASDSISDRL VWEFLPSQMY AYLSKIRSSR GIVDGRVVDW
DL54 repli	DGSLATIDLS SASDSISDRL VWEFLPPQMY AYLSKIRSPR GIVDGRMIDW
DL1 replic	DGSLATIDLS SASDSISDRL VWSFLPPELY SYLDRIIRSHY GIVDGETIRW
DL2 replic	DGSLATIDLS SASDSISDRL VWSFLPPELY SYLDRIIRSHY GIVDGETIRW
DL13 repli	DGSLATIDLS SASDSISDRL VWSFLPPELY SYLDRIIRSHY GIVDGETIRW
DL16 repli	DGSLATIDLS SASDSISDRL VWSFLPPELY SYLDRIIRSHY GIVDGETIRW
ST4 replic	DGSLATIDLS SASDSISDRL VWSFLPPELY SYLDRIIRSHY GIVDGETIRW
R17 replic	DGSLATIDLS SASDSISDRL VWNFLPPELY SYLDRIIRSHY GIVDGETIRW
J20 replic	DGSLATIDLS SASDSISDRL VWSFLPPELY SYLDRIIRSHY GIIDGETIRW
MS2 replic	DGSLATIDLS SASDSISDRL VWSFLPPELY SYLDRIIRSHY GIVDGETIRW
fr replica	DGSLATIDLS SASDSISDRL VWSFLPPELY SYLDMIRSHY GYVNGKMIRW

	310 320 330 340 350
Consensus	LFSTMGNGF TFELESMIFW AIV AT HF N GTIGIYG DDIICP EIA
DL52 repli	HLFSTMGNGF TFELESMIFW AIVKATMIHF GNLGTIGIYG DDIICPTEIA
DL54 repli	HLFSTMGNGF TFELESMIFW AIVKATMTHF GNLGTIGIYG DDIICPTEIA
DL1 replic	ELFSTMGNGF TFELESMIFW AIVKATQIHF GNAGTIGIYG DDIICPSEIA
DL2 replic	ELFSTMGNGF TFELESMIFW AIVKATQIHF GNAGTIGIYG DDIICPSEIA
DL13 repli	ELFSTMGNGF TFELESMIFW AIVKATQIHF GNAGTIGIYG DDIICPSEIA
DL16 repli	ELFSTMGNGF TFELESMIFW AIVKATQIHF GNAGTIGIYG DDIICPSEIA
ST4 replic	ELFSTMGNGF TFELESMIFW AIVKATQIHF GNAGTIGIYG DDIICPSEIA
R17 replic	ELFSTMGNGF TFELESMIFW AIVKATQIHF GNAGTIGIYG DDIICPSEIA
J20 replic	ELFSTMGNGF TFELESMIFW AIVKATQIHF GNAGTIGIYG DDIICPSEIA
MS2 replic	ELFSTMGNGF TFELESMIFW AIVKATQIHF GNAGTIGIYG DDIICPSEIA
fr replica	ELFSTMGNGF TFELESMIFW AIVRATQIHF RNTGTIGIYG DDIICPTEIA

	360 370 380 390 400
Consensus	PRVLEAL Y GFKPN KTF G FRESC AH G D K P YI KP
DL52 repli	PRVLEALSFY GFKPNQSKTF ITGRFRESCG AHYFGGADCK PIYIKKPVNN
DL54 repli	PRVLEALSFY GFKPNQSKTF ITGRFRESCG AHYFGGADCK PIYIKKPVNN
DL1 replic	PRVLEALAYY GFKPNLRKTF VSGLFRESCG AHFYRGVDVK PFYIKKPVDN
DL2 replic	PRVLEALAYY GFKPNLRKTF VSGLFRESCG AHFYRGVDVK PFYIKKPVDN
DL13 repli	PRVLEALAYY GFKPNLRKTF VSGLFRESCG AHFYRGVDVK PFYIKKPVDN
DL16 repli	PRVLEALAYY GFKPNLRKTF VSGLFRESCG AHFYRGVDVK PFYIKKPVDN
ST4 replic	PRVLEALAYY GFKPNLRKTF VSGLFRESCG AHFYRGVDVK PFYIKKPVDN
R17 replic	PRVLEALAYY GFKPNLRKTF VSGLFRESCS AHFYRGVDVK PFYIKKPVDN
J20 replic	PRVLEALAYY GFKPNLRKTF VSGLFRESCG AHFYRGVDVK PFYIRKPVDN
MS2 replic	PRVLEALAYY GFKPNLRKTF VSGLFRESCG AHFYRGVDVK PFYIKKPVDN
fr replica	PRVLEALSFY GFKPNLRKTF TSGSFRESCG AHYFRGVDVK PFYIKKPITD

	410 420 430 440 450
Consensus	LF L NR RGWGVV G DPRL W LS VP FGG L ADY
DL52 repli	LFAVCLLLNR LRGWGVVNGV SDPRLFETWK WLSERVPSIL FGGSNLDADY
DL54 repli	LFAVCLLLNR LRGWGVVNGV SDPRLFETWK WLSERVPSIL FGGSNLDADY
DL1 replic	LFSMLIMNR LRGWGVVGGM SDPRLYKVWV RLSSLVPSMF FGGLDLAADY
DL2 replic	LFSMLIMNR LRGWGVVGGM SDPRLYKVWV RLSSLVPSMF FGGLDLAADY
DL13 repli	LFSMLIMNR LRGWGVVGGM SDPRLYKVWV RLSSLVPSMF FGGLDLAADY
DL16 repli	LFSMLIMNR LRGWGVVGGM SDPRLYKVWV RLSSLVPSMF FGGLDLAADY
ST4 replic	LFALMLILNR LRGWGVVGGM SDPRLYKVWV RLSSQVPSMF FGGLDLAADY
R17 replic	LFALMLILNR LRGWGVVGGM SDPRLYKVWV RLSSQVPSMF FGGLDLAADY
J20 replic	LFSMLIMNR LRGWGVVGGM SDPRLYKVWV RLSSLVPSMF FGGLDLAADY
MS2 replic	LFALMLILNR LRGWGVVGGM SDPRLYKVWV RLSSQVPSMF FGGLDLAADY
fr replica	LFSMLILNR IRGWGVVNGI ADPRLYEVWE KLSRLVPRYL FGGLDLQADY

	460 470 480 490 500
Consensus	YVVS R L R FS SGR
DL52 repli	YVVSQRQAKK RYESHPSYGR TLSHFHSTPH RLYRVPVSKR EFSPREESGR
DL54 repli	YVVSQRQARK RCEDHPVYGR TLAHFHSSPH RLYRVPASKR EFSSREESGR
DL1 replic	YVVSPPTAVS VYTKTA-YGR LLADTRTSGF RLARIAKERK RFSEKHDSGR
DL2 replic	YVVSPPNAVS VYTKTA-YGR LLADARTSGF RLARIAKERK HFSEKHDSGR
DL13 repli	YVVSPPNAVS VYTKTA-YGR LLADARTSGF RLARIAKERK HFSEKHGSGR

DL16 repli	YVVSPPNAVS	VYTKTA-YGR	LLADARTSGF	RLARTAKERK	HFSEKHDSGR
ST4 replic	YVVSPPTAVS	VYTKTP-YGR	LLADTRTSGF	RLARIARERK	FFSEKHDSGR
R17 replic	YVVSPPTAVS	VYTKTP-YGR	LLADTRTSGF	RLARIARERK	FFSEKHDSGR
J20 replic	YVVSPPTAVS	VYTKTA-YGR	LLADARTSGF	RLARIAKERK	HFSEKHDSGR
MS2 replic	YVVSPPTAVS	VYTKTP-YGR	LLADTRTSGF	RLARIARERK	FFSEKHDSGR
fr replica	YVVSPPILKG	IYSKMN-GRR	EYAEARTTGF	KLARIARWRK	HFSDKHDSGR

	510 520 530 540
Consensus	I W H GG D V R RTSEWL VP FPQE
DL52 repli	LITWYHNGGQ VIDTTTTTPRV RLVRTSEWLT VVPLFPQEDG NCELS-
DL54 repli	LITWYHNGGQ IIDTTTTTPRV RLVRTSEWLA VVPSFPQEDD TRELS-
DL1 replic	YIAWFHTGGE ITDSMKSAGV RVMRTSEWLT PVPTFPQECG PASSPR
DL2 replic	YIAWFHTGGE ITDSMKSAGV RVIRTSEWLT PVPIFPQECG PASSPR
DL13 repli	YIAWFHTGGE ITDSMKSAGV RVIRTSEWLT PVPIFPQECG PASSPR
DL16 repli	YIAWFHTGGE ITDSMKSAGV RVIRTSEWLT PVPIFPQECG PASSPR
ST4 replic	YIAWFHTGGE VTDSMKSAGV RIMRTSEWLT PVPTFPQECG PASSPR
R17 replic	YIAWFHTGGE ITDSMKSAGV RIMRTSEWLT PVPTFPQECG PASSPR
J20 replic	YIAWFHTGGE ITDSMKSAGV RVMRTSEWLT PVPTFPQECG PASSPR
MS2 replic	YIAWFHTGGE ITDSMKSAGV RVIRTSEWLT PVPTFPQECG PASSPR
fr replica	YIAWFHTGGE ITDSMKSAGV RVMRTSEWLQ PVPVFPQECG PASSPQ

Alignment: Group II replicase and JS strains.

	10 20 30 40 50
Consensus	M LC D RD S GS DP S DF AYLRD
DL52 repli	MSKTTKKFNS LCIDLPRDLS LEIYQSIASV ATGSGDPHSR DFTAIAYLRD
DL54 repli	MSKTTKKFNS LCIDLPRDLS LEIYQSIASV ATGSGDPHSR DFTAIAYLRD
DL10 repli	MFRFTEIEKT LCMDRTRDCA VRFHVYLQSL DMGSSDPHSP DFDGLAYLRD
DL20 repli	MFRFTEIEKT LCMDRTRDCA VRFHVYLQSL DLGSSDPHSP DFDGLAYLRD
GA replica	MFRFTEIEKT LCMDRTRDCA VRFHVYLQSL DLGSSDPLSP DFDGLAYLRD
T72 replic	MFRFTEIKKT LCMDRTRDCA VRFHVYLQSL DLGSSDPHSP DFDGLAYLRD
KU1 replic	MFRFTEIEKT LCMDRTRDCA VRFHVYLQSL DLGSSDPHSP DFDGLAYLRD

	60 70 80 90 100
Consensus	E LTKHPSLG N A R LA AKL RC N G
DL52 repli	ELLTKHPSLG NGNDEATRR LAI AKLREAN ERCGQINREG FLHDKSLSWD
DL54 repli	ELLTKHPSLG NGNDEATRR LAI AKLREAN ERCGQINREG FLHDKSLSWD
DL10 repli	ECLTKHPSLG NSNSDARRKE LAYAKLMDS QRCKIQNSNG YDYSHIESV
DL20 repli	ECLTKHPSLG DSNSDARRKE LAYAKLMDS QRCKIQNSNG YDYSHIESG
GA replica	ECLTKHPSLG DSNSDARRKE LAYAKLMDS QRCKIQNSNG YDYSHIESG
T72 replic	ECLTKHPSLG DSNSDALRKE LAYAKLMDS QRCKIQNSNG YDLSHIDSG
KU1 replic	ECLTKHPSLG DSNSDALRKE LAYAKLMDS QRCKIQNSNG YDLSHIDAGV

	110 120 130 140 150
Consensus	LL G S C FS NGAS G KL DAAP KK A
DL52 repli	PDVLQTSIRS LIGNLLSGYR SSLFGQCTFS NGASMGHKLQ DAAPYKKFAE
DL54 repli	PDVLQTSIRS LIGNLLSGYR SSLFGQCTFS NGASMGHKLQ DAAPYKKFAE
DL10 repli	LSGILKTAQA LVANLLTGFE SHFLNDCSFS NGASQGFKLR DAAPFKKIAG
DL20 repli	LSGILKTAQA LVANLLTGFE SHFLNDCSFS NGASQGFKLR DAAPFKKIAG
GA replica	LSGILKTAQA LVANLLTGFE SHFLNDCSFS NGASQGFKLR DAAPFKKIAG
T72 replic	LNGILLTAKA SIAKLLMGFE SHFLNDCSFS NGASQGFKLQ DAAPFKKIAG
KU1 replic	LNGILLTAKA LIAKLLIGFE SHFLNDCSFS NGASQGFKLR DAAPFKKIAG

	160 170 180 190 200
Consensus	QATVT A A C PW E FR V GNG F VPK
DL52 repli	QATVTPRALR AALLVRDQCV PWIRHAVRYN --ESYEFRLV VGNGVFTVPK
DL54 repli	QATVTPRALR AALLVRDQCA PWIRHAVHYS --ESYEFRLV VGNGVFTVPK
DL10 repli	QATVTAPAYN IAVA AVKTCA PWYAYMQETY GDETRWFRRV YGNGLFSVPK
DL20 repli	QATVTAPAYD IAVA AVKTCA PWYAYMQETY GDETKWFRRV YGNGLFSVPK
GA replica	QATVTAPAYD IAVA AVKTCA PWYAYMQETY GDETKWFRRV YGNGLFSVPK
T72 replic	QATVTAPAYD LAVHAVKTCG PWLRYMQETY GDETRWFRRV YGNGLFSVPK
KU1 replic	QATVTAPAYD LAVLAVKTCG PWLRYMQETY GDETRWFRRV YGNGLFSVPK

	210 220 230 240 250
Consensus	NNKIDRAACK EPD NMYLQK G G FIR RL RS IDLNDQ T NQ LA LG
DL52 repli	NNKIDRAACK EPDANMYLQK GVGDFIRRL RSIGIDLNDQ TINQRLAKLG
DL54 repli	NNKIDRAACK EPDANMYLQK GVGGFIRRL RSIGIDLNDQ TINQRLAKLG
DL10 repli	NNKIDRAACK EPDMNMYLQK GAGSFIRKRL RSVGIDLNDQ TCNQELARLG
DL20 repli	NNKIDRAACK EPDMNMYLQK GAGSFIRKRL RSVGIDLNDQ TRNQELARLG
GA replica	NNKIDRAACK EPDMNMYLQK GAGSFIRKRL RSVGIDLNDQ TRNQELARLG
T72 replic	NNKIDRAACK EPDMNMYLQK GAGSFIRRL RSVNIDLNDQ TRNQELARLG
KU1 replic	NNKIDRAACK EPDMNMYLQK GAGSFIRRL RSVNIDLNDQ TRNQELARLG

	260 270 280 290 300
Consensus	SIDGSLATID LSSASDS SD RLVW LP Y YL IR DG
DL52 repli	SIDGSLATID LSSASDSISD RLVWEFLPSQ MYAYLSKIRS SRGIVDGRVV
DL54 repli	SIDGSLATID LSSASDSISD RLVWEFLPPQ MYAYLSKIRS PRGIVDGRMI
DL10 repli	SIDGSLATID LSSASDSVSD RLVWDLLPPH VYSYLARIRS SFTMIDGRLH
DL20 repli	SIDGSLATID LSSASDSISD RLVWDLLPPH VYSYLARIRS SFTMIDGRLH
GA replica	SIDGSLATID LSSASDSISD RLVWDLLPPH VYSYLARIRT SFTMIDGRLH
T72 replic	SIDGSLATID LSSASDSVSD RLVWDLLPPH VYSYLHRIRS SFTMIDGQLH
KU1 replic	SIDGSLATID LSSASDSVSD RLVWDLLPPH VYSYLHRIRS SFTMIDGRLH

	310 320 330 340 350
Consensus	W LFSTMG N GFTFELES MI FWA M G G G YGDDII P
DL52 repli	DWHLFSTMG N GFTFELES MI FWAIVKATMI HFGNLGTIGI YGDDIICPTE
DL54 repli	DWHLFSTMG N GFTFELES MI FWAIVKATMT HFGNLGTIGI YGDDIICPTE
DL10 repli	KWGLFSTMG N GFTFELES MI FWALSKSVML SMGVTGSLGI YGDDIIVPVE
DL20 repli	KWGLFSTMG N GFTFELES MI FWALSKSVML SMGVTGSLGV YGDDIIVPVE
GA replica	KWGLFSTMG N GFTFELES MI FWALSKSIML SMGVTGSLGI YGDDIIVPVE
T72 replic	KWNLFSTMG N GFTFELES MI FWALSKSVMS YLGVTGLLGI YGDDIIVPTK
KU1 replic	KWNLFSTMG N GFTFELES MI FWALSNTVMS YLGVTGLLGI YGDDIIVPTK

	360 370 380 390 400
Consensus	P L LS F PN K TF TG FRES CGAH F A KP Y K P
DL52 repli	IAPRVLEALS FYGFKPNQSK TFITGRFRES CGAHYFGGAD CKPIYIKKPV
DL54 repli	IAPRVLEALS FYGFKPNQSK TFITGRFRES CGAHYFGGAD CKPIYIKKPV
DL10 repli	CAPTLLKVLS AVNFLPNQKK TFFTGYFRES CGAHFFKGAD MKPFYCKRPM
DL20 repli	CAPTLLKVLS AVNFLPNQKK TFFTGYFRES CGAHFFKGAD MKPFYCKRPM
GA replica	CRPTLLKVLS AVNFLPNEEK TFFTGYFRES CGAHFFKDAD MKPFYCKRPM
T72 replic	CAPLLLQVLS AVNFLPNQKK TFFTGYFRES CGAHFFKGAS VKPFYCKRPM
KU1 replic	CAPLLLQVLS AVNFLPNQKK TFFTGYFRES CGAHFFKGAS VKPFYCKRPM

	410 420 430 440 450
Consensus	L LL NR RGW G SDPRLF WK P GG NLD
DL52 repli	NNLFAVCLLL NRLRGWGVVN GVSDPRLFET WKWLSERVPS ILFGGSNLDA
DL54 repli	NNLFAVCLLL NRLRGWGVVN GVSDPRLFET WKWLSERVPS ILFGGSNLDA
DL10 repli	ETLPDVMLLC NRIRGWQTVG GMSDPRLFPI WKEFADMIPP KFKGGCNLDR
DL20 repli	ETLPDVMLLC NRIRGWQTVG GMSDPRLFPI WKEFADMIPP KFKGGCNLDR
GA replica	ETLPDVMLLC NRIRGWQTVG GMSDPRLFPI WKEFADMIPP KFKGGCNLDR

T72 replic	ETLPDIMLLC	NRIRGWTIG	GISDPRLFPI	WKEFADMIPP	KFKGGCNLDR	
KU1 replic	ETLPDVLLLC	NRIRGWTIG	GISDPRLFPI	WKEFADMIPP	KFKGGCNLDR	
	
		460	470	480	490	500
Consensus	D Y VS		G L		F E	
DL52 repli	DYYVVSQRQA	KKRYESHPSY	GRTLSHFHST	PHRLYRVPVS	KREFSPREES	
DL54 repli	DYYVVSQRQA	RKRCEDHPVY	GRTLAHFHSS	PHRLYRVPAS	KREFSSREES	
DL10 repli	DTYLVSPDKP	-----	GVTLVRVAKV	RSGFN-----	-HSFPYGHEN	
DL20 repli	DTYLVSPDKP	-----	GVTLVRIAKV	RSGFN-----	-HAFPYGYEN	
GA replica	DTYLVSPDKP	-----	GVSLVRIAKV	RSGFN-----	-HAFPYGYEN	
T72 replic	DTYLVSPDKP	-----	GKTLVRVAKK	RSGFN-----	-HKFRSDYEN	
KU1 replic	DTYLVSPDKP	-----	GVTLVRDATV	RSGFN-----	-YKFRRRQEN	
	
		510	520	530	540	
Consensus	GR W H G	G T	R R	SE W	P FPQ E	LS
DL52 repli	GRLITWYHNG	-GQVIDTTTT	PRVRLVRTSE	WLTVVPLFPQ	EDGNCELS	
DL54 repli	GRLITWYHNG	-GQIIDTTTT	PRVRLVRTSE	WLAVVPSFPQ	EDDTRELS	
DL10 repli	GRYVHWLHMG	SGEVLETISS	ARFRCKPNSE	WRTQIPLFPQ	ELEACVLS	
DL20 repli	GRYVHWLHMG	SGEVLETISS	ARFRCKPNSE	WRTQIPLFPQ	ELEACVLS	
GA replica	GRYVHWLHMG	SGEVLETISS	ARYRCKPNSE	WRTQIPLFPQ	ELEACVLS	
T72 replic	GRYIHWLHMG	SGEVLETISS	ARFRCKPNSE	WRTQIPLFPQ	EIEACVLS	
KU1 replic	GRYIHWLHMG	SGEVSETISS	ARFRCKPNSE	WRIQIPLFPQ	EVEACVLS	

Appendix C4
Group I and JS Replicase Nucleotides

Alignment: Group I and JS replicase nucleotides.

	ORF4 START									
									
	10 20 30 40 50									
Consensus	ATGTCGAAGA	CAACAAAGAA	GTTCAACTCT	TTATGTATTG	ATCTTCCTYG					
DL1 IC.					
DL2 IC.					
DL13 IC.					
DL16 IT.					
ST4 IC.					
R17 IC.					
J20 IC.					
MS2 IC.					
DL52C.					
DL54C.					
									
	60 70 80 90 100									
Consensus	CGATCTTTCT	CTCGAAATTT	ACCAATCAAT	TGCTTCTGTC	GCTACTGGAA					
DL1 I					
DL2 I					
DL13 I					
DL16 I					
ST4 I					
R17 I					
J20 I					
MS2 I					
DL52					
DL54					
									
	110 120 130 140 150									
Consensus	GCGGTRATCC	GCACAGTRRM	GACTTTACRG	CAATTGCTTA	CYTAAGRGAY					
DL1 IG....AGAA.C....G..C					
DL2 IG....AGAA.C....A..C					
DL13 IG....AGAA.C....A..C					
DL16 IG....AGAA.C....A..C					
ST4 IG....GACA.T....G..C					
R17 IA....GACA.C....G..C					
J20 IG....AGAG.C....A..C					
MS2 IG....GACA.T....G..C					
DL52G....AGAA.C....A..C					
DL54G....AGAA.C....A..T					

	160170180190200
Consensus	GARTTGCTHA CWAAGCATCC VWCHTTRGGH WMTGGTAATG ACGAGGCGAC
DL1 I	..G.....A. .T..... CT.C..A..C AA.....
DL2 I	..A.....A. .T..... GT.C..A..A AA.....
DL13 I	..A.....A. .T..... GT.C..A..A AA.....
DL16 I	..A.....A. .T..... GT.C..A..A AA.....
ST4 I	..A.....C. .A..... GA.C..A..T TC.....
R17 I	..A.....T. .A..... GA.T..A..C TC.....
J20 I	..G.....C. .A..... GT.T..A..A AA.....
MS2 I	..A.....C. .A..... GA.C..A..T TC.....
DL52	..A.....A. .T..... GT.C..A..A AA.....
DL54	..G.....T. .T..... AT.A..G..C AA.....

	210220230240250
Consensus	CCGYCGNRCY YTAGCTATYG CTAAGCTNCG GGAGGCGAAT GRDSRDYGYG
DL1 I	...C..AG.T T.....T.T.. AACGAT.T.
DL2 I	...C..GG.T C.....C.C.. AGCGGT.C.
DL13 I	...C..GG.T C.....C.C.. AGCGGT.C.
DL16 I	...C..GG.T C.....C.C.. AGCGGT.C.
ST4 I	...T..TA.C T.....C.A.. ATCGGT.C.
R17 I	...T..CA.C T.....T.G.. ATCGGT.C.
J20 I	...C..GG.T T.....C.C.. AACGAT.T.
MS2 I	...T..TA.C T.....C.A.. GTGATC.C.
DL52	...C..GG.T C.....C.C.. AGCGGT.C.
DL54	...C..GG.T C.....T.C.. AGCGAT.T.

	260270280290300
Consensus	GYCAGATHAA YAGRGARGGT TTCTTACAYG AYAAATCCTT RTCRTGGGAT
DL1 I	.C.....C.. T..A..G... ..T.. .C..... G..A.....
DL2 I	.C.....T.. T..G..A... ..C.. .T..... A..G.....
DL13 I	.C.....T.. T..G..A... ..C.. .T..... A..G.....
DL16 I	.C.....T.. T..G..A... ..C.. .T..... A..G.....
ST4 I	.T.....A.. T..A..A... ..T.. .C..... G..A.....
R17 I	.C.....A.. T..A..A... ..T.. .C..... G..A.....
J20 I	.C.....T.. C..A..A... ..T.. .C..... G..G.....
MS2 I	.T.....A.. T..A..A... ..T.. .C..... G..A.....
DL52	.C.....T.. T..G..A... ..C.. .T..... A..G.....
DL54	.C.....T.. T..A..A... ..T.. .C..... G..A.....

	310320330340350
Consensus	CCGGATGTTT TACAAACCAG CATCCGTAGC CTTATYGGCA AYCTYCTCTC
DL1 IT.... .C..T.....
DL2 IT.... .C..T.....
DL13 IT.... .C..T.....
DL16 IT.... .C..T.....
ST4 IT.... .C..C.....
R17 IC.... .T..T.....
J20 IT.... .C..T.....
MS2 IT.... .C..C.....

DL52	T.....	.C..T.....
DL54	T.....	.C..C.....

	360 370 380 390 400
Consensus	TGGCTAYCRH TCGTCGTTGT TTGGGCAATG CACGTTYTCC AACGGTGCYY
DL1 IT.GTC... ..CT
DL2 IC.GCC... ..CT
DL13 IC.GCC... ..CT
DL16 IC.GCC... ..CT
ST4 IC.GAC... ..CT
R17 IC.AAC... ..CT
J20 IT.GTT... ..CT
MS2 IC.GAC... ..TC
DL52C.GCC... ..CT
DL54T.GTC... ..CT

	410 420 430 440 450
Consensus	CDATGGGGCA CAAGTTGCAG GATGCAGCGC CTTACAAGAA GTTCGCTGAA
DL1 I	.G..... ..
DL2 I	.A..... ..
DL13 I	.A..... ..
DL16 I	.A..... ..
ST4 I	.T..... ..
R17 I	.T..... ..
J20 I	.T..... ..
MS2 I	.T..... ..
DL52	.A..... ..
DL54	.G..... ..

	460 470 480 490 500
Consensus	CAAGCAACCG TTACCCCCCG CGCTCTRAGA GCGGCNYTAY TGGTCMGAGA
DL1 IA... ..CC..C ..C...
DL2 IG... ..AC..C ..C...
DL13 IG... ..AC..C ..C...
DL16 IG... ..AC..C ..C...
ST4 IG... ..TC..T ..C...
R17 IG... ..TC..T ..C...
J20 IG... ..GT..C ..A...
MS2 IG... ..TC..T ..C...
DL52G... ..AC..C ..C...
DL54G... ..TC..C ..C...

	510 520 530 540 550
Consensus	CCARTGTGYG CCGTGATYA GACACGCGGT CCRCTAYARY GARTCATATR
DL1 I	...G...T.T.G...C.AT ..G.....G
DL2 I	...G...T.C.G...C.AC ..G.....G
DL13 I	...G...T.C.G...C.AC ..G.....G
DL16 I	...G...T.C.G...C.AC ..G.....G
ST4 I	...A...C.C.G...T.AC ..G.....G
R17 I	...A...C.C.A...T.AC ..G.....G
J20 I	...G...C.C.G...T.AC ..A.....A
MS2 I	...A...C.C.G...T.AC ..G.....G

DL52	...G....T.C.G...C.AC	..G.....G
DL54	...G....C.C.A...C.GC	..G.....G

	560	570	580	590	600
Consensus	AATTTAGRCT	CGTYGTAGGG	AACGGAGTGT	TYACAGTTCC	GAAGAATAAT
DL1 IG..	...T.....T.....
DL2 IG..	...C.....T.....
DL13 IG..	...C.....T.....
DL16 IG..	...C.....T.....
ST4 IG..	...T.....T.....
R17 IG..	...T.....T.....
J20 IA..	...T.....T.....
MS2 IG..	...T.....T.....
DL52G..	...C.....T.....
DL54G..	...C.....C.....

	610	620	630	640	650
Consensus	AAAATAGATC	GGGCTGCCTG	YAAGGAGCCH	GATRYGAAYA	TGTACCTYCA
DL1 I	T.....C	...AT...T.T..
DL2 I	T.....C	...AT...T.C..
DL13 I	T.....C	...AT...T.C..
DL16 I	T.....C	...AT...T.C..
ST4 I	T.....T	...AT...T.C..
R17 I	T.....T	...AT...T.C..
J20 I	T.....C	...AT...T.T..
MS2 I	T.....T	...AT...T.C..
DL52	C.....T	...GC...C.T..
DL54	T.....A	...GC...C.T..

	660	670	680	690	700
Consensus	GAAAGGRGTY	GGYGVYTTYA	THMGVCGYCG	BCTCMRHTCY	RTYGGTATMG
DL1 IG..T	..C.CC..C.	.TA.A..C..	C...AAA..T	G.C.....A.
DL2 IA..T	..C.CC..C.	.AA.G..T..	C...AAA..T	G.C.....A.
DL13 IA..T	..C.CC..C.	.AA.G..T..	C...AAA..T	G.C.....A.
DL16 IA..T	..C.CC..C.	.AA.G..T..	C...AAA..T	G.C.....A.
ST4 IG..C	..T.CC..T.	.CA.A..C..	G...AAA..C	G.T.....A.
R17 IG..C	..C.CT..T.	.TA.A..C..	G...AAA..C	G.T.....A.
J20 IA..T	..C.CT..T.	.AC.G..T..	T...AGA..C	G.T.....A.
MS2 IG..C	..T.CT..C.	.CA.A..C..	G...AAA..C	G.T.....A.
DL52A..T	..C.AT..T.	.AC.C..T..	T...CGC..T	A.C.....C.
DL54G..T	..T.GC..C.	.AC.C..C..	C...CGT..T	A.C.....C.

	710	720	730	740	750
Consensus	AYCTGAATGA	TCARWCGATC	AAYCARCKTY	TDGCWMARCW	NGGCAGYRYM
DL1 I	.C.....	...AA.....	..T..G.G.C	.A..TC.A.A	G.....CATA
DL2 I	.C.....	...AA.....	..T..G.G.T	.A..TC.A.A	G.....CATA
DL13 I	.C.....	...AA.....	..T..G.G.T	.A..TC.A.A	G.....CATA
DL16 I	.C.....	...AA.....	..T..G.G.T	.A..TC.A.A	G.....CATA
ST4 I	.T.....	...AT.....	..C..G.T.C	.G..TC.G.A	G.....CGTA
R17 I	.C.....	...GT.....	..C..G.G.C	.A..TC.G.A	A.....CGCA

J20 I	.C.....	...AA.....	..C..G.G.C	.G..TC.A.A	A.....CGTC
MS2 I	.C.....	...AT.....	..C..G.G.C	.G..TC.G.A	G.....CGTA
DL52	.C.....	...AA.....	..T..G.G.C	.T..AA.A.T	T.....CATC
DL54	.C.....	...AA.....	..T..A.G.C	.T..AA.A.T	C.....TATC

	760770780790800
Consensus	GATGGYTCDY TDGCRACGAT WGAYYTATCG TCNGCNTCBG AYTCYATHTC
DL1 IT..TT .A..G..... A..CC..... ..A..T..G. .T..C..C..
DL2 IC..TT .A..A..... A..TT..... ..G..T..T. .T..T..C..
DL13 IC..TT .A..A..... A..TT..... ..G..T..T. .T..T..C..
DL16 IC..TT .A..A..... A..TT..... ..G..T..T. .T..T..C..
ST4 IT..GC .T..G..... A..CT..... ..T..A..C. .T..C..C..
R17 IT..GC .T..G..... A..CT..... ..C..G..C. .T..C..T..
J20 IC..AT .G..G..... A..CT..... ..G..C..T. .C..C..C..
MS2 IT..GC .T..G..... A..CT..... ..T..A..C. .T..C..C..
DL52T..TT .A..G..... T..TT..... ..T..T..T. .T..C..A..
DL54T..TT .A..G..... A..TT..... ..T..T..T. .C..C..A..

	810820830840850
Consensus	BGAYCGCCTN GTKTGGRRNT TYCTYCCAYC KSARHTRTAY KCRTAYCTBK
DL1 I	T..C.....G ..G...AGC. .T..C...C. TG.GC.A..T T.A..T..CG
DL2 I	T..C.....G ..G...AGT. .C..C...C. TG.GT.A..T T.A..T..CG
DL13 I	T..C.....G ..G...AGT. .C..C...C. TG.GT.A..T T.A..T..CG
DL16 I	T..C.....G ..G...AGT. .C..C...C. TG.GT.A..T T.A..T..CG
ST4 I	C..T.....G ..G...AGT. .T..C...C. TG.GC.A..T T.A..T..CG
R17 I	C..C.....A ..G...AAT. .C..T...C. TG.GC.A..T T.A..T..TG
J20 I	T..C.....G ..G...AGT. .C..C...C. TG.GC.A..T T.A..T..CG
MS2 I	C..T.....G ..G...AGT. .T..C...C. TG.GC.A..T T.A..T..CG
DL52	G..C.....C ..T...GAA. .C..C...T. GC.AA.G..C G.G..C..GT
DL54	G..C.....T ..T...GAG. .T..C...C. GC.AA.G..T G.G..C..GT

	860870880890900
Consensus	MKMRWATYCG YTCVYMMYRH GGAATCRTWG AYGGVSRKRY RRTMSRVTTGG
DL1 I	ATCGT..C.. C..CCACTACG.A. .T..CGAGAC GA.ACGG...
DL2 I	ATCGT..C.. C..CCACTACG.A. .T..GGAGAC GA.ACGA...
DL13 I	ATCGT..C.. C..CCACTACG.A. .T..GGAGAC GA.ACGA...
DL16 I	ATCGT..C.. C..CCACTACG.A. .T..GGAGAC GA.ACGA...
ST4 I	ATCGT..C.. C..ACACTACG.A. .T..CGAGAC GA.ACGA...
R17 I	ATCGT..C.. C..GCACTACG.A. .T..CGAGAC GA.ACGA...
J20 I	ATCGT..C.. C..CCACTATA.A. .T..AGAGAC GA.ACGA...
MS2 I	ATCGT..C.. C..ACACTACG.A. .T..CGAGAC GA.ACGA...
DL52	CGAAA..T.. T..GTCACGAG.T. .C..ACGTGT AG.CGAC...
DL54	CTAAA..T.. T..GCCACGAG.T. .C..ACGTAT GA.CGAC...

	910920930940950
Consensus	SAMCTATTTT CCACDATGGG WAATGGDDTTY ACNTTYGARC TAGAGTCCAT
DL1 I	G.A..... ..A..... A.....G..C ..T..C..A.
DL2 I	G.A..... ..A..... T.....G..C ..T..C..A.
DL13 I	G.A..... ..A..... T.....G..C ..T..C..A.
DL16 I	G.A..... ..A..... T.....G..C ..T..C..A.
ST4 I	G.A..... ..A..... A.....G..C ..G..T..G.
R17 I	G.A..... ..G..... A.....G..T ..A..T..G.
J20 I	G.A..... ..A..... A.....G..C ..T..T..A.
MS2 I	G.A..... ..A..... A.....G..C ..A..T..G.

DL52	C.C.....A.....	T.....T..T	..C..T..G.
DL54	C.C.....T.....	T.....A..T	..C..T..A.

	960 970 980 990 1000
Consensus	GATMTTYTGG GCWATAGTNA ARGCRACYMW RAYYCATT TT GGTAACSYYG
DL1 I	...A..T... ..T.....G. .A..G..CCA G.TC..... ..GCC.
DL2 I	...C..T... ..T.....G. .A..A..CCA G.TC..... ..GCC.
DL13 I	...C..T... ..T.....G. .A..A..CCA G.TC..... ..GCC.
DL16 I	...C..T... ..T.....G. .A..A..CCA G.TC..... ..GCC.
ST4 I	...A..C... ..A.....C. .A..G..CCA A.TC..... ..GCC.
R17 I	...A..C... ..A.....C. .A..A..CCA A.TC..... ..GCC.
J20 I	...A..T... ..T.....A. .A..A..CCA G.TC..... ..GCC.
MS2 I	...A..C... ..A.....C. .A..G..CCA A.TC..... ..GCC.
DL52	...A..T... ..A.....C. .G..A..TAT G.TC..... ..CTT.
DL54	...C..T... ..A.....T. .G..A..CAT G.CT..... ..CTT.

	1010 1020 1030 1040 1050
Consensus	GAACMATWGG CATCTAYGGG GACGATATYA TATGTCCCAS WGAGATTGCA
DL1 IC..A..C...T.G T.....
DL2 IC..A..C...T.G T.....
DL13 IC..A..C...T.G T.....
DL16 IC..A..C...T.G T.....
ST4 IC..A..C...T.G T.....
R17 IC..A..C...C.G T.....
J20 IC..A..C...T.G T.....
MS2 IC..A..C...T.G T.....
DL52A..T..T...C.C A.....
DL54A..T..T...C.C A.....

	1060 1070 1080 1090 1100
Consensus	CCYCGTGTGC TRGAGGCDCT HRSCTWCTAC GGTTTYAAAC CGAATCWBY S
DL1 I	..C..... .A.....A. TGC..A....T.... ..TTCG
DL2 I	..C..... .A.....A. TGC..A....C.... ..TTCG
DL13 I	..C..... .A.....A. TGC..A....C.... ..TTCG
DL16 I	..C..... .A.....A. TGC..A....C.... ..TTCG
ST4 I	..C..... .G.....A. TGC..A....C.... ..TCCG
R17 I	..C..... .A.....A. TGC..A....T.... ..TTCG
J20 I	..C..... .A.....G. AGC..A....C.... ..TTCG
MS2 I	..C..... .A.....A. TGC..A....T.... ..TTCG
DL52	..T..... .A.....T. CAG..T....T.... ..AGTC
DL54	..T..... .A.....T. CAG..T....T.... ..AGTC

	1110 1120 1130 1140 1150
Consensus	KAARACGTTC RTSWCVGK C KCTTTCGCGA GWSCTGYRGY GCRCAYTWYT
DL1 I	T..A..... G.GT.G..G. T..... ..AG...CG.C ..G..C.TT.
DL2 I	T..A..... G.GT.A..G. T..... ..AG...TG.C ..G..C.TT.
DL13 I	T..A..... G.GT.A..G. T..... ..AG...TG.C ..G..C.TT.
DL16 I	T..A..... G.GT.A..G. T..... ..AG...TG.C ..G..C.TT.
ST4 I	T..A..... G.GT.C..G. T..... ..AG...CG.C ..G..C.TT.
R17 I	T..A..... G.GT.C..G. T..... ..AG...CA.C ..G..C.TT.
J20 I	T..A..... G.GT.A..G. T..... ..AG...TG.C ..G..C.TT.
MS2 I	T..A..... G.GT.C..G. T..... ..AG...CG.C ..G..C.TT.

DL52	G..G.....	A.CA.G..T.	G.....	.TC...TG.T	..A..T.AC.
DL54	G..G.....	A.CA.G..T.	G.....	.TC...TG.T	..A..T.AC.

	1160	1170	1180	1190	1200
Consensus	WCSGYGGTGY	YGATKKCAAA	CCGWTYTAYA	TCARGAAACC	HGTYRACAAY
DL1 I	A.C.T....T	C...GT....	...T.T..C.	...A.....	A..TG....C
DL2 I	A.C.T....T	C...GT....	...T.C..C.	...A.....	A..TG....T
DL13 I	A.C.T....T	C...GT....	...T.C..C.	...A.....	A..TG....T
DL16 I	A.C.T....T	C...GT....	...T.C..C.	...A.....	A..TG....T
ST4 I	A.C.T....T	C...GT....	...T.T..C.	...A.....	T..TG....T
R17 I	A.C.T....T	C...GT....	...T.T..C.	...A.....	T..TG....C
J20 I	A.C.T....T	C...GT....	...T.T..C.	...G.....	A..CG....T
MS2 I	A.C.T....T	C...GT....	...T.T..C.	...A.....	T..TG....T
DL52	T.G.C....C	T...TG....	...A.T..T.	...A.....	C..TA....C
DL54	T.G.C....C	T...TG....	...A.T..T.	...A.....	C..TA....C

	1210	1220	1230	1240	1250
Consensus	CTCTTYKCCS	TBWKKCTGWT	MHTNAA YMGR	CTDMGSGGNT	GGGGNGTTGT
DL1 ICT..C	.TATG...A.	AA.G..TC.G	..TA.G..G.C.....
DL2 ICT..C	.TATG...A.	CA.G..TC.G	..TA.G..G.C.....
DL13 ICT..C	.TATG...A.	CA.G..TC.G	..TA.G..G.C.....
DL16 ICT..C	.TATG...A.	CA.G..TC.G	..TA.G..G.C.....
ST4 ICG..C	.TATG...A.	AT.G..TC.G	..AC.G..T.A.....
R17 ITG..C	.GATG...A.	AT.A..TC.G	..AC.G..T.G.....
J20 ICT..C	.TATG...A.	CA.G..TC.G	..TA.G..A.T.....
MS2 ICG..C	.GATG...A.	AT.A..TC.G	..AC.G..T.A.....
DL52CG..G	.CTGT...T.	AC.C..CA.G	..AC.C..G.T.....
DL54CG..G	.CTGT...T.	AC.T..CA.A	..GC.C..C.T.....

	1260	1270	1280	1290	1300
Consensus	SRRMGGWRTG	TCAGATCCDC	GCCTHTWYRA	GRYKTGGRWA	YGRCTVTCK
DL1 I	CGGA..TA..G.T.ATA.	.GTT...GT.	C.A..G...T
DL2 I	CGGA..TA..G.T.ATA.	.GTT...GT.	C.A..A...T
DL13 I	CGGA..TA..G.T.ATA.	.GTT...GT.	C.A..A...T
DL16 I	CGGA..TA..G.T.ATA.	.GTT...GT.	C.A..A...T
ST4 I	CGGA..TA..A.T.ACA.	.GTG...GT.	C.A..C...T
R17 I	CGGA..TA..A.T.ACA.	.GTG...GT.	C.A..C...T
J20 I	CGGA..TA..T.A.ATA.	.GTT...GT.	C.A..G...T
MS2 I	CGGA..TA..A.C.ATA.	.GTG...GT.	C.G..C...T
DL52	GAAC..AG..T.C.TCG.	.ACT...AA.	T.G..A...G
DL54	GAAC..TG..T.C.TCG.	.ACT...AA.	T.G..A...G

	1310	1320	1330	1340	1350
Consensus	MMCDKGTDC	HTCGATRYTY	TTCGGTGGGW	CDRACCTYGM	TGCHGACTAY
DL1 I	CC.TG..T..	A.....GT.CA	.GG....T.C	...C.....T
DL2 I	CC.TG..T..	A.....GT.CA	.GG....T.C	...C.....C
DL13 I	CC.TG..T..	A.....GT.CA	.GG....T.C	...C.....C
DL16 I	CC.TG..T..	A.....GT.CA	.GG....T.C	...C.....C
ST4 I	CC.AG..G..	T.....GT.TA	.GG....C.C	...C.....C
R17 I	CC.AG..A..	T.....GT.CA	.GG....C.C	...C.....C

J20 I	CC.TG..T.. A.....GT.C	A .GG....T.C ...T.....C
MS2 I	CC.AG..G.. T.....GT.C	A .GG....C.C ...C.....C
DL52	AA.GT..T.. T.....AC.T	T .TA....T.A ...A.....C
DL54	AA.GT..T.. C.....AC.T	T .AA....T.A ...A.....T

		First nt insertion									
		
		1360	1370	1380	1390	1400					
Consensus		TACGTRGTYA	GYCSBCMBRM	NGCYADRWMK	CRKTDYGABR	VYMANMCHBY					
DL1 I	A..C.	.C.CC.CGAC	C.--AGTCT	.AG.TT-.TA	CTA.AA.TGC					
DL2 I	A..T.	.C.CT.CGAA	T.--TGTCT	.GG.TT-.TA	CCA.AA.TGC					
DL13 I	A..T.	.C.CT.CGAA	T.--TGTCT	.GG.TT-.TA	CCA.AA.TGC					
DL16 I	A..T.	.C.CT.CGAA	T.--TGTCT	.GG.TT-.TA	CCA.AA.TGC					
ST4 I	A..C.	.C.CG.CCAC	G.--AGTCT	.GG.AT-.TA	CCA.GA.TCC					
R17 I	A..C.	.C.CG.CTAC	G.--GGTCT	.GG.AT-.TA	CCA.GA.TCC					
J20 I	A..C.	.C.CC.CGAC	A.--GGTCT	.AG.TT-.TA	CTA.GA.CGC					
MS2 I	A..C.	.C.CG.CTAC	G.--AGTCT	.GG.AT-.CA	CCA.GA.TCC					
DL52	G..T.	.T.GT.AGGA	A..T.AAAAG	.GT.AC..GA	GTC.CC.ATC					
DL54	G..C.	.C.GC.AGGA	A..C.GGAAG	.GT.GC..GG	ATC.TC.AGT					
		
		1410	1420	1430	1440	1450					
Consensus		NTAYGGRMGB	HYVCTYKCKS	AYDYCRYAS	CWCKSSTYWY	CGKYTKKMYC					
DL1 I		C..C..GA.G	TTA..CG.GG	.TACC.GT.C	.T.GGG.TTC	..TC.TGCT.					
DL2 I		A..T..GA.G	TTA..CG.GG	.CGCC.GT.C	.T.GGG.TTC	..TC.TGCT.					
DL13 I		A..T..GA.G	TTA..CG.GG	.CGCC.GT.C	.T.GGG.TTC	..TC.TGCT.					
DL16 I		A..T..GA.G	TTA..CG.GG	.CGCC.GT.C	.T.GGG.TTC	..TC.TGCT.					
ST4 I		G..T..GC.G	CTA..CG.GG	.TACC.GT.C	.T.GGG.TTC	..TC.TGCT.					
R17 I		G..T..AC.G	CTG..CG.GG	.TACC.GT.C	.T.GGG.TTC	..TC.TGCT.					
J20 I		A..T..GA.G	TTG..CG.GG	.CGCC.GT.C	.T.GGG.TTC	..TC.TGCT.					
MS2 I		G..C..GC.G	CTG..CG.GG	.TACC.GT.C	.T.GGG.TTC	..TC.TGCT.					
DL52		T..T..GC.C	ACC..CT.TC	.CTTT.AT.G	.A.TCC.CAT	..GC.TTAC.					
DL54		A..T..AC.T	ACC..TG.TC	.TTTC.AC.G	.T.TCC.CAT	..GT.GTAC.					
		
		1460	1470	1480	1490	1500					
Consensus		GYRYRCCCGY	NHVWRARCGH	RARYDYTCTY	MRCGHRAAGM	RYSRCAGYGG					
DL1 I		.TAT---.C	AAAAG.G..A	A.GCGC.-.T	AG..AG...C	ATGA...T..					
DL2 I		.TAT---.C	GAAAG.G..A	A.GCAC.-.T	AG..AG...C	ATGA...T..					
DL13 I		.TAT---.C	GAAAG.G..A	A.GCAC.-.T	AG..AG...C	ATGG...T..					
DL16 I		.TAC---.C	GAAAG.G..A	A.GCAC.-.T	AG..AG...C	ATGA...T..					
ST4 I		.TAT---.C	TCGAG.A..C	A.GTTC.-.C	AG..AA...C	ATGA...T..					
R17 I		.TAT---.C	TCGAG.A..C	A.GTTC.-.C	AG..AA...C	ATGA...T..					
J20 I		.TAT---.C	GAAAG.G..T	A.GCAC.-.T	AG..AG...C	ATGA...T..					
MS2 I		.TAT---.C	TCGAG.A..C	A.GTTC.-.C	AG..AA...C	ACGA...T..					
DL52		.CGTG....T	ATCTA.A..T	G.ATTT...C	CA..TG...A	ATC---C..					
DL54		.CGTA....C	CTCTA.A..C	G.ATTT...T	CG..CG...A	GTC---C..					
		
		1510	1520	1530	1540	1550					
Consensus		YCGCYWCATM	RCDTGGTWCC	ATAMTGGHGG	TSARRTYAAY	GAYASYAYKA					
DL1 I		T...TA...A	G.A....T..	...C...A..	.G.GA.C.CC	..T.GT.TG.					
DL2 I		T...TA...A	G.A....T..	...C...A..	.G.AA.C.CC	..T.GT.TG.					
DL13 I		T...TA...A	G.A....T..	...C...A..	.G.AA.C.CC	..T.GT.TG.					
DL16 I		T...TA...A	G.A....T..	...C...A..	.G.AA.C.CC	..T.GT.TG.					
ST4 I		C...TA...A	G.G....T..	...C...A..	.G.AG.C.CC	..C.GT.TG.					
R17 I		T...TA...A	G.G....T..	...C...A..	.G.AA.C.CC	..C.GT.TG.					
J20 I		T...TA...A	G.A....T..	...C...A..	.G.AA.C.CC	..T.GT.TG.					
MS2 I		T...TA...A	G.G....T..	...C...A..	.G.AA.C.CC	..C.GC.TG.					
DL52		T...CT...C	A.T....A..	...A...T..	.C.AG.T.TT	..C.CC.CT.					

DL54

T...CT...C A.T....A... ..A...C... .C.AA.T.TT ..C.CC.CT.

	15601570158015901600
Consensus	MGWCCSCHRG SGTCKGYVTH RTRCGMACDT CGGARTGGCT RRCRSYRGTK
DL1 I	A.T..G.TG. C..G..CG.A A.G..C..T.G..... GA.GCCG..T
DL2 I	A.T..G.TG. C..G..TG.A A.A..C..T.G..... GA.GCCG..T
DL13 I	A.T..G.TG. C..G..TG.A A.A..C..T.G..... GA.GCCG..T
DL16 I	A.T..G.TG. C..G..TG.A A.A..C..T.G..... GA.GCCG..T
ST4 I	A.T..G.CG. C..G..TA.T A.G..C..T.G..... AA.GCCG..T
R17 I	A.T..G.CG. C..G..CA.C A.G..C..T.G..... AA.GCCG..T
J20 I	A.T..G.CG. C..G..TG.T A.G..C..T.A..... AA.GCCG..T
MS2 I	A.T..G.CG. C..G..CG.T A.A..C..T.G..... AA.GCCG..T
DL52	C.A..C.AA. G..T..CC.T G.G..A..G.A..... AA.AGTG..G
DL54	C.A..C.AA. G..T..CC.T G.G..A..A.A..... AG.GGTA..G

		(Group I) ORF4 STOP	
		
	16101620163016401650		
Consensus	CCMHYMTTCC CKCAGGARKR TGRCRMHGC GAGCTCTCCT MGK YAG CWSR		
DL1 I	..CACA.... .T.....GTG ..G-GCCA.. C.G T ...TGA		
DL2 I	..CATA.... .T.....ATG ..G-GCCA.. C.G T ...TGA		
DL13 I	..CATA.... .T.....ATG ..G-GCCA.. C.G T ...TGA		
DL16 I	..CATA.... .T.....ATG ..G-GCCA.. C.G T ...TGA		
ST4 I	..CACA.... .T.....GTG ..G-GCCA.. C.G T ...TGA		
R17 I	..CACA.... .T.....GTG ..G-GCCA.. C.G T ...TGA		
J20 I	..CACA.... .G.....GTG ..G-GCCA.. C.G T ...TGA		
MS2 I	..CACA.... .T.....GTG ..G-GCCA.. C.G T ...TGA		
DL52	..ACTC.... .T.....AGA ..G.AACT.. A.TC...ACG		
DL54	..ATCC.... .T.....AGA ..A.ACAC.. A.TC...ACG		

		ORF4 STOP (JS Group)	
		
	16601670168016901700		
Consensus	SCKAGG GACC CCCGTAAWCG GGGTGGGTGT GCTCGAAAGA GCACGGGTCC		
DL1 I	C.G..... ..A..		
DL2 I	C.G..... ..A..		
DL13 I	C.G..... ..A..		
DL16 I	C.G..... ..A..		
ST4 I	C.G..... ..A..		
R17 I	C.G..... ..A..		
J20 I	C.G..... ..A..		
MS2 I	C.G..... ..A..		
DL52	G. TT.. -----		
DL54	G. TT.. -----		

	17101720173017401750
Consensus	GCGWAAGCGG YCCGKMTYCA YCGWAAGGTG GGCGGGCTTC GGCCCARSGA
DL1 I	...A..... T--.GC.C.. C..A.....GG..
DL2 I	...A..... T--.GC.C.. C..A.....GG..
DL13 I	...A..... T--.GC.C.. C..A.....GG..
DL16 I	...A..... T--.GC.C.. C..A.....GG..
ST4 I	...A..... T--.GC.C.. C..A.....GG..
R17 I	...A..... T--.GC.C.. C..A.....GG..
J20 I	...T..... T--.GC.C.. C..A.....GG..
MS2 I	...A..... T...GC.C.. C..A.....GG..
DL52	...A..... C---TA.T.. T..T..---- - - - - - .AC..

DL54 ...A..... C---TA.T.. T..T..---- ----- ----..AC..

...A..... C---TA.T.. T..T..---- ----- ----..AC..

	1760 1770 1780 1790 1800
Consensus	YCTCCCCYTR ARGAKAGGRC CCGGKAWYCT CCYRATTYGG TRACTARCTT
DL1 I	C.....C.A .A..G...A.G.TT.. ..CG...T.. .A....G..-
DL2 I	C.....C.A .A..G...A.G.TT.. ..CG...T.. .A....G..-
DL13 I	C.....C.A .A..G...A.G.TT.. ..CG...T.. .A....G..-
DL16 I	C.....C.A .A..G...A.G.TT.. ..CG...T.. .A....G..-
ST4 I	C.....T.G .A..G...G.G.TT.. ..CG...T.. .A....G..-
R17 I	C.....C.G .A..G...A.G.TT.. ..CG...T.. .A....G..-
J20 I	C.....C.A .A..G...A.G.TT.. ..CG...T.. .A....G..-
MS2 I	C.....C.A .A..G...A.G.TT.. ..CG...T.. .A....G..-
DL52	T----- -G..T...A- ---.T.AT.. ..TA...T.. .G....A...
DL54	T----- -G..T...A- ---.T.AC.. ..TA...C.. .G....A...

	1810 1820
Consensus	TGCTTGGCTA GTBACCACCC A
DL1 I	-..... ..T..... .
DL2 I	-..... ..T..... .
DL13 I	-..... ..T..... .
DL16 I	-..... ..T..... .
ST4 I	-..... ..T..... .
R17 I	-..... ..G..... .
J20 I	-..... ..T..... .
MS2 I	-..... ..T..... .
DL52C..... .
DL54C..... .